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An Isolate of Human Immunodeficiency Virus Type 1 Originally Classified as Subtype I Represents a Complex Mosaic Comprising Three Different Group M Subtypes (A, G, and I)

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Full-length reference clones and sequences are currently available for eight human immunodeficiency virus type 1 (HIV-1) group M subtypes (A through H), but none have been reported for subtypes I and J, which have only been identified in a few individuals. Phylogenetic information for subtype I, in particular, is limited since only about 400 bp of env gene sequences have been determined for just two epidemiologically linked viruses infecting a couple who were heterosexual intravenous drug users from Cyprus. To characterize subtype I in greater detail, we employed long-range PCR to clone a full-length provirus (94CY032.3) from an isolate obtained from one of the individuals originally reported to be infected with this subtype. Phylogenetic analysis of C2-V3 env gene sequences confirmed that 94CY032.3 was closely related to sequences previously classified as subtype I. However, analysis of the remainder of its genome revealed various regions in which 94CY032.3 was significantly clustered with either subtype A or subtype G. Only sequences located in vpr and nef, as well as the middle portions of pol and env, formed independent lineages roughly equidistant from all other known subtypes. Since these latter regions most likely have a common origin, we classify them all as subtype I. These results thus indicate that the originally reported prototypic subtype I isolate 94CY032 represents a triple recombinant (A/G/I) with at least 11 points of recombination crossover. We also screened HIV-1 recombinants with regions of uncertain subtype assignment for the presence of subtype I sequences. This analysis revealed that two of the earliest mosaics from Africa, Z321B (A/G/? and MAL (A/D/?), contain short segments of sequence which clustered closely with the subtype I domains of 94CY032.3. Since Z321 was isolated in 1976, subtype I as well as subtypes A and G must have existed in Central Africa prior to that date. The discovery of subtype I in HIV-1 hybrids from widely distant geographic locations also suggests a more widespread distribution of this virus subtype, or at least segments of it, than previously recognized.

A molecular epidemiological survey of human immunodeficiency virus type 1 (HIV-1) conducted in Cyprus in 1994 revealed a surprising degree of genetic variation among locally circulating strains (14). Kostrikis and colleagues screened 25 HIV-1-infected individuals attending an outpatient clinic in Nicosia by performing phylogenetic analyses of C2-V3 env gene sequences amplified from their uncultured peripheral blood mononuclear cells (PBMCs). This identified representatives of several different HIV-1 group M clades, including subtype A, C, and F strains, that are not commonly found in European populations. Moreover, both members of a heterosexual couple with a history of intravenous drug use and documented travel outside of Cyprus were found to be infected with similar viruses that could not be assigned to any of the previously defined HIV-1 subtypes. These viruses formed an independent lineage roughly equidistant from all other group M subtypes, and so it was proposed to classify them as members of a new clade, termed subtype I (14). At about the same time as this initial description of subtype I, it was realized that numerous HIV-1 strains are mosaics of sequences from more than one clade (18, 19). Subsequent confirmation of the widespread occurrence of such hybrid viruses, often with evidence of multiple recombination crossovers along the genome (3, 4, 6, 9, 10, 17, 23), indicated that classification and definition of new subtypes should be based on complete genomic sequences (9, 16, 17). This is particularly important for viruses originating from geographic regions where multiple subtypes cocirculate since these have a high probability of being recombinant. To characterize subtype I in greater detail, we thus cloned a full-length provirus from a short-term-cultured, primary isolate established from one of the two individuals (HO32) infected with this subtype (14). Using primers corresponding to the tRNA primer binding site (5'-TCTCTAagctGGCCGCCAAGGGAC-3', lowercase letters indicate an MluI site) and the polyadenylation signal in the 3' long terminal repeat (LTR) (5'-ACCAGaagctACTAAC AGACGGGACACACTTT-3'), we used long-range PCR

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to amplify nearly full-length genomic fragments (9, 10, 21) that contained all coding and regulatory regions except for 102 bp of 5′ unique LTR sequences (U5). Amplification products were subcloned into a plasmid vector and mapped by restriction enzyme digestion. One clone, termed 94CY032.3, was selected for further analysis. A 694-bp fragment spanning the remainder of the LTR was amplified separately using a seminested approach (10).

The complete sequence of 94CY032.3 was determined by the primer-walking approach. Examination of potential coding regions revealed the expected reading frames for \textit{gag}, \textit{pol}, \textit{vif}, \textit{vpr}, \textit{tat}, \textit{rev}, \textit{vpu}, \textit{env}, and \textit{nef} (data not shown). None of the genes contained major deletions, insertions, or rearrangements. However, both env and vif genes contained single in-frame stop codons. There was also a single-base-pair insertion at position 5199 which caused a frameshift and altered six amino acid residues at the C terminus of the Vpr protein. All other protein domains of known function as well as major regulatory sequences, including the primer binding site, the packaging signal, and major splice sites, appeared to be intact. Similarly, the number, position, and consensus sequences of promoter and enhancer elements in the 94CY032.3 LTR were indistinguishable from those of most other HIV-1 strains, except for the presence of an unusual TATA sequence (TAAAA), thus far only found in subtype E (A/E) viruses from Thailand and the Central African Republic (4, 10).

