Cdk1-Dependent Phosphorylation of Cdc13 Coordinates Telomere Elongation during Cell-Cycle Progression

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SUMMARY

Elongation of telomeres by telomerase replenishes the loss of terminal telomeric DNA repeats during each cell cycle. In budding yeast, Cdc13 plays an essential role in telomere length homeostasis, partly through its interactions with both the telomerase complex and the competing Stn1-Ten1 complex. Previous studies in yeast have shown that telomere elongation by telomerase is cell cycle dependent, but the mechanism underlying this dependence is unclear. In S. cerevisiae, two major telomeric DNA-binding proteins are Rap1 and Cdc13. While Rap1 binds duplex C1-3A/TG1-3 DNA repeats (Conrad et al., 1990; Wright et al., 1992; Wright and Zakian, 1995), Cdc13 binds the single-stranded TG1-3 3′ overhang (Bourns et al., 1998). Cdc13 has been implicated as the major regulator of telomere access by the telomerase complex (Evans and Lundblad, 1999; Hughes et al., 2000; Lendvay et al., 1996; Lin and Zakian, 1996; Nugent et al., 1996). Cdc13 can interact with two distinct complexes that either positively or negatively regulate telomere elongation (Chandra et al., 2001). Association of Cdc13 with Est1 is essential for recruitment of the telomerase holoenzyme, which contains the protein catalytic subunit Est2 and the integral RNA template TLC1, as well as Est1 and Est3 (Lendvay et al., 1996). Cdc13 also interacts with the Stn1-Ten1 protein complex, which negatively regulates telomere elongation and plays an essential role in telomere end protection (Grandin et al., 2001). Hence, Cdc13 tightly regulates telomere elongation through its interactions with both Est1-Est2-Est3-TLC1 complex and the competing Stn1-Ten1 complex (Chandra et al., 2001). However, the mechanism by which Cdc13 coordinates the binding of these two complexes during telomere elongation had not previously been analyzed.

Telomere length homeostasis is maintained through a dynamic process. Telomerase extends the telomeric TG1-3 single-stranded DNA overhang by copying the intrinsic telomerase RNA template, while lagging strand synthesis by DNA polymerases α and δ is inferred to fill the terminal 5′ gap during each cell cycle (Diede and Gottschling, 1999). Previous studies have shown that telomere elongation by telomerase is restricted to late S to G2 phases in vivo (Diede and Gottschling, 1999; Marcand et al., 2000). The timing of telomere elongation in late S and G2 phases correlates with the binding of protein factors involved in telomere elongation, including Est1, Est2, and Cdc13 (Schramke et al., 2004; Taggart et al., 2002). These data have suggested that the assembly of a functional telomerase complex at the telomeres is restricted to late S to G2 phases of the cell cycle.

INTRODUCTION

In most eukaryotes, the ends of linear chromosomes are capped by telomeres. Telomeres are essential for both the stability of linear chromosomes and the complete replication of genomic information (Blackburn, 2000). Telomeres are maintained by telomerase, whose activity is highly regulated. In yeast, more than 150 genes affect telomere length maintenance (Askree et al., 2004; Gatbonton et al., 2006). In humans, haploinsufficiency for telomerase RNA in dyskeratosis congenita patients leads to progressive bone marrow failure and premature aging (Chen and Greider, 2004). Hence even a reduction in gene dosage has severe clinical consequences, highlighting the importance of telomerase regulation.

In the budding yeast S. cerevisiae, the telomeric DNA consists of 250–350 base pairs of double-stranded C1-3A/TG1-3 repeats with a short single-stranded TG1-3 3′ overhang (Wellinger et al., 1993; Zakian, 1996). Sequence-specific telomeric DNA-binding proteins and their associated factors form a dynamic structure at telomeres (Laroche et al., 2000; Smith et al., 2003). This dynamic nucleoprotein structure is essential for telomere silencing, telomere end protection, and telomere length regulation. In S. cerevisiae, two major telomeric DNA-binding proteins are Rap1 and Cdc13. While Rap1 binds duplex C1-3A/TG1-3 DNA repeats (Conrad et al., 1990; Wright et al., 1992; Wright and Zakian, 1995), Cdc13 binds the single-stranded TG1-3 3′ overhang (Bourns et al., 1998). Cdc13 has been implicated as the major regulator of telomere access by the telomerase complex (Evans and Lundblad, 1999; Hughes et al., 2000; Lendvay et al., 1996; Lin and Zakian, 1996; Nugent et al., 1996). Cdc13 can interact with two distinct complexes that either positively or negatively regulate telomere elongation (Chandra et al., 2001). Association of Cdc13 with Est1 is essential for recruitment of the telomerase holoenzyme, which contains the protein catalytic subunit Est2 and the integral RNA template TLC1, as well as Est1 and Est3 (Lendvay et al., 1996). Cdc13 also interacts with the Stn1-Ten1 protein complex, which negatively regulates telomere elongation and plays an essential role in telomere end protection (Grandin et al., 2001). Hence, Cdc13 tightly regulates telomere elongation through its interactions with both the Est1-Est2-Est3-TLC1 complex and the competing Stn1-Ten1 complex (Chandra et al., 2001). However, the mechanism by which Cdc13 coordinates the binding of these two complexes during telomere elongation had not previously been analyzed.

