While K-ras is essential for mouse development, expression of the K-ras 4A splice variant is dispensable

Citation for published version:

Digital Object Identifier (DOI):

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Molecular and Cellular Biology

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
While K-ras Is Essential for Mouse Development, Expression of the K-ras 4A Splice Variant Is Dispensable

Sarah J. Plowman,1 D. James Williamson,2† Maureen J. O’Sullivan,2 Jennifer Doig,1 Ann-Marie Ritchie,1 David J. Harrison,2 David W. Melton,1 Mark J. Arends,3* Martin L. Hooper,1 and Charles E. Patek1*

Sir Alastair Currie Cancer Research UK Laboratories, Molecular Medicine Centre, Western General Hospital, Crewe Road, Edinburgh EH4 2XU,1 Division of Pathology, University of Edinburgh Medical School, Edinburgh EH8 9AG,2 and Department of Pathology, Addenbrooke’s Hospital, University of Cambridge, Cambridge CB2 2QQ,3 United Kingdom

Received 30 June 2003/Returned for modification 22 August 2003/Accepted 18 September 2003

In mammals, the three classical ras genes encode four highly homologous proteins, N-Ras, H-Ras, and the isoforms K-Ras 4A and 4B. Previous studies have shown that K-ras is essential for mouse development and that while K-ras 4A and 4B are expressed during development, K-ras 4A expression is regulated temporally and spatially and occurs in adult kidney, intestine, stomach, and liver. In the present study, the pattern of K-ras 4A expression was examined in a wide range of wild-type adult mouse tissues, and gene targeting was used to generate K-ras 4A-deficient mice to examine its role in development. It was found that K-ras 4A is also expressed in uterus, lung, pancreas, salivary glands, seminal vesicles, bone marrow cells, and cecum, where it was the major K-Ras isoform expressed. Mating between K-ras4A+/+ mice produced viable K-ras4A+/tm and K-ras4A+/tm offspring with the expected Mendelian ratios of inheritance, and these mice expressed the K-ras 4B splice variant only. K-ras4A4A/tm mice were fertile and showed no histopathological abnormalities on inbred (129/Ola) or crossbred (129/Ola × C57BL/6) genetic backgrounds. The results demonstrate that K-Ras 4A, like H- and N-Ras, is dispensable for normal mouse development, at least in the presence of functional K-Ras 4B.

The three classical mammalian ras genes, K-, N-, and H-ras, encode proteins that are members of the guanine nucleotide binding protein superfamily (8). Ras proteins are small (~21 kDa) GTPases which cycle between inactive (GDP-bound) and active (GTP-bound) conformations at the plasma membrane, by interaction with a variety of guanine nucleotide exchange factors and GTPase activating proteins in response to stimulation by a diverse array of cell surface receptors, including the epidermal growth factor receptor and cytokine receptors such as interleukin-2 (14). Following activation, Ras proteins bind and activate a plethora of downstream effector proteins, including Raf kinases and phosphatidylinositol 3-kinases, and by this means control many cellular functions, including proliferation, differentiation, migration, and apoptosis (4, 11, 28).

Activating point mutations leading to constitutive activation of the Ras proteins are associated with some 30% of all human malignancies (1), and these mutations render Ras insensitive to the regulatory action of GTPase activating proteins, leading to excessive and inappropriate signaling, resulting in the promotion of cellular transformation. However, while mutationally active ras genes are generally believed to act as dominant oncogenes, recent reports indicate that wild-type K- and N-Ras proteins have tumor suppressor activity (7, 32). Thus, K- and N-ras appear to exert a dual function in that they promote cancer development as gain-of-function oncogenes when mutated and inhibit cancer by tumor suppressor activity when wild type (proto-oncogene).

The high degree of homology between Ras proteins suggests that functional redundancy may exist among these proteins, yet mounting evidence exists for unique roles for the ras gene family members. For example, most human cancers with ras activating mutations are associated exclusively with a particular ras gene, suggesting tissue-specific activity: K-ras mutations are prevalent in lung (~30%), colon (~40%), and pancreatic (~90%) cancers, while H-ras and N-ras activating mutations are associated with bladder cancer and myeloid leukemia, respectively (1). Furthermore, while N-ras and H-ras are dispensable for development both individually and in combination (10, 26), mice harboring a homozygous K-ras null mutation (K-ras+/−) are not viable and die between day 12.5 post coitum and term (depending on the genetic background) due to cardiac, liver, neurological, and hematopoietic defects (18, 19).

