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Cells Expressing Murine RAD52 Splice Variants Favor Sister Chromatid Repair†•

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The RAD52 gene is essential for homologous recombination in the yeast Saccharomyces cerevisiae. RAD52 is the archetype in an epistasis group of genes essential for DNA damage repair. By catalyzing the replacement of replication protein A with Rad51 on single-stranded DNA, Rad52 likely promotes strand invasion of a double-stranded DNA molecule by single-stranded DNA. Although the sequence and in vitro functions of mammalian RAD52 are conserved with those of yeast, one difference is the presence of introns and consequent splicing of the mammalian RAD52 pre-mRNA. We identified two novel splice variants from the RAD52 gene that are expressed in adult mouse tissues. Expression of these splice variants in tissue culture cells elevates the frequency of recombination that uses a sister chromatid template. To characterize this dominant phenotype further, the RAD52 gene from the yeast Saccharomyces cerevisiae was truncated to model the mammalian splice variants. The same dominant sister chromatid recombination phenotype seen in mammalian cells was also observed in yeast. Furthermore, repair from a homologous chromatid is reduced in yeast, implying that the choice of alternative repair pathways may be controlled by these variants. In addition, a dominant DNA repair defect induced by one of the variants in yeast is suppressed by overexpression of RAD51, suggesting that the Rad51-Rad52 interaction is impaired.

Homology-directed repair (HDR) allows cells to repair DNA double-strand breaks (DSBs) in an error-free way and is a key mechanism for maintaining genome integrity in all eukaryotes (11, 48, 52). RAD52 is an essential gene for HDR in the yeast Saccharomyces cerevisiae (20, 30, 46). Identified as one member of an epistasis group of radiation-sensitive yeast mutants, rad52 strains are defective in HDR, the major DSB repair pathway in yeast. RAD52 is largely conserved throughout eukaryotes, and both the human and yeast Rad52 proteins stimulate the replacement of replication protein A with Rad51 on single-stranded DNA (4, 29, 39, 45). Additionally, Rad52 promotes DNA annealing and strand exchange between a DNA duplex and complementary single-stranded DNA (14, 21, 27). Three-dimensional structures of the N terminus of human RAD52 (hRAD52) reveal a multimeric ring containing a positively charged surface groove that has been suggested to bind single-stranded DNA (15, 40). Relative to the N terminus of Rad52, the C terminus is poorly conserved between homologs. The C terminus of Rad52 is essential for its interaction with Rad51, a bacterial RecA homolog (26, 37). Consistent with its ability to bind both itself and other DNA repair pro-
more, for one variant in yeast, the defect in DNA repair is suppressed by overexpression of RAD51. These data show that the splice variants have the potential to alter the ratio of repair from the homologous to the sister chromosome, possibly by impairing the interaction of Rad52 with Rad51.

MATERIALS AND METHODS

RNA isolation and reverse transcription-PCR (RT-PCR). Whole-tissue samples were homogenized first by passing approximately 15 mg tissue several times through a 27-gauge needle in RNeAlter buffer (Ambion) and then through a Qiashredder column (QIAGEN). RNA was purified from the resulting suspension without delay using the RNAeasy column purification system (QIAGEN). Three to 5 μg of RNA was used for cDNA synthesis using random hexamer primers, thus ensuring that the cDNA represents all the transcripts present in the tissue. Either the Superscript first-strand synthesis system for RT-PCR (Invitrogen) or first-strand cDNA synthesis kit for RT-PCR (Roche) was used for this cDNA synthesis. Approximately 10% of the resulting cDNA was used as a template for PCR using gene specific primers.