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In budding yeast, the G-rich overhang is very short (about 13 bases) throughout most of the cell cycle but becomes longer around late S to G2 phases (Larrivee et al., 2004; Wellinger et al., 1993). Available data have suggested that Cdk1 activity is required for the generation of this extended 3′ single-strand overhang, although the details of the mechanism were unknown (Frank et al., 2006; Vodenicharov and Wellinger, 2006). Increased binding of Cdc13 to such an extended 3′ single-strand overhang could serve to subsequently recruit telomerase through the interaction of Cdc13 with Est1. Of the four telomerase components, Est1, Est2, TLC1, and Est3, only the expression of Est1 is cell cycle regulated, peaking at late S and G2 phases (Osterhage et al., 2006). Hence, expression of Est1 at late S and G2 phases likely restricts the assembly of functional telomerase complex to late S and G2 phases. How cells coordinate cell-cycle progression and the recruitment of telomerase complex to telomere has been an open question.

In budding yeast, the regulation of cell-cycle progression depends on a single cyclin-dependent kinase, Cdk1 (Cdc28). Cdk1 regulates cell-cycle progression by phosphorylating hundreds of different protein substrates (Ubersax et al., 2003). The association with various, periodically expressed cyclins regulates the substrate specificity of Cdk1. While it is known that telomere elongation is cell cycle dependent, no Cdk1 substrates that regulate telomere elongation have been identified. Here we show that Cdk1-dependent phosphorylation of Cdc13 at threonine 308 plays an important role in the efficient recruitment of the telomerase complex to telomeres in late G2 phases of the cell cycle. Both the telomerase complex and the Stn1-Ten1 complex are recruited to telomeres during late S and G2 phases of cell-cycle progression. Therefore, since these two complexes counteract each other in terms of telomere length regulation, it is necessary to coordinate their binding to telomeres in order to ensure active telomerase function. Our data show that phosphorylation of Cdc13 by Cdk1 plays such a key regulatory role, by coordinating the subsequent recruitment of these two complexes to telomeres to ensure proper telomere elongation and telomere protection.

RESULTS

Cdc13 Is Phosphorylated by Cdk1-as1 In Vitro
To identify potential Cdk1 substrates among components of the telomerase and telomere complexes that might contribute to the coordination of telomere elongation and cell-cycle progression in budding yeast, we utilized a previously developed strategy for specific labeling of substrates by a single kinase (Ubersax et al., 2003). In this approach, the conserved phenylalanine 88 in the ATP-binding domain of Cdk1 is replaced by glycine (Cdk1-as1). Previous data have shown that Cdk1-as1 is functional in vivo and exhibits a much higher affinity and selectivity for the bulky ATP analog N^6-Benzyl-ATP (Bishop et al., 2000; Ubersax et al., 2003). Cdk1-as1/cyclin complexes were purified from an asynchronous yeast culture (Figure 1A). Thus, the purified Cdk1-as1/cyclin complexes contain various Cdk/cyclin complexes with kinase activity for different cell-cycle stages. We chose this strategy because we did not know when any potential Cdk1 substrates from telomerase and telomere complexes might be phosphorylated in vivo. For in vitro kinase assays using this preparation, 6xHis tagged recombinant protein versions of two telomerase subunits (Est1 and Est3) and two telomere-binding factors (Cdc13 and Ten1) were used as substrates (Figure 1B). As a control for Cdk1-as1 phosphorylation specificity, we used a 6xHis-Cdc13-7A, in which alanine residues replace all seven predicted Cdk1 phosphorylation sites (as indicated in Figure S1A available online). Figure 1C shows that only wild-type Cdc13 recombinant protein is specifically phosphorylated by the Cdk1-as1 in the presence of [γ-32P]N^6-Benzyl-ATP, with only background phosphorylation detected for 6xHis-Cdc13-7A, 6xHis-Est1, 6xHis-Est3, and 6xHis-Ten1. For Est2 and Stn1, insufficient protein was obtained from the bacterial expression system. Instead, we used partially purified Est2-13myc and Stn1-13myc proteins from yeast lysates as substrates. However, no specific Cdk1-as1 phosphorylation was identified for these proteins in vitro (data not shown).