Thus, of the classical ras gene family members (K-, N-, and H-ras), only K-ras is necessary for embryonic development. However, the findings that (i) N-ras−/− mice are healthy but N-ras+/−, K-ras−/−/− mice die in utero (18), (ii) K-ras−/−/− N-ras−/−/− mice exhibit a more severe phenotype than K-ras−/−/− mice (18), and (iii) fewer than expected H-ras−/− N-ras−/− mice survive embryogenesis (10) underline the essential role for K-Ras in development and imply partial functional overlap between different Ras proteins.

Understanding the role(s) of K-Ras in development and tissue function is an important route to gain insight as to how K-ras activating mutations promote neoplastic change. How-

*Corresponding author. Mailing address: Sir Alastair Currie Cancer Research United Kingdom Laboratories, Molecular Medicine Centre, Western General Hospital, Crewe Road, Edinburgh EH4 2XU, United Kingdom. Phone: 44(0)131-651-1080. Fax: 44(0)131-651-1072. E-mail: charles.patek@ed.ac.uk.
†Present address: Department of General Medicine, Sir Charles Gardiner Hospital, Nedlands, Perth 6009, Western Australia, Australia.
ever, this is complicated by the fact that the K-ras gene encodes two protein isoforms, K-Ras 4A and K-Ras 4B, of 189 and 188 residues, respectively, by alternative splicing of the fourth coding exons 4A and 4B, and K-ras activating mutations that usually affect codons 12, 13, and 61 jointly affect both isoforms (5, 9, 21). The isoforms differ significantly at their C termini after residue 165, including the hypervariable domains and CAAX motifs. These regions are involved in membrane association, which is essential for Ras function (16) via a series of posttranslational modifications, which include isoprenylation, endoproteolysis, and methylation (25). However, these modifications differ between the isoforms due to the sequence differences at the hypervariable domain: K-Ras 4A, like N- and H-Ras, is palmitoylated at cysteine residues upstream of the CAAX motif, which are replaced with a polylsine domain in K-Ras 4B.

The difference in C-terminal modifications of Ras proteins leads to alternative trafficking pathways to the plasma membrane (3) and ultimately localization to different plasma membrane microdomains (23, 24). Indeed, reports that H-, N-, and K-Ras differentially affect Raf-1, phosphatidylinositol 3-kinase, and Rap (28, 29, 31) suggest that they elicit divergent biological responses by interacting with different subsets of downstream effectors. Thus, the posttranslational differences between K-Ras 4A and K-Ras 4B could affect their membrane localization and, therefore, interaction with different membrane targets. The specific association between K-Ras 4B and the guanine nucleotide exchange factor Smg GDS (27) and reports that the C-terminal modifications of Ras proteins play an important role in the recruitment of the guanine nucleotide exchange factor to the plasma membrane support this hypothesis.

The targeting vector contains a neomycin resistance cassette (containing a phosphoglycerate kinase I promoter) and the herpes simplex virus thymidine kinase gene of exon 4B (13). The neomycin resistance cassette (containing a phosphoglycerate kinase I promoter) and the herpes simplex virus thymidine kinase gene were employed for positive and negative selection, respectively (Fig. 1A), which amplify a 2.1-kb product. The PCR conditions were 1.0 μM each primer (Sigma), 1× PCR buffer (Invitrogen), 1.5 mM MgCl₂ (Invitrogen), 200 μM each deoxynucleoside triphosphate (Amersham) (1× TdT) and the primer set genomic DNA per reaction. Following denaturing for 4 min at 94°C, DNA was amplified for 45 cycles (94°C for 1 min, 58°C for 1 min, and 72°C for 2.5 min).

Homologous recombination was confirmed by Southern blotting. Genomic DNA (20 μg) was digested with HindIII or PvuII and, after electrophoresis, was transferred to a nylon membrane (Genescreen) in denaturing buffer (0.5 M NaOH, 1.5 M NaCl). The membrane was neutralized and prehybridized at 65°C for 2 h (10% dextran sulfate, 1% sodium dodecyl sulfate, 6× SSC [1× SSC is 0.15 M NaCl plus 0.015 M sodium citrate]), 200 μg of herring sperm DNA per ml). The 3′ internal and 5′ external probes (Fig. 1A) were labeled with [32P]dCTP by random priming (Stratagene) and hybridized to the membrane at 65°C overnight. Membranes were washed at room temperature with 2× SSC (twice for 5 min), at 65°C with 2× SSC–1% sodium dodecyl sulfate (twice for 30 min), then at room temperature with 0.1× SSC (twice for 30 min). Membranes were exposed to film overnight at –70°C.