As a control for cDNA yield, primers were designed to amplify a 200-bp product spanning exon 4 and 5 from a splice transcript of the mouse glyceraldehyde-3-phosphate dehydrogenase gene (GAPDH). Each RT reaction includes a matched control that omits the reverse transcriptase enzyme to confirm that the subsequent PCR products are derived from cDNA and not from DNA contamination. GAPDH primers used for PCR were 5′-TGGAGAGCAGAACGTCTCA-3′ and 5′-GTTGCAAGATGTTGCTGA-3′. Rad52 exon 1 to 7 primers were 5′-TGAGGGCATCTGTTCCAGAAGCTTCC-3′ and 5′-TGAGGGCATCTGTTCCAGAAGCTTCC-3′. Rad52 exons 6 to 11 primers were 5′-GGAAAGGGCTCTGACTGATG-3′ and 5′-GGATTCTGCTCTCAACAGGGG-3′ (all oligonucleotides were obtained from Sigma-Genosys). All PCR amplifications used the following protocol: 95°C for 5 min, followed by 30 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 1 min. The resulting PCR products were separated on Tris-borate-EDTA (TBE)-agarose gel electrophoresis, and the gels were blotted overnight at 48°C. Blots were washed several times in 4× SSC (0.15 M NaCl plus 0.015 M sodium citrate) and then incubated with trypsin-EDTA (Invitrogen) and then washed once in PBS. Cells (2 × 10⁶) were plated on 60-mm tissue culture dishes and grown overnight in serum-plus medium. The following day the cells were transfected in OPTIMEM medium (Invitrogen) with 4 μg/dish plasmid DNA (2 μg of pSel-C and 2 μg of experimental plasmid as indicated), using 4 μl/dish lipofectin (Invitrogen) following the manufacturer’s instructions (this DNA/lipofectin ratio was determined to be optimal for transfection of CHO cells using the pIRES-GFP reporter vector). Transfection was carried out for 5 h, after which the medium was replaced with serum-plus medium. After 48 h, the cells were collected using trypsin-EDTA, washed twice in PBS, and resuspended in 200 to 600 μl PBS. FACs analysis was performed on the cells as previously described (49). Expression levels of I-SceI from the pSel vector were assumed to be equivalent in each case. The expression of the mammalian variants was assessed by a parallel experiment in which CHO cells were transfected with plasmids containing the pIRES vector and those described above but including the internal ribosomal entry site (IRES)-GFP sequence. This GFP sequence produces fluorescence signal in all cells that successfully take up and express the plasmid vector. FACs analysis of the resulting transfections allowed the measurement of the proportion of CHO cells transfected (GFP+). For each of the four vectors (pIRES-GFP, pRAD52-IRES-GFP, pRAD52-IRES-GFP, and pRAD52+intb5-IRES-GFP) the number of cells transfected was equivalent (Fig. S8 in the supplemental material).

Mammalian cell synchronization. To obtain a synchronous culture of mouse ST3 cells, the cells were grown to near confluence in 150-cm² tissue culture flasks and then arrested with 400 ng/ml (1.3 μM) nocodazole for 6 h. The mitotic cells were then collected by mitotic shake-off, which involves mechanical shaking of the flask for 5 min. The mitotic cells were collected by centrifugation, washed twice in nocodazole-free media, and then grown in nocodazole-free media. Cells were plated in 60-cm² dishes and harvested at various time points using trypsin. One aliquot was fixed in 100% ethanol at −20°C for at least 30 min. These fixed cells were then stained in PBS with 50 μg/ml propidium iodide and 200 μg/ml RNase A (Sigma) at 4°C for at least 30 min prior to FACs analysis. A second aliquot was placed directly into RLT buffer (QIAGEN) and stored at −80°C for RNA preparation as described above.

Yeast strains and methods. Standard genetic methods were used for the growth and manipulation of yeast (36). The yeast strains used in this study are listed in Table 1. The strains encoding Rad52 tagged with yellow fluorescent protein (YFP) and cyan fluorescent protein (CFP) have been reported previously (23). We modified W3749-14C (Rad52-YFP) using our previously published allele replacement method (8). Briefly, PCR primers were designed to generate targeting fragments that eliminate either the 3′ 1,281 bp of RAD52 for rad52Δ77-YFP or the 3′ 660 bp of RAD52 for rad52Δ2284. The final five amino acids of yeast Rad52 encoded by rad52Δ2277 and rad52Δ2284 are -FGTSR and -DTSHK, respectively. These two resulting Rad52 deletion strains (WS5001-9C and WS5002-5A) maintain the truncated yeast RAD52 ORF in frame with the downstream YFP gene. The sensitivity of yeast strains to γ-irradiation was determined by growing yeast strains inYPD to log phase and then plating on YPD plates and incubating them at 30°C. The survival rates of the mutants were determined by measuring the proportion of S/G 2-phase cells in log-phase culture containing rad52Δ77-YFP and -CFP foci were determined using fluorescence microscopy as previously described (23, 25). The fluorescence of tagged Rad52 protein in individual cells was measured by reconstructing cells in three dimensions from fluorescent images.
TABLE 1. Yeast strains used in this study