To identify the putative Cdk1 phosphorylation sites in Cdc13 phosphorylated by Cdk1-as1 in vitro, we engineered and purified 6xHis-Cdc13 recombinant proteins containing an alanine mutation at each of the seven individual potential Cdk1 phosphorylation sites in turn (as indicated in Figure S1A). As shown in Figure 1D, mutation of Cdc13 threonine 308 to alanine (T308A) dramatically reduced the phosphorylation by Cdk1-as1 in vitro. Since some residual phosphorylation of Cdc13 was seen in the presence of the single threonine 308 to alanine mutation (Figure 1D, bottom panel, compare lanes 1 and 2 to lanes 3 and 4), we further engineered and purified 6xHis-Cdc13 recombinant proteins containing the T308A mutation in combination with each of the other six additional potential Cdk1 phosphorylation site mutations. As shown in Figure 1E, when both threonine 308 and serine 336 in Cdc13 were mutated to alanine, no phosphorylation above background was observed (Figure 1E, bottom panel, compare lanes 3 and 6). Mutation of the other five putative Cdk1 phosphorylation sites in addition to the T308A mutation did not affect phosphorylation by Cdk1-as1 in vitro. These results suggested that both threonine 308 and serine 336 could be phosphorylated by Cdk1-as1 in vitro.

Phosphorylation of Cdc13 by Cdk1 In Vivo
To determine whether both Cdc13 threonine 308 and serine 336 are phosphorylated in vivo, we first asked whether mutation of either threonine 308 or serine 336 affected telomere length regulation. A diploid yeast strain (A364a background) heterozygous for the threonine 308 to alanine mutation cdc13-T308A or the serine 336 to alanine mutation cdc13-S336A was sporulated and dissected. The telomere length of each individual yeast colony derived from the dissected spores was measured after ~100 population doubling (PD100). Southern blot hybridization using a telomere probe (teloblot) showed that the cdc13-S336A mutation did not affect telomere length maintenance (Figure 2, compare lanes 1 and 2 to lanes 3 and 4). In contrast, the cdc13-T308A mutation resulted in 75 ± 10 bp telomere shortening (a 25% reduction in normal telomere length; Figure 2, compare lanes 5 and 6 to lanes 7 and 8). Such telomere shortening had largely taken place by ~50 cell divisions after sporulation (Figure S2). Similar results were observed in the S288C yeast strain background (Figure S3A). Simultaneous mutation of threonine 308 and serine 336 to alanine or mutation of all seven
putative Cdk1 phosphorylation sites to alanine caused telomere shortening similar to that of cdc13-T308A alone (Figure 2, compare lanes 9 and 10 to lanes 11 and 12; lanes 13 and 14 to lanes 15 and 16). Replacement of Cdc13 threonine 308 with aspartic or glutamic acid to provide a negative charge similar to phosphorylation failed to rescue the telomere shortening phenotype (Figure 2, compare lanes 17 and 18 to 19 and 20; lanes 21 and 22 to 23 and 24), suggesting that phosphorylation itself, rather than the negative charge associated with the phosphorylation event, is important for Cdc13 function in vivo.

Since mutation of Cdc13 threonine 308 to alanine results in significant telomere shortening in vivo, these findings suggested that Cdc13 threonine 308 may be phosphorylated and this phosphorylation event may be necessary for normal telomere maintenance in vivo. Consistent with this hypothesis, analysis of the Cdc13 amino acid sequence alignments across different fungal species in the yeast genome database shows that Cdc13 threonine 308 is well conserved, but serine 336 is not (Figure S1B).

Mutation of threonine 308 to alanine did not result in obvious changes in Cdc13 protein mobility on SDS-PAGE (data not shown). To ask whether Cdc13 threonine 308 is phosphorylated in vivo, we raised a Cdc13 threonine 308 phospho-specific rabbit

![Figure 1. Phosphorylation of Cdc13 by Cdk1-as1 In Vitro](image)

(A) A silver-stained gel showing the purification of TAP-tagged Cdk1-as1 and copurified cyclins from asynchronous yeast cell cultures.