Production of K-Ras 4A-deficient mice. Embryonic stem cells harboring the heterozygous deletion of exon 4A (K-ras<sup>tm<sub>4A/4A</sub></sup>) were injected into day 3.5 post coitum C57BL/6 blastocysts, which were implanted into the uterus of day 2.5 post coitum C57BL/6 pseudopregnant recipient females. Male chimeras were mated with 129/Ola or C57BL/6 females to generate inbred and crossed lines, and germ line transmission of embryonic stem cell markers was identified by coat color (light yellow and agouti, respectively).

Since long-range PCR proved unreliable for analyzing tail biopsy DNA, mice were genotyped with two different PCRs. In reaction 1, the primer set neo52 (5′-GATGTCCTCTGGCAGATACGGT-3′) and neo22 (5′-CGATAGAGCGCGATGCTGGCAATG-3′), positioned in the neomycin cassette (Fig. 1A), amplify a 206-bp product in K-ras<sup>tm<sub>4A/4A</sub></sup> and K-ras<sup>tm<sub>4A/4A</sub></sup> mice. In reaction 2, the primer set PxsAS (5′-CATTGGTGAGAGAGTCGGCATGCACAGT-3′) and PxsAA (5′-TCACAGCCAGGATCCTTTCTTC-3′), positioned in exon 4A (Fig. 1A), amplify a 72-bp product in wild-type and K-ras<sup>tm<sub>4A/4A</sub></sup> mice. In both reactions, DNA was denatured for 4 min at 94°C and amplified for 30 cycles at 94°C for 30 s and 58°C for 30 s. The genotypes were confirmed by Southern blotting with internal and external probes as described above.

In the inbred colony, first-generation K-ras<sup>tm<sub>4A/4A</sub></sup> males (which do not harbor the Hprt null mutation) were mated with wild-type 129/Ola mice to generate an inbred stock that was wild type for Hprt. In the crossed colony, the Hprt mutation was allowed to segregate. All animal work was carried out under Home Office license, with ethically approved methods as set out by the UK Coordinating Committee on Cancer Research in their guidelines on the Welfare of Animals in Experimental Neoplasia.

RESULTS

Generation of K-Ras 4A-deficient mice. The targeting vector pPTKiNKi4A was designed to replace exon 4A of K-ras with a neomycin resistance cassette (Fig. 1A). Of 324 embryonic stem clones screened by PCR, 17 were positive for homologous recombination, amplifying the correct 2.1-kb PCR product (Fig. 1B). The genotype of seven of these clones was confirmed by Southern analysis with a 5′ external probe that identified a 5-kb wild-type and a 4-kb targeted band and a 3′ internal probe that identified a 2-kb wild-type and a 6-kb targeted band (Fig.
1C). Of nine chimeras (generated from three different embryonic stem cell clones), five transmitted embryonic stem cell-derived coat color markers through the germ line. Since the effects of mutations, including the K-ras null mutation (18), can vary greatly with the genetic background, the consequence of deleting K-ras 4A expression was examined in inbred (129/Ola) and crossbred (129/Ola/129X/H11003/C57BL/6) strains of mice. The embryonic stem cell-derived offspring were genotyped at weaning by PCR (with the primer sets neo22/neo52 and Px4AS/Px4AA, which amplify the neomycin cassette and exon 4A, respectively; Fig. 2A) and by Southern blotting (with the 5' external and 3' internal probes; Fig. 2B), and it was found that K-ras<sup>tm/-/H9004</sup>4A/+ and wild-type mice were present in the expected Mendelian ratio (54 and 61, respectively; χ² = 0.426, P = 0.514).