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</table>

* All strains in this study are RADS derivatives of W303 (47, 54). In addition to the genotype listed above, all strains are MATA/MATA his3A1/Y281 trp1-1/1TRP1 and homozygous for ade2-2 alleles. As the ade2 allele contains a fill-in mutation of the 5' Ndel restriction endonuclease cleavage site of the yeast ade2 gene, and the ade2-a allele contains a fill-in mutation of the 5' Ndel restriction endonuclease cleavage site (2). The frequency of heteroallelic recombination was measured for at least nine independent isolates of each diploid strain. Cells were grown overnight in synthetic complete (SC) medium and washed once in H2O before being briefly sonicated. Serial 10-fold dilutions were plated both on SC medium and in duplicate on adenine-deficient medium. Colonies were counted after 48 h, and the method of Lea and Coulson was used to estimate "m," the mean number of recombinants per culture (22).

The strains encoding the two heteroalleles of ADE2 were a kind gift from Lorraine Symington, and their use has been described elsewhere (12). The ade2-a allele contains a fill-in mutation of the 3' Ndel restriction endonuclease cleavage site of the yeast ADE2 gene, and the ade2-a allele contains a fill-in mutation of the 5' Ndel restriction endonuclease cleavage site (2). The frequency of heteroallelic recombination was measured for at least nine independent isolates of each diploid strain. Cells were grown overnight in synthetic complete (SC) medium and washed once in H2O before being briefly sonicated. Serial 10-fold dilutions were plated both on SC medium and in duplicate on adenine-deficient medium. Colonies were counted after 48 h, and the method of Lea and Coulson was used to estimate "m," the mean number of recombinants per culture (22).

The strains used for the direct-repeat assay contain two defective LEU2 alleles separated by a URA4 gene and were derived from previously published strain W1470 (41). Strains for direct-repeat recombination were treated in the same way as those for heteroallelic recombination, except that the cells were plated on SC and, in duplicate, on leucine-deficient medium. This direct-repeat recombination assay also allows us to measure the frequency of single-strand annealing (SSA) events, which are scored by the loss of a URA4 marker (pop-out), which is placed between the two repeated leu2 alleles. As the ade2 direct repeat is present as a diploid, LEU2 cells resulting from SSA events will be ura3Δ/URA3, compared to URA3/URA3 for replacement events. Plating the LEU2+ colonies on medium containing 5-fluoroorotic acid (5-FOA), which selects against URA3, allowed us to differentiate the URA3 genotype. The colonies that were ura3Δ/URA3, when replica-plated to 5-FOA, produced small papillae, indicative of their ability to lose the single URA3+ gene. In contrast, the URA3+/URA3- colonies failed to grow at all on 5-FOA. Sporulating a selection of each class of LEU2+ colonies and testing the ability of the haploid spores to grow on uracil-deficient medium confirmed their URA3 genotypes (data not shown). This allowed us to estimate the proportion of LEU+ recombinants resulting in pop-out events compared to replacement events (illustrated in Fig. 6A).

RESULTS

Identification of mouse splice variants. Preliminary data derived from our sequencing of the mouse RAD52 gene (unpublished observations) and from RT-PCR from an array of different mouse tissues (see Fig. SA in the supplemental material) suggested that at least two splice variants of mouse RAD52 are commonly expressed. To investigate these findings further, tissue samples of lung and liver were dissected from two C57BL/6J mice and used for RT-PCR. A series of overlapping PCRs were performed covering the length of the mouse RAD52 cDNA, as illustrated in Fig. 1A. The murine RAD52 gene contains 12 exons. Three PCRs amplify cDNA from exons 1 to 7, 6 to 11, and 10 to 12. These PCRs were designed to detect most alterations in splicing that are pre-
dicted from earlier studies (Fig. 5A), such as the removal of exon sequence or the inclusion of intron sequences. Gels were blotted and probed with radiolabeled oligonucleotides to ensure that the RT-PCR products are specific for the RAD52 gene. Using a relatively low number of PCR cycles and subsequent gel blotting also allows a semiquantitative assessment of the template RNA, as the RT-PCRs have not saturated.