(B) Coomassie blue stain gel showing the input of recombinant 6xHis tagged telomerase subunits and telomere-binding proteins. Cdc13-7A is used as negative control with all seven potential Cdk1 phosphorylation sites mutated to alanine. Asterisks mark proteins of expected molecular weight.

(C) Autoradiograph showing the specific phosphorylation of wild-type Cdc13 by Cdk1-as1/cyclin complexes in vitro in the presence of [\(\gamma\)-32P]N6-Benzyl ATP but not Cdc13-7A, Est1, Est3, or Ten1.

(D) In vitro phosphorylation assay of mutant Cdc13 proteins that contain mutations at individual putative Cdk1 phosphorylation site as indicated. Mutant Cdc13 with threonine 308 to alanine mutation results in dramatic reduction of phosphorylation by Cdk1-as1/cyclins in vitro.

(E) In vitro phosphorylation assay of mutant Cdc13 proteins that contain putative Cdk1 phosphorylation site mutations in addition to threonine 308 to alanine mutation. Mutation of both threonine at position 308 and serine at position 336 results in complete loss of phosphorylation by Cdk1-as1 in vitro.
Telomere shortening (lanes 5–8).

As shown in Figure 3B, lambda phosphatase treatment results in ing that this threonine 308 phospho-specific antibody is specific. (Figure 3B, top panel, compare lane 3 to lanes 1 and 2). These data detection of Cdc13 by the threonine 308 phospho-specific antibody in the presence of phosphatase inhibitor still results in specific de- while untreated lysate or lysate treated with lambda phosphatase the loss of Cdc13 detection by the phospho-specific antibody, 

Polyclonal antibody. As shown in Figure 3A, this Cdc13 threonine 308 phospho-specific antibody recognized endogenously expressed wild-type Cdc13-13myc protein immunoprecipitated from asynchronous A364a yeast cell lysate but not the Cdc13- T308A-13myc mutant (Figure 3A, top panel). Reprobing the same western blot with anti-myc antibody showed that the expression levels of Cdc13-13myc and Cdc13-T308A-13myc muta- tant proteins are comparable (Figure 3A, bottom panel), suggest- ing that this threonine 308 phospho-specific antibody is specific. As shown in Figure 3B, lambda phosphatase treatment results in the loss of Cdc13 detection by the phospho-specific antibody, while untreated lysate or lysate treated with lambda phosphatase in the presence of phosphatase inhibitor still results in specific detection of Cdc13 by the threonine 308 phospho-specific antibody (Figure 3B, top panel, compare lane 3 to lanes 1 and 2). These data confirm that Cdc13 threonine 308 is phosphorylated in vivo.

To confirm that Cdc13 threonine 308 is phosphorylated by Cdk1, it is important to show that the phosphorylation of threo- nine 308 in vivo is dependent on Cdk1 activity. Previous studies have shown that in a yeast strain carrying the Cdk1-as1 allele, the activity of CDK can be inhibited by 5–25 μM of 1-NM-PP1 within 10 min (Bishop et al., 2000). This rapid inhibition of Cdk1-as1 activity by 1-NM-PP1 reduces the potential for indirect effects due to shift of cell-cycle position. As shown in Figure 3C, addition of 10 μM 1-NM-PP1 to asynchronous yeast culture at 30°C results in rapid loss of Cdc13 detection by the threonine 308 phospho-specific antibody within 15 min (Figure 3C, top panel, compare lane 1 to lanes 2 and 3). These data suggest that Cdc13 threonine 308 is phosphorylated by Cdk1 in vivo. Treatment of the asynchronous yeast culture with 1-NM-PP1 also results in some enrichment of yeast in G1 phase of the cell cycle, as shown by FACS analysis of cellular DNA content (Figure 3D, compare 0 min to 15 and 30 min time points).

To identify the specific cell-cycle stages during which Cdc13 is phosphorylated, alpha-factor synchronized yeast cultures were analyzed in a series of time points every 15 min. As shown in Figure 3E, phosphorylation of Cdc13 can be readily detected in the time span from 45 min to 75 min after alpha-factor release. FACS analysis of cellular DNA content suggested that the phosphory- lation of Cdc13 occurred in late S to G2 phases of the cell cycle (Figure 3F), which coincides with the ability of telomerase to perform telomere elongation in vivo, as shown previously (Diede and Gottschling, 1999; Marcard et al., 2000; Schramke et al., 2004; Taggart et al., 2002). Consistent with these data obtained in the A364a yeast strain background, phosphorylated Cdc13 could also be detected in asynchronous S288C yeast lysates but not from alpha-factor arrested S288C cell lysate (Figure S3B).