To compare 94CY032.3 to previously reported subtype I sequences, we constructed a phylogenetic tree from C2-V3 sequences, including representatives of all 10 known group M subtypes (Fig. 1). As expected, 94CY032.3 clustered most closely with CYHO321 and CYHO322, sequences amplified from uncultured PBMC DNA of the same individual (HO32) from whom the 94CY032 isolate was derived. 94CY032.3 also clustered very closely with CYHO311, a sequence derived from the sexual partner of HO32 (14), strongly suggesting that the two infections were epidemiologically linked. Finally, as observed in the past (14), all subtype I sequences clustered independently, forming a distinct lineage roughly equidistant from all other subtypes, including subtype J (15). These findings thus confirmed the authenticity of the 94CY032.3 clone and validated it as a representative of subtype I in the C2-V3 region of the viral envelope.

FIG. 1. Confirmation of 94CY032.3 (highlighted) as a subtype I representative in the C2-V3 envelope region. Reference sequences for all known group M subtypes were obtained from the Los Alamos sequence database (13), aligned using CLUSTAL W (26), and adjusted manually using the alignment editor MASE (7). Sites for which there was a gap in any of the sequences were excluded from all sequence comparisons. Pairwise evolutionary distances were estimated by using Kimura’s two-parameter method to correct for superimposed substitutions (12). Phylogenetic trees were constructed by the neighbor-joining method (20), and the reliability of topologies was estimated by performing bootstrap analysis using 1,000 replicates (8). Values at nodes indicate the percentage of bootstraps in which the cluster to the right was found (bootstrap values of 80% and higher are shown). Branch lengths are drawn to scale.
To characterize the remainder of the 94CY032.3 genome, we next performed pairwise sequence comparisons with recently reported nonmosaic reference sequences for subtypes A through H (9, 16) as well as selected intersubtype recombinants (17). We have used this approach in the past to screen newly derived sequences for regions of unusual sequence similarity or dissimilarity that might indicate recombination (9, 10). Briefly, 94CY032.3 was added to a multiple genome align-
FIG. 3. Bootstrap plot analysis to map recombination breakpoints in 94CY032.3. Bootscanning was performed essentially as described previously (9, 10, 22), plotting the magnitude of the bootstrap value supporting the clustering of 94CY032.3 with 92UG037.1 (subtype A in red) in comparison with that of 94CY032.3 and 92NG083.2 (subtype G in green) for a window of 400 bp moved in increments of 10 bp along the alignment. Regions of subtype A or G origin are identified by very high bootstrap values (>80%). The location of eight recombination crossovers is indicated (see text for details). Breakpoint analysis between positions 4200 and 4800 was not possible due to the recombinant nature of 92NG083.2 (shaded) (9). The beginnings of gag, pol, vif, vpr, env, and nef open reading frames are shown. The y axis indicates the percent bootstrap replicates which support the clustering of 94CY032.3 with representatives of the respective subtypes.
fied two A/G crossovers around positions 1200 and 5600 and one G/A crossover around position 4100. The bootstrap plots also outlined regions with no peaks or peaks below 80%, which coincided with segments that clustered independently in the exploratory tree analysis and thus were likely in subtype I. Delineating the boundaries of these regions suggested five additional breakpoint positions: G/I at 1500, I/G at 3800, G/I at 6000, I/A at 6900, and A/I at 7200. Because full-length, non-mosaic reference sequences for the parental lineages (G and I) were not available, most of the breakpoints could not be

FIG. 4. Recombination breakpoint analysis of 94CY032.3 in the vif/vpr region. (A) Diversity plots comparing 94CY032.3 to reference sequences from the database (U455, MN, and ELI) as well as to a recently identified (partial) nonrecombinant subtype G sequence (95ML045) (2). Sequences are color coded. Distance values were calculated for a window of 300 bp moved in steps of 10 nucleotides. The x axis indicates the nucleotide positions along the alignment (gaps were stripped and removed from the alignment). The positions of the start codons of the vif, vpr, and tat genes are shown. The y axis denotes the distance between the viruses compared (0.05 = 5% difference). (B) Neighbor-joining trees depicting the position of 94CY032.3 in regions flanking the breakpoints identified in panel A. Trees were constructed from the genomic regions indicated. Subtypes are identified by brackets. Four sequences from Mali represent subtype G; these are the only available subtype G reference sequences in this region, since all other characterized subtype G viruses contain A fragments (2, 9, 16, 17). Numbers at nodes indicate the percentage of bootstrap values with which the adjacent cluster is supported; only values above 80% are shown. Branch lengths are drawn to scale.

FIG. 5. Inferred structure of the mosaic 94CY032.3 genome. Regions of different subtype origin are color coded. All breakpoints are tentative, since full-length nonrecombinant subtype G and I reference sequences are not available. LTR sequences were not analyzed and are thus shown as hatched boxes.
mapped with certainty. However, the A/G breakpoints at positions 1200 and 5600 were confirmed by informative site analysis (data not shown). The recombinant nature of 92NG083.2 prohibited reliable breakpoint analysis between positions 4200 and 4800 (9, 16) (highlighted in Fig. 3).