We observed two variant products consistent with alternative splicing in addition to the expected RAD52 transcript products corresponding to the annotated RAD52 sequence (Fig. 1B and C). The expected product from exons 1 to 7 is 591 bp; however, a smaller, ~500-bp product was also identified (Fig. 1B). As depicted in Fig. 1C, products amplified from exons 6 to 11 give rise to a larger product of 750 to 800 bp in addition to the expected 631-bp product. No alternatively spliced products were observed from exons 10 to 12 (data not shown). Figure 1D shows the GAPDH control.

Both of these variant products are predicted to encode truncated RAD52 peptides (Fig. 2). DNA sequencing of the novel products revealed the alternative splicing. The small-variant product from exons 4 corresponds to a deletion of amino acids 1 to 248 of RAD52, and 42 “novel” amino acids are encoded by intron 8 (underlined and italicized).

Splice variants are associated with polyribosomes. The RAD52 splice variants are predicted to encode C-terminal truncations of the RAD52 protein. Since degradation products of normal RAD52 could confound the results of a protein blot and specific antibodies to the N terminus of murine RAD52 are not available, we had to rely on additional evidence to determine if the splice variants are being translated. In particular, we searched for these splice-variant mRNAs on polyribosomes by identifying the variants (described above) to screen for RAD52Δexon4 and RAD52+intron8. The normal RAD52 transcript and both of the splice variants are associated with polyribosomes (Fig. 3), indicating that the normal and variant RAD52 transcripts are being actively translated in these mouse cells.

Splice variants are expressed uniformly throughout the cell cycle. Since Rad52 is thought to act primarily in S and G2 and to be inactive in G1, it is of interest to assay the expression of the splice variants throughout the cell cycle. Therefore, the expression of the variants in synchronized mouse 3T3 cells was examined as described in Materials and Methods. For each time point, an aliquot was stained with propidium iodide and used to analyze DNA content by FACS analysis, and a second aliquot was used for RNA preparation. RT-PCR was performed to determine if the splice variants are being translated in these mouse cells.

FIG. 2. RAD52 peptide is aligned with the predicted product of the two variant transcripts, RAD52Δexon4 and RAD52+intron8. The RAD52Δexon4 sequence includes the first 63 amino acids of RAD52 followed by 18 “out of frame” amino acids (underlined and italicized), encoded by exon 5. The RAD52+intron8 sequence includes the first 248 amino acids of RAD52, followed by 42 amino acids encoded by intron 8 (underlined and italicized).

FIG. 3. Two autoradiographs are shown of blots of RT-PCRs specific for RAD52Δexon4 (primers amplify exons 1 to 7, top) and RAD52+intron8 (primers amplify exons 6 to 11, bottom). Each blot contains 10 lanes, representing the 10 fractions from the sucrose gradient, 1 to 10 in the order they came off the gradient. The lane to the right (1) represents products obtained from the bottom of the gradient; this may include some residual nuclei and high-molecular-mass complexes. The lanes to the left (8 to 10) contain low-molecular-mass complexes including free RNA molecules. Lanes 2 to 7 represent samples obtained from the polyribosome-containing fraction of the gradient. The UV absorbance profile of the polyribosome gradient is shown separately in Fig. SC of the supplemental material.
and D) for the RT-PCR products used to identify products result: replacements, where one of the transduction products; for example, if it is difficult to differentiate number of functional assays that distinguish between recombinant products; where the combination products; for example, if it is difficult to differentiate repair from a homolog as opposed to a sister chromosome. Consequently, we employed the budding yeast Saccharomyces cerevisiae to assay for dominant-negative behavior of the splice variants. To mimic the C-terminal truncations that are predicted to be encoded by the two mouse splice variants, RAD52Δexon4 and RAD52Δintron8, the RAD52-YFP (encoding Rad52 fused to yellow fluorescent protein) gene from Saccharomyces cerevisiae (23) was modified at the yeast chromosomal locus. The RAD52Δexon4 and RAD52Δintron8 mouse splice variants were recreated in the yeast RAD52-YFP gene as rad52Δ77 and rad52Δ284, respectively (the numbers correspond to the final amino acid in the deleted Rad52 sequences). It is important to note that the mouse sequences were not transferred into yeast; rather, the yeast sequence was used to model the mouse transcripts by creating truncations of the endogenous yeast RAD52 gene. We refer to these two yeast alleles as “splice variant equivalents.”