Cdc13 Phosphorylation Affects Telomerase-Dependent Telomere Maintenance

To further understand the functional significance of Cdk1- dependent phosphorylation of Cdc13, we tested whether the phosphorylation of Cdc13 threonine 308 is involved in telomere- dependent or telomerase-independent telomere maintenance pathways. For this purpose, we compared the telomere length in the cdc13-T308A single mutant with single mutants of known telomerase-dependent telomere maintenance factors (est2Δ, tel1Δ, mec1Δ, yku70Δ, and yku80Δ) as well as in double mutants containing cdc13-T308A in combination with est2Δ, tel1Δ, mec1Δ, yku70Δ, or yku80Δ. As shown in Figure 5A, mutation of Cdc13 threonine 308 to alanine did not result in additive or synergistic telomere shortening phenotypes with est2Δ, tel1Δ, mec1Δ, yku70Δ, or yku80Δ deletions, indicating that Cdk1-dependent phosphorylation of Cdc13 likely plays a role in the telomerase-dependent telomere maintenance mechanism. As predicted from these findings, no accelerated senescence phenotype was observed in cdc13-T308A est2Δ or cdc13-T308A tel1Δ double mutants compared to est2Δ or tel1Δ single mutants (Figure S5).

Cdc13 Phosphorylation Is Necessary for Efficient Telomerase Recruitment to the Telomeres

Several functional domains have been identified in Cdc13, as shown in Figure S6A. Previous data from yeast two-hybrid
analysis also identified several protein interaction regions in Cdc13, including the N-terminal region (amino acids 1–252) that is necessary for interaction with Pol1, Imp4, Sir4, and Zds2 (Hsu et al., 2004), the Est1-interacting region (recruitment domain, RD) from amino acids 190–340, and the Stn1-interacting region from amino acids 252–924 (Figure S6A) (Petreaca et al., 2006). To understand the molecular function of Cdc13 phosphorylation, we first tested whether the mutation of Cdc13 threonine 308 to alanine affects the recruitment of Cdc13 to telomeres during cell-cycle progression. Chromatin immunoprecipitation (ChIP) and Q-PCR analysis (Figure 4A) showed that wild-type Cdc13 is recruited to telomeres in a cell-cycle-dependent manner, as previously reported (Schramke et al., 2004; Taggart et al., 2002). Mutation of threonine 308 to alanine did not affect this cell-cycle-dependent recruitment of Cdc13 to telomeres (Figures 4A and S7). To understand the molecular function of Cdc13 phosphorylation, we first tested whether the mutation of Cdc13 threonine 308 to alanine affects the interaction of Cdc13 to telomeres during cell-cycle progression. Chromatin immunoprecipitation (ChIP) and Q-PCR analysis (Figure 4A) showed that wild-type Cdc13 is recruited to telomeres in a cell-cycle-dependent manner, as previously reported (Schramke et al., 2004; Taggart et al., 2002). Mutation of threonine 308 to alanine did not affect this cell-cycle-dependent recruitment of Cdc13 to telomeres (Figures 4A and S7). These results are consistent with the location of threonine 308 being outside the Cdc13 DNA-binding motif (Figure S6A). Mutation of threonine 308 to alanine also did not significantly affect the cell-cycle-regulated generation of single-stranded G-rich overhangs (Figure S8). Although the single-stranded overhang signal in CDC13 yeast appears slightly stronger than that in cdc13-T308A yeast during S/G2, this is likely due to the fact that the CDC13 yeast is more efficient in elongating telomeres (during S/G2 phase) and hence elongating the G-rich strand more.