**K-ras 4A-deficient mice are healthy and fertile.** Offspring from crosses between K-ras<sup>tm/-/H9004</sup>4A/+ mice were genotyped by PCR (Fig. 2A) and Southern blotting (Fig. 2B). Wild-type, K-ras<sup>tm/-/H9004</sup>4A/+, and K-ras<sup>tm/-/H9004</sup>4A/- offspring were present in the expected Mendelian ratios on both crossbred (48, 101, and 58, respectively; χ² = 1.087, P = 0.581) and inbred (11, 20, and 10, respectively; χ² = 0.073, P = 0.964) genetic back-

---

**FIG. 1.** Targeted deletion of the K-ras 4A splice variant in embryonic stem cells. (A) Schematic representation of the mouse K-ras locus and the targeting vector used to replace exon 4A. The exons are indicated by open boxes. The neomycin resistance cassette and the herpes simplex virus thymidine kinase gene cassette were employed as selectable markers. In the targeting vector, the thin line denotes plasmid DNA. The positions of the 5' external and 3' internal probes (solid boxes) and PCR primers (arrows) used for genotyping are indicated. Restriction enzyme sites: E, EcoRI; P, PvuII; H, HindIII. (B) Embryonic stem cells were genotyped by PCR with the primer set neo22 and Px4BA, which amplify a 2.1-kb product in clones that harbor the targeted allele (clones 173, 187, 194, 196, 221, 226, and 248). +/+, wild-type embryonic stem cells; −, PCR negative control; M, 1-kb DNA ladder showing the 2.0- and 2.1-kb size markers. (C) Confirmation of homologous recombination in embryonic stem cells by Southern blotting. Following digestion with HindIII, a wild-type 2-kb and a targeted 6-kb fragment were detected with the 3' internal probe (upper panel). Following digestion with PvuII, a wild-type 5-kb and a targeted 4-kb band were detected with the 5' external probe (lower panel). +/+, wild-type embryonic stem cells.
grounds, indicating that all K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice develop normally. In addition, further breeding studies found that male and female K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} offspring were present in the expected Mendelian ratios in both crossbred (113 and 113, respectively) and inbred (20 and 25, respectively; \( \chi^2 = 0.556, P = 0.456 \)) stocks. Male and female K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice were fertile, and females successfully weaned their young.

Detailed histopathological analysis was undertaken of 24 crossbred mice at 3 months, including 12 wild-type and 12 K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice, with six males and six females in each cohort. No abnormalities were detected at necropsy, and examination of liver, kidney, large and small intestine, stomach, pancreas, spleen, thymus, heart, lung, brain, ovary, uterus, seminal vesicles, and testis found no difference between tissues from wild-type and K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice. Inbred K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice were also fertile and outwardly healthy, and histopathological analysis of a single animal at 3 months found no abnormalities. The oldest K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice in the crossbred colony are currently 8 months old, and these too are outwardly healthy.

**FIG. 2.** Genotyping of mice. (A) The upper panel shows amplification of K-ras exon 4A by PCR. The 72-bp product is present in wild-type (+/+), lanes 9 and 10) and K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} (+/−, lanes 14 and 15) mice, but not in K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice (−/−, lanes 5 and 11). The lower panel shows amplification of the neomycin cassette by PCR. The 206-bp product is present in K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} and K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice but not in wild-type mice. (B) Confirmation of genotypes by Southern blotting. Tail DNA digested with HindIII and hybridized with the 3’ internal probe (upper panel) gave a 2-kb fragment for the wild-type allele and a 6-kb fragment from the targeted allele. Only the 6-kb fragment is present in K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice. Digestion with PvuII and hybridization with the 5’ external probe (lower panel) gave a 5-kb band for the wild-type allele and a 4-kb fragment for the targeted allele. Only the 4-kb fragment is present in K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice.

K-ras 4A-deficient mice express the K-ras 4B splice variant only. To confirm that the modification of the K-ras gene, by the introduction of the targeting vector, resulted in the deletion of the K-ras 4A splice variant, RT-PCR was used to examine K-ras expression in the large intestine of wild-type and K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice. Only the K-ras 4B splice variant was expressed by K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice (Fig. 3).

**FIG. 3.** Expression of K-ras in the large intestine from wild-type and K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} mice. RT-PCR analysis of K-ras 4A and 4B mRNAs with primers positioned in exons 1 and 4B that amplify both splice variants in the same reaction (22). The 687-bp band corresponds to K-ras 4A, and the 565-bp band corresponds to K-ras 4B. Wild-type (+/+), but not K-ras\textsuperscript{tm4A\textit{A}tm4A\textit{A}} (−/−) mice express K-ras 4A. Lane M, 100-bp size markers; the major band shown is 600 bp.