As the mouse variants showed increased HDR when expressed in mammalian cells (Fig. 5), a similar repair assay was performed in yeast. Recombination between direct repeats was assayed using a reporter system containing two defective alleles of LEU2 (leu2ΔEcoRI and leu2ΔBstEI) (Fig. 6A). Similar to the HDR assay used in CHO cells, the leu2 direct-repeat reporter assays primarily intrachromatid and sister chromatid recombination (17, 41). Commonly, two kinds of LEU2+ products result: replacements, where one of the leu2 alleles is repaired without loss of the intervening URA3 sequence; or pop-out events, where the URA3 is lost by deletion of the sequence between the two LEU2 alleles (Fig. 6A). As described in Materials and Methods, these two events are distinguished by a sequence to restore a functional GFP open reading frame. CHO cells were transfected with vectors expressing the normal RAD52 cDNA, the two variant cDNAs, RAD52Δexon4 and RAD52Δintron8, or the control (summarized in Fig. 5B). Expression of RAD52 has no significant effect on HDR in this assay (Fig. 5C), similar to the expression of RAD52 in a cell line mutated in rad52 (43). Interestingly, expression of either of the splice variant constructs elevates the frequency of repair compared to that of full-length RAD52 (Fig. 5C).

Since there is evidence from chicken cells to suggest that RAD52 acts in parallel with XRCC3 in DNA damage repair (10), the mammalian HDR experiments were repeated with a CHO cell line that lacks XRCC3 (CHO-XRCC3−/−) to see how the splice variants act in this mutant. Consistent with previous observations (31), the overall frequency of HDR in this cell line is more than 10-fold lower than in wild-type cells. However, overexpression of the RAD52 splice variants did not change the levels of HDR (Fig. 5D).

Finally, we attempted to knock down the level of the RAD52Δintron8 variant in CHO cells by designing small interfering RNA against the hamster intron 8 sequence. However, transfection of this small interfering RNA had no detectable effect on the level of the intron 8 variant (as judged by RT-PCR) or the frequency of recombination of the GFP targets in the presence of I-SceI (data not shown).

**Splice variants modeled in yeast cause dominant recombination effects.** The ability to conduct functional recombination studies in mammalian cells is currently hindered by the limited number of functional assays that distinguish between recombination products; for example, if it is difficult to differentiate repair from a homolog as opposed to a sister chromosome. Consequently, we employed the budding yeast Saccharomyces cerevisiae to assay for dominant-negative behavior of the splice variants. To mimic the C-terminal truncations that are predicted to be encoded by the two mouse splice variants, RAD52Δexon4 and RAD52Δintron8, the RAD52-YFP (encoding Rad52 fused to yellow fluorescent protein) gene from Saccharomyces cerevisiae (23) was modified at the yeast chromosomal locus. The RAD52Δexon4 and RAD52Δintron8 mouse splice variants were recreated in the yeast RAD52-YFP gene as rad52Δ77 and rad52Δ284, respectively (the numbers correspond to the final amino acid in the deleted Rad52 sequences). It is important to note that the mouse sequences were not transferred into yeast; rather, the yeast sequence was used to model the mouse transcripts by creating truncations of the endogenous yeast RAD52 gene. We refer to these two yeast alleles as “splice variant equivalents.”
simple replica-plating assay. Since the mouse splice variants detected in our screen are expressed together with the normal transcript (Fig. 1B and C, 3, and 4B), we next created diploid yeast strains that contain a full-length *RAD52* allele in addition to a splice variant-equivalent allele to screen for dominant effects on direct-repeat recombination. Both of the *RAD52* splice variant equivalents elevate the frequency of *LEU2* recombinants (Fig. 6B). While the effects upon recombination tested in the same way (D). The difference between the wild-type *RAD52* plasmid and the pRAD52Δexon4 plasmid is not significant (one-tailed *t* test, *P* = 0.094). The difference between the wild-type *RAD52* plasmid and the pRAD52+intron8 plasmid is significant (one-tailed *t* test, *P* = 0.044).
are modest, they are significantly different from those of a heterozygous strain (one-tailed $t$ tests, $P = 8.5 \times 10^{-3}$ and $4.4 \times 10^{-6}$ for rad52Δ77 and rad52Δ284, respectively). These effects are similar to those seen in mammalian cell culture (Fig. 5). Interestingly, in yeast, the rad52Δ77 allele increases replacement events, while the rad52Δ284 allele increases the frequency of URA3 pop-outs (Table 2).