Since threonine 308 lies in both the Est1- and the Stn1-interacting regions (Figure S6A), we next asked whether Cdk1-dependent phosphorylation of Cdc13 plays a role in coordinating the interaction of Cdc13 with Est1 and Stn1. As previously reported, we could not detect the interaction of endogenous Cdc13 with Est1 or Stn1 by coimmunoprecipitation (Chandra et al., 2001; Petreaca et al., 2007). However, when 13myc-Est1 or 13myc-Stn1 was overexpressed using a galactose-inducible promoter, we readily detected the interaction of 13myc-Est1 or 13myc-Stn1 with endogenously expressed Cdc13-3HA. Mutation of threonine 308 to alanine did not abolish the interaction of Cdc13 with Est1 or Stn1 (Figure 4B, top panel, compare lanes 3 and 4, lanes 5 and 6), although the interaction of Cdc13-3HA with 13myc-Est1 seems to be slightly stronger than that of Cdc13-T308A-3HA (Figure 4B, compare lanes 3 and 4 on top panel). To address whether T308A mutation affects the interaction of Cdc13 and Est1 in a quantitative way under physiological conditions, we took advantage of the fact that the interaction of Cdc13 and Est1 is necessary for the telomeric recruitment of telomerase holoenzyme (which contains the integral RNA template component, TLC1) by Cdc13. By using QRT-PCR to analyze the amount of TLC1 that coimmunoprecipitated with Cdc13, we were able to address whether phosphorylation of Cdc13 affects its interaction with Est1 quantitatively. Indeed, mutation of threonine 308 to alanine...
results in 30% reduction of TLC1 coimmunoprecipitated with Cdc13 but not of the control actin mRNA (Figures 4C and S9A). The recruitment of Est1 to telomeres was also correspondingly reduced by at least 25% in the cdc13-T308A mutant as shown by ChIP from unsynchronized yeast culture (Figure 4D).

We further demonstrated that cell-cycle-dependent recruitment of Est1 and Est2 to telomeres is also reduced in yeast harboring the Cdc13 threonine 308 to alanine mutation compared to wild-type (Figures 4E, 4F, and S10). Together, these data indicated that Cdk1-dependent phosphorylation of Cdc13 is necessary for maximal recruitment of the telomerase complex to telomeres during cell-cycle progression but is not absolutely required.

In S. cerevisiae, the double-stranded telomeric DNA-binding protein Rap1 associates with Rif1 and Rif2 through its C-terminal domain and has been shown to negatively regulate telomere lengthening through a potential Rap1/Rif1/Rif2 counting mechanism (Levy and Blackburn, 2004; Marcand et al., 1997). In yeast strains lacking the Rap1 C terminus, Rif1, or Rif2, telomere length homeostasis is disabled and long telomeres ensue (Levy and Blackburn, 2004; Marcand et al., 1997). Reduction of Rap1, Rif1, and Rif2 association with telomeres—similar to the situation when an individual telomere is shortened—results in telomeres being elongated. Such telomere elongation induced by rap1Δ, rif1Δ, or rif2Δ is telomerase dependent (Marcand et al., 1997). To ask whether Cdk1-dependent phosphorylation of Cdc13 is important for coordination of this increased telomerase action during cell-cycle progression in yeast lacking Rif1 or Rif2, we first compared the telomere length of haploid strains obtained via dissection of sporulation products from cdc13-T308A/rif1Δ or cdc13-T308A/rif2Δ diploid heterozygotes. As shown in Figure 5A, the telomere overelongation induced by the absence of Rif1 or Rif2 depended on Cdk1-dependent phosphorylation of Cdc13. This impairment in telomere overelongation in cdc13-T308A rif1Δ or cdc13-T308A rif2Δ yeast was attributable to lower telomerase recruitment by the mutant Cdc13-T308A compared to wild-type Cdc13, as indicated by reduced association of Cdc13-T308A with TLC1 in coimmunoprecipitation experiments (Figures 5B and S9A). Consistent with the long telomere length in rif1Δ or rif2Δ cells harboring wild-type Cdc13, a strong increase of telomerase recruitment by Cdc13 was observed, as indicated by increased coimmunoprecipitation of Cdc13 and TLC1 in cdc13-T308A yeast compared to wild-type yeast expressing Est1-13myc (p values < 0.01).

Figure 4. Cdc13 Threonine 308 Phosphorylation Is Necessary for Efficient Recruitment of Telomerase Complex to Telomere

(A) ChIP assays show that Cdc13 threonine 308 mutation does not affect binding of Cdc13 to telomeres during cell-cycle progression.

(B) Cdc13-T308A still interacts with Est1 and Stn1 when Est1 and Stn1 are overexpressed.

(C) Coimmunoprecipitation of Cdc13 and TLC1 in vivo. Reduced telomerase recruitment efficiency by Cdc13-T308A as indicated by reduced association of Cdc13-T308A and TLC1 (p values < 0.01), quantified by real-time QRT-PCR.