To map the recombination breakpoints in this remaining region, we made use of four recently reported, partial but nonmosaic subtype G sequences from Mali which spanned the \textit{vif/vpr} region and thus bridged the subtype A gap of 92NG083.2 (2). Figure 4A illustrates a set of distance plots that compare 94CY032.3 to one of these newly derived G sequences (95ML045) as well as representatives of subtype A (U455), B (MN), and D (ELI), respectively. Consistent with the results from the exploratory tree analysis (Fig. 2), 94CY032.3 was disproportionately more closely related to U455 in the 5' and 3' thirds of this fragment, suggesting the presence of subtype A-like segments. However, in the middle of the fragment, 94CY032.3 was clearly equidistant from U455 and the other subtypes, suggesting an independent position. Thus, noting the points at which the “A” distance increased and decreased relative to the other distances allowed us to map the two remaining breakpoints, one near position 4650 and the other near position 5000. Trees constructed from sequences surrounding these two breakpoints (Fig. 4B) confirmed that 94CY032.3 switched position from subtype A (panel 4255-4650) to subtype I (4651-5000) and back to subtype A (5001-
5300). Note that the new subtype G sequences only cover the region between positions 4255 and 5300.

Figure 5 summarizes the results from all phylogenetic analyses, depicting a schematic representation of the mosaic genome structure of 94CY032.3. Segments of different subtype origin are color coded, and there are a total of 10 recombination breakpoints between the 5′ end of gag and the 3′ end of nef LTR sequences were not separately analyzed for mosaicism, but the discordant subtype assignments of the gag and nef regions necessitate at least one more breakpoint within either the viral LTR or the gag leader sequence. Given this extent of mosaic complexity, 94CY032.3 must be the result of multiple recombination events, either in the same or different individuals.

Having identified several fragments of subtype I in 94CY032.3, we next determined whether there was any evidence for its presence in other full-length recombinants from the database. Two known mosaiics, MAL (1, 19) and Z321 (5), were of particular interest, because previous analyses had indicated that these viruses contain regions of uncertain subtype assignment (17, 19). For example, MAL has long been known to represent a mosaic of subtypes A and D (color coded as red and blue, respectively, in Fig. 6A) but also contains a sizable pol fragment that has defined previous subtype classification (white in Fig. 6A) (17, 19). Similarly, Z321 is a known mosaic of subtypes A and G (red and green, respectively, in Fig. 6A (5)), but a recent reanalysis of its recombination breakpoints also identified regions that could not be assigned to any known subtype (17). To determine whether any of these regions represented subtype I, we again performed distance plot analysis, looking for dips in the diversity profiles of 94CY032.3, with MAL and Z321 as an indication of relatively greater sequence similarity. Indeed, two such dips were identified, one in the pol region of MAL and another in the vif/vpr region of Z321, both of which coincided with previously unclassified segments of their genomes (data not shown). Subsequent phylogenetic tree analysis confirmed that these regions were indeed of subtype I origin, since MAL and Z321 clustered with very high bootstrap values with the subtype I domains of 94CY032.3 (Fig. 6B and C). However, subtype I did not account for all of the unclassifiable regions in MAL and Z321 (17), and it thus remains unclear whether these represent still other, as yet unidentified, subtypes or regions of multiple breakpoints that cannot be mapped by using current methods.

In summary, we show here that a strain of HIV-1, proposed in 1995 as a prototypic subtype I isolate (14), in fact represents a complex mosaic comprising subtypes A, G, and I, respectively. We also show that two of the oldest known isolates from Africa, MAL isolated in 1984 (1) and Z321 isolated in 1976 (11, 25), contain short segments of sequence closely related to the subtype I domains of 94CY032.3. These findings support the following conclusions. (i) Although initially detected in Cyprus, subtype I must have existed in Africa as early as 1976; it is unknown whether full-length, nonmosaic representatives of subtype I still exist but have not yet been sampled or whether this subtype is represented only by fragments in present-day recombinants. (ii) The ancestry of 94CY032.3 has likely involved multiple recombination events; it remains unclear whether these occurred in Africa and/or in Cyprus, where a number of different subtypes have also been documented (14). (iii) Subtype I, along with subtypes A and G, must have diverged substantially earlier than the 1970s in order to be detectable as a distinct segment in the Z321 genome: this is consistent with the recent molecular characterization of a virus from 1959 which in phylogenetic analyses appears to have postdated the group M radiation (27). (iv) Finally, the finding of subtype I in several different recombinants, including one from an intravenous drug user (14), suggests that this subtype may be more widespread than previously thought, at least in the form of mosaic genome fragments. It will be interesting to screen additional viruses from drug user populations and their contacts in Cyprus and Greece to determine the current prevalence and geographic distribution of subtype I-containing viruses.

**Nucleotide sequence accession numbers.** The complete sequence of 94CY032.3 has been deposited at GenBank under accession no. AF049337 and AF049338.

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