The yeast system has the advantage that the variants are easily assayed for effects on other types of DNA repair. Therefore, we analyzed the splice variant equivalents for their effects on heteroallelic recombination, focus formation, and DNA damage sensitivity. First, spontaneous heteroallelic recombination was assayed in diploid yeast with two defective ade2 heteroalleles (ade2-n and ade2-a). Recombination between these alleles, which reside at each end of the ADE2 gene on homologous chromosomes, restores a functional ADE2 gene (Fig. 7A). Surprisingly, in contrast to the elevated frequency of direct-repeat recombination seen in both mammalian cells (Fig. 5) and yeast (Fig. 6B), both of the splice variant equivalents lowered the frequency of yeast heteroallelic recombination (Fig. 7B). These disparate phenotypes suggest that the variants may be promoting sister chromatid repair while at the same time suppressing repair from a homologous chromosome.

In yeast, Rad52 and other DNA repair proteins form distinct nuclear foci at the sites of double-strand break repair (23–25). These DNA repair centers appear predominantly in S and G2 phases of the cell cycle. Altered levels of foci within yeast strains are indicative of DNA repair deficiencies (25). Next, the formation of foci in the splice variant equivalent strains was measured using fluorescence microscopy. Both splice variant equivalents fail to form spontaneous foci in a haploid strain (data not shown). Diploid strains containing the splice variant equivalents tagged with YFP and wild-type Rad52 tagged with CFP (cyan fluorescent protein) were assayed for focus formation. Interestingly, in these strains, the Rad52Δ284-YFP protein forms foci that also contain wild-type Rad52-CFP (Fig. 8A), suggesting that Rad52Δ284-YFP and wild-type Rad52 are incorporated into the same DNA repair complex. The same phenotype is not observed for Rad52Δ77-YFP. Additionally, both splice variant equivalents elevate the proportion of S/G2 cells that contain spontaneous wild-type Rad52 foci, indicative of either increased DNA damage or an altered DNA repair activity within these cells (Fig. 8B).

Unlike in mammalian cells, RAD52 is essential for HDR in yeast, and furthermore HDR is its major DSB repair pathway. Therefore, DNA damage sensitivity is a standard assay for the function of yeast Rad52. First, we estimated protein levels in diploid strains containing differentially tagged Rad52 peptides (CFP and YFP) to ensure that the splice variant equivalent alleles do not alter the expression of RAD52 from either the wild-type or mutant allele. Table 3 shows the amount of the YFP and CFP detected in the nucleus of the different strains, and there is no significant difference between the two mutant strains and the wild-type diploid. Next, the repair capacity of the splice variant equivalent strains was assayed by measuring viability with increasing doses of $\gamma$-irradiation. Both rad52Δ77 and rad52Δ284 are as sensitive to $\gamma$-irradiation as rad52 null strains, either as haploid strains (data not shown) or as heterozygous diploids containing a null allele (Fig. 9A, first four rows). The diploid strains containing both a mutant and wild-type RAD52 allele were tested for sensitivity to $\gamma$-irradiation, and rad52Δ77 (Fig. 9A, sixth row) is as sensitive as a wild type/null (Fig. 9A, top row). In contrast, rad52Δ284 confers a dominant-negative phenotype for $\gamma$-irradiation sensitivity (Fig. 9A, last row, indicated by the white arrow).

![FIG. 7.](image)