(D) ChIP assays show that recruitment of Est1 to telomeres is reduced in cdc13-T308A yeast compared to wild-type yeast expressing Est1-13myc (p values < 0.01).

(E) ChIP assays show that cell-cycle-dependent recruitment of Est1 to telomeres is reduced in cdc13-T308A yeast compared to wild-type yeast expressing Est1-13myc (p values < 0.01).

(F) ChIP assays show that cell-cycle-dependent recruitment of Est2 to telomeres is reduced in cdc13-T308A yeast compared to wild-type yeast expressing Est2-13myc (p values < 0.01).
enrichment using a standard Y^1 subtelomeric PCR primer set (data not shown). However, ChIP clearly showed increased Est1 recruitment to telomeres in CDC13 rif2Δ yeast, which was reduced in cdc13-T308A rif2Δ yeast (Figure 5C). Together these data suggested that phosphorylation of Cdc13 threonine 308 plays a central role in coordinating telomerase recruitment to telomeres during telomere elongation in yeast lacking Rif1 or Rif2.

**Stn1-ΔC199 Rescues Cdc13-T308A Deficiency**

Since Cdc13 threonine 308 is also located inside the Stn1 interaction domain (Figure S6A), we tested whether Cdc13 phosphorylation affects the recruitment of Stn1 to telomeres. ChIP analysis showed that cell-cycle-dependent recruitment of Stn1-13myc to telomeres in CDC13 rif2Δ yeast, which was generally similar in CDC13 or cdc13-T308A cells. The Stn1 telomere association peaked in late S to G2 phases, coincident with the recruitment of the telomerase complex during cell-cycle progression, as shown previously (Schramke et al., 2004; Taggart et al., 2002). Because both the telomerase complexes and the Stn1-Ten1 complexes are recruited to telomeres during late S and G2 phases of cell-cycle progression and potentially counteract each other in terms of telomere length regulation, we asked whether the phosphorylation of Cdc13 plays a role in coordinating its interaction with these two distinct complexes. Yeast lacking wild-type Stn1 and expressing Stn1-ΔC199 (which has the C terminus necessary for its interaction with Cdc13 deleted, Figure S6B) are viable but show dramatic telomere lengthening (Petreaca et al., 2006), as shown in Figure 6B. Deletion of RAD52, which is required for homologous DNA recombination events in yeast, did not affect telomere elongation in stn1-ΔC199 strain, indicating that telomere overelongation in stn1-ΔC199 strains is due to an effect on telomerase (Figure 6B, compare lanes 1 and 2 to lanes 5 and 6). Therefore, we tested whether the mutation of Cdc13 threonine 308 to alanine affected telomere elongation in stn1-ΔC199 yeast. In striking contrast to rif1Δ or rif2Δ yeast, loss of Cdc13 phosphorylation did not affect telomere lengthening in stn1-ΔC199 yeast (Figure 6B, compare lanes 5 and 6). Consistent with this observation, stn1-ΔC199 also rescued the inhibition of telomerase recruitment by the cdc13-T308A mutation, as indicated by dramatically increased and similar association of Cdc13 and Tel1 complexes at telomeres in cdc13-T308A stn1-ΔC199 yeast.
Cdc13-T308A with TLC1 (Figures 6C and S9A). The increased associations of Cdc13 and Cdc13-T308A with TLC1 are also commensurate with the degree of telomere elongation in stn1-DC199 yeast, compared to wild-type yeast (Figure S9A). The expression level of endogenous Cdc13 or Cdc13-T308A was not affected by Stn1-DC199 (Figure S9B). Hence, these data provide evidence that interaction of Stn1 and Cdc13 is responsible for the inhibition of telomerase recruitment to telomeres that we observed in cdc13-T308A yeast. This is likely due to the increased competition of Stn1-Ten1 complex with telomerase complex for binding to Cdc13 in the absence of Cdk1-dependent phosphorylation. Since it is very difficult to accurately quantify the recruitment of Est1 or Est2 to telomeres using ChIP in stn1-DC199 yeast, due to the extremely long telomeres, it was still possible that the Cdc13-T308A mutation may affect the binding of Stn1 or Est1 to Cdc13 indirectly. To further address these possibilities, we showed that overexpression of Stn1 in cdc13-T308A yeast resulted in further telomere shortening (Figure 6D, lanes 7 and 8). In contrast, overexpression of Est1 in yeast with wild-type CDC13 resulted in slight telomere elongation (Figure 6E, lanes 2 and 6). Overexpression of Est1 in yeast with cdc13-T308A also resulted in slight telomere elongation but not complete rescue of the telomere shortening attributable to the cdc13-T308A mutation (Figure 6E, lanes 4 and 8). In summary, these data indicate that phosphorylation of Cdc13 by Cdk1 plays a central role in regulating the access of the telomerase complex to telomeres via competition with the Stn1-Ten1 complex during cell-cycle progression.