**FIG. 4.** K-ras 4A and 4B expression in adult mouse tissues. The PCR primers used amplify both K-ras splice variants in the same reaction (22). The K-ras 4A and 4B mRNAs generate PCR products of 687 and 565 bp, respectively. Tissues: Ht, heart; Li, large intestine; Si, small intestine; Ki, kidney; Liv, liver; St, stomach; Lu, lung; Ov, ovary; Ut, uterus; Adr, adrenal gland; Sg, salivary gland; Sp, spleen; Br, brain; Pan, pancreas; Bm, bone marrow; Th, thymus; Cae, cecum; Sv, seminal vesicles; Tt, testis. −, PCR negative control. *, 100-bp ladder; the major band shown is 600 bp.
contrast, K-ras 4B was expressed ubiquitously and was the only isoform expressed in brain, spleen, heart, adrenal gland, thymus, testis, and ovary. While both isoforms were expressed at similar levels in stomach and large intestine, K-ras 4A was the major isof orm expressed in the occum.

DISCUSSION

To gain further understanding concerning the role(s) of K-Ras in development and tissue function, we examined the pattern of K-ras 4A expression in wild-type adult mouse tissues and used gene targeting to delete K-ras 4A expression. While previous studies have established that expression of the K-ras gene is essential for mouse development (18, 19), here we found that, even though K-ras 4A is expressed during development and in adult tissues (22, 30; this study), all K-ras\textsuperscript{tm\Delta 4A/tm\Delta 4A} mice on both inbred and crossbred genetic backgrounds developed normally, and the adult mice are fertile and healthy.

The finding that K-Ras 4A is dispensable for development suggests that the embryonic lethality of K-ras\textsuperscript{−−/−} mice (which do not express either isof orm) may result solely from failure of expression of the K-Ras 4B isof orm. However, the very fact that K-ras 4A is expressed during development, albeit in a spatially and temporally regulated manner (22), raises the possibility that the lethal K-ras\textsuperscript{−−/−} phenotype could result from loss of synergistic function of the two protein isoforms or else from loss of a critical function(s) that can be performed by either isof orm, such that one or the other is necessary but in the absence of both development cannot occur. While the latter possibility is less likely, given that only the K-Ras 4B isof orm is expressed ubiquitously throughout development (22), questions concerning whether expression of the K-Ras 4B isof orm is essential for normal development and whether K-Ras 4A influences its action wait to be addressed by comparing the phenotypes of K-ras\textsuperscript{−−/−} and K-ras 4B-deficient mice on the same inbred genetic background. It is nevertheless important to stress that whatever the result of these future studies, our conclusion that K-Ras 4A is dispensable for normal mouse development in the presence of functional K-Ras 4B remains valid.

The C-terminal modifications of Ras proteins are essential for protein function (16), and functional differences between Ras proteins may reflect, at least in part, these different modifications. Therefore, the findings by Hancock et al. that N-, H-, and K-Ras 4A undergo similar C-terminal modifications which involve palmitoylation of cysteine residues upstream of the common CAAX motif (14) are significant because these proteins are dispensable for normal development. Furthermore, the viability of K-ras\textsuperscript{tm\Delta 4A/tm\Delta 4A} mice is unlikely to reflect a compensatory upregulation of these closely related Ras proteins, since the simultaneous deletion of both K-ras splice variants does not result in the upregulation of H-ras or N-ras expression in either tissues or fibroblast cultures from K-ras\textsuperscript{−−/−} embryos (18). Likewise, H-ras\textsuperscript{−−/−} N-ras\textsuperscript{−−/−} mutant mice show no change in K-ras expression (10).

Importantly, since N-ras\textsuperscript{−−/−}, H-ras\textsuperscript{−−/−}, and K-ras\textsuperscript{tm\Delta 4A/tm\Delta 4A} mice are fertile, it is now possible, by crossing these mice, to formally test whether the viability of K-ras\textsuperscript{tm\Δ4A/tm\Δ4A} mice is indeed independent of N- and H-ras expression. Also, since K-Ras 4B, unlike K-Ras 4A, associates with the plasma mem-
why they are linked with particular types of epithelial cell-derived tumors.

ACKNOWLEDGMENTS

This study was supported by a Medical Research Council studentship and grants from the Cancer Research Campaign (CRC, now Cancer Research UK), the Melville Trust for the Care and Cure of Cancer, Action Research for the Crippled Child (new Action Medical Research), and the Row Fogo Charitable Trust.

REFERENCES