**FIG. 7.** (A) Heteroallelic recombination is measured using two ade2 alleles as shown schematically. The dark triangles indicate the two mutants that inactivate the gene: $ade2-n$ is a fill-in mutation of the AatII site in the 5' end of the gene, and $ade2-a$ is a fill-in mutation of the NdeI site in the 3' end of the gene (12). Recombination between the two homologs restores a functional $ADE2$ gene, as indicated. The gene conversion event depicted has lost the 3' $ade2-n$ mutation, leading to restoration of a functional $ADE2$ gene. The frequency of heteroallelic recombination is calculated using the methods described by Lea and Coulson (22). (B) The median $ade2$ recombination frequency $m$ as defined by Lea and Coulson; the error bars indicate $\pm$ standard deviation of $m$. The wild-type allele is abbreviated wt. Both variant alleles (rad52Δ77/null and rad52Δ284/null) have a lower frequency of heteroallelic recombination than the heterozygous wild-type strain (wt/null) (one-tailed $t$ tests, $P < 10^{-3}$). The frequency of recombination in the diploid wild-type strain (wt/wt) does not differ significantly from the heterozygous wild-type strain (wt/null) (two-tailed $t$ test, $P = 0.87$). The rad52Δ77/null strain has a significantly lower frequency of recombination than the rad52Δ284/null strain (one-tailed $t$ test, $P < 2 \times 10^{-4}$).
Rad52 recruits Rad51 to sites of DNA damage and stimulates Rad51-mediated DNA strand exchange in vitro. Furthermore, overexpression of Rad51 suppresses the phenotype of a Rad52 C-terminal truncation (1, 26). Therefore, we tested whether overexpression of the Rad51 gene could rescue the dominant-negative γ-irradiation phenotype seen for rad52Δ284. The seven yeast strains shown in Fig. 9A were transformed with a plasmid containing the yeast Rad51 gene under the control of a galactose-inducible promoter (pYES-S10-51; a kind gift from Thomas Kodadek). This plasmid complements a rad51 null strain in the presence of galactose (data not shown). Figure 9B shows that the dominant-negative phenotype of rad52Δ284 is partially rescued after induction of Rad51 expression (compare the bottom rows of Fig. 9B, as indicated with a white arrow). This suggests that the radiation-sensitive phenotype of rad52Δ284 is dependent on its disruption of Rad51 function(s).

**DISCUSSION**

**Identification of novel RAD52 transcripts from adult mouse tissue.** Here we identify two novel RAD52 transcripts from adult mouse tissue which are the major alternatively spliced mRNAs from this gene (Fig. 1). One variant aberrantly splices the RAD52 mRNA from exon 3 directly to exon 5, skipping exon 4 (RAD52Δexon4), while the other variant fails to remove the short intron 8 (RAD52+intron8). These two variant mRNAs are predicted to encode truncations of the RAD52 peptide and are likely translated in vivo, since both are found on polyribosomes in the cytoplasm of mouse cells (Fig. 3). Furthermore, a RAD52 IMAGE clone sequence was identified that lacks exon 4 (locus id:BC010673, gi:14715025), and sequences including RAD52 intron 8 were identified from the National Center for Biotechnology Information Mouse UniGene Clusters Map (data not shown). However, definitive identification of these variants using database screening fails, because many cDNA clone sequences are derived from partially spliced transcripts. In addition, examination of the sequences at the intron-exon boundaries of murine RAD52 does not identify deviations from the canonical splice sites to account for the alternate splicing. Thus, a solely bioinformatics approach is inadequate for identifying these novel splice variants, which requires the rigor of the RT-PCR analysis described here.

**Testing the recombination phenotype of variant RAD52 transcripts in cell culture.** RAD52Δexon4 and RAD52+intron8 are predicted to encode truncated mouse RAD52 peptides of 80 and 290 amino acids, respectively (Fig. 2). We next tested the effects of the mouse variants on HDR in mammalian cells using direct repeats of GFP (Fig. 5A). Expression of the murine RAD52 variants increase the frequency of gene replacement compared to wild-type mouse RAD52 (Fig. 5), establishing that these variants are capable of altering the DNA repair phenotype of mammalian cells. Although the increase in HDR is small, it is consistent with the relatively modest effect on this type of assay seen when complementing rad52 null cells (43). Several years ago, Kito et al. speculated that human RAD52 splice variants similar in size to the mouse RAD52Δexon4 transcript might confer a dominant-negative phenotype (16). Here we demonstrate that C-terminal truncations of RAD52 do in

**TABLE 3. Quantification of Rad52 protein levels**

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<th>Quantification of YFP signal mean fluorescence</th>
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</thead>
<tbody>
<tr>
<td>W5937</td>
<td>RAD52-CFP/RAD52ΔYFP</td>
<td>54</td>
<td>477.5 (±71)</td>
<td>257.2 (±12.5)</td>
</tr>
<tr>
<td>W5938</td>
<td>RAD52-CFP/Rad52Δ77-YFP</td>
<td>47</td>
<td>504.6 (±40.7)</td>
<td>239.0 (±2.9)</td>
</tr>
<tr>
<td>W5939</td>
<td>RAD52-CFP/Rad52Δ284-YFP</td>
<td>62</td>
<td>454.4 (±39.9)</td>
<td>264 (±12.5)</td>
</tr>
</tbody>
</table>

*The full genotypes are listed in Table 1. n indicates the number of cells measured for fluorescence. The amount of CFP is greater than YFP in all cases, as CFP gives a brighter fluorescence signal. It should be noted that nuclear fluorescence was measured in all cases, as Rad52 is localized within the nucleus. However, Rad52Δ77-YFP is localized throughout the cell.*
Fact confer a dominant phenotype in mammalian cells by increasing recombination.