**DISCUSSION**

We have shown that Cdc13 is phosphorylated by Cdk1 in vivo during late S to G2 phases of the cell cycle, a time in the cell cycle that coincides with the previously reported recruitment of the telomerase complex to telomeres (Schramke et al., 2004; Taggart et al., 2002). Our data provide evidence that phosphorylation of Cdc13 is important in regulating the recruitment efficiency of the telomerase complex to telomeres by competing with the Stn1-Ten1 complexes during telomere elongation. Uncoupling of this cell-cycle-dependent event by inactivation of Cdc13 phosphorylation results in less telomerase recruitment to telomeres during cell-cycle progression and compromised telomere elongation.
To put these findings in context, we note that Cdk1 phosphorylation of Cdc13 at T308 will, in turn, need to be coordinated with other levels of Cdc13 regulation. In addition to Cdk1, the Tel1 and Mec1 kinases (homologs of mammalian ATM and ATR) have previously been shown to play important roles in telomere length maintenance. Tel1 and Mec1 control the DNA-damage response in budding yeast through phosphorylation of proteins involved in checkpoint control (such as Rad53) and DNA replication (such as RPA) (Brush et al., 1996; Sanchez et al., 1996). In budding yeast, in contrast to Mec1, Tel1 plays a minor role in the cellular response to DNA damage but a major role in telomere length maintenance. Loss of Tel1 results in telomere shortening (Lustig and Petes, 1986). While tel1 mec1 double mutants have a senescence phenotype (Ritchie et al., 1999), they have normal telomerase enzymatic activity (Chan et al., 2001). Consistent with this observation, Tel1 and Mec1 are required for normal levels of association of Est1 and Est2 with telomeres, suggesting a potential role in promoting the recruitment of telomerase complex to telomeres (Goudsouzian et al., 2006). Recent studies also showed that Tel1 kinase activity is crucial for preferential recruitment of telomerase to a shortened telomere (Chang et al., 2007; Hector et al., 2007; Hirano and Sugimoto, 2007; Sabourin et al., 2007). Furthermore, Tel1 and Mec1 can phosphorylate Cdc13 at multiple sites in vitro. Single mutation of either of these in vitro phosphorylation sites (serine 249 and serine 255) to alanine resulted in telomere shortening, and double mutation of these two potential phosphorylation sites resulted in progressive telomere shortening and senescence in vivo (Tseng et al., 2006).

While combination of Cdc13 T308A with S249A and S255A mutations did not result in obvious synergistic telomere shortening (Figure S11), it is still conceivable that phosphorylation of Cdc13 by Cdk1 and by Tel1 and/or Mec1 may act synergistically to determine which telomere is elongated through preferential recruitment of a functional telomerase complex in late S to G2 phases. Further identification of potential Tel1/Mec1-dependent phosphorylation sites in Cdc13 and confirmation of these Tel1/Mec1 phosphorylation events in vivo using phospho-specific antibodies will be important to probe their potential roles in telomerase action on telomeres.

Based on the results reported here and on previous data, we propose the following model: cell-cycle progression into late S phase results in telomere end processing and elongation of the G-strand overhang. Cdk1 phosphorylates Cdc13 at T308. As depicted in Figure 7, the threonine 308 phosphorylated Cdc13 on the G-strand overhang results in preferential recruitment of the telomerase complex through its interaction with Est1 because the T308 phosphorylation biases the binding of Cdc13 toward Est1. Conversely, lack of T308 phosphorylation results in reduced interaction of Cdc13 and Est1, hence increasing the association of Cdc13 with the Stn1-Ten1 complex. Thus, this Cdk1-dependent phosphorylation of Cdc13 provides a mechanism that coordinates cell-cycle progression with the recruitment of telomerase relative to the Stn1-Ten1 complexes to telomeres in late S to G2. Deletion of the Stn1 C terminus results in loss of Cdc13 binding, hence alleviating negative telomere length regulation by the Stn1-Ten1 complex and resulting in

Figure 7. A