**Testing the recombination phenotype of variant RAD52 transcripts in yeast.** The recombination assays available in mammalian cells have a number of limitations. We therefore modeled the splice variants in yeast to allow a further assessment of their role in DNA repair and recombination. A key advantage of modeling the mouse splice variants in yeast is that a range of well-characterized assays are available against which other defined alleles of ScRad52 have previously been tested. Furthermore, the HDR assays available in yeast are more diverse and informative than the current mammalian assays.

The two mouse variants, **RAD52**/Exon4 and **RAD52**/Intron8, were modeled in the ScRad52 gene (**rad52**/Exon4 and **rad52**/Intron8, respectively). These alleles are referred to as “splice variant equivalents,” since they truncate the yeast Rad52 protein at po-
sitions homologous to the mouse variants. The mammalian assay suggests that expression of the mouse variants elevates repair from the same or a sister chromosome (Fig. 5), therefore an equivalent yeast assay was performed. Direct-repeat recombination in yeast (Fig. 6) shows results similar to those seen in mammalian cells (Fig. 5), suggesting that the yeast splice variant equivalents do indeed model their mammalian counterparts. The splice variant equivalents show additional functional differences, which were revealed by further analysis of the LEU2 recombination products from the leu2 direct-repeat assay. For rad52Δ77, the increased recombination is due to a modest increase in replacement events, whereas for rad52Δ284 the increase is due to pop-out events (Table 2). In contrast to the increase in leu2 direct-repeat recombination, the splice variant equivalents dominantly decrease the frequency of recombination between ade2 heteroalleles (Fig. 7). These opposing recombination phenotypes suggest that the variants may bias the HDR pathway away from a homologous template toward a sister or intrachromatid template.

To probe further the dominant effects of the two splice variant equivalents, DNA damage sensitivity was tested. While mammalian rad52 mutant cells exhibit limited DNA damage sensitivity, yeast rad52 strains are exquisitely sensitive to DNA damage. Thus, in yeast, sensitivity to DNA damage provides a simple assay for RAD52 function. As haploid cells, both rad52Δ77 and rad52Δ284 are sensitive to γ-irradiation (data not shown). These results were not surprising, since previous studies showed that truncations removing more than ~200 C-terminal amino acids of ScRad52 render strains as sensitive as a rad52 null strain to methyl methanesulfonate (MMS), a DNA-damaging agent (1, 6). In addition, a deletion of the last 175 codons of ScRAD52 confers a defective DSB repair phenotype (51).

Since the mammalian variants are coexpressed with full-length transcripts (Fig. 1B, C, 3, and 4B), diploid strains containing a splice variant equivalent allele and a wild-type ScRAD52 allele were tested for γ-ray sensitivity. The Rad52Δ284 allele is dominant negative, while the Rad52Δ77 allele is not (Fig. 9A). Previous work has shown that overexpression of two ScRad52 C-terminal truncations confers a dominant-negative phenotype for sensitivity to MMS (26), suggesting that N-terminal Rad52 peptides are capable of disrupting the DNA repair ability of the full-length protein. Interestingly, structural analysis of both the full-length human RAD52 protein and an N-terminal fragment reveals a multimeric complex with RAD52 monomers arranged in a ring configuration (15, 40, 44). Therefore, it is likely that the RAD52+intron8 variant is incorporated into the ring and disrupts its function. This view is supported by our finding that Rad52Δ284 and wild-type Rad2 colocalize in foci (Fig. 8A). In addition, increased levels of wild-type Rad52 foci caused by the splice variant equivalents may indicate increased DNA damage. However, it is unclear how increased damage could cause a specific preference for repair from the same or a sister chromosome. Indeed, DNA damage elevates repair from both sister and homologous chromosomes (13). In addition, elevated Rad52 foci levels in yeast mutants typically indicate delayed or impaired repair processes.

Disruption of the Rad52-Rad51 interaction and its effects on recombination. C-terminal truncations of Rad52 are known to disrupt its interaction with Rad51, thus impairing the formation of Rad51 filaments (19, 26, 37, 39). Such Rad52 C-termi-
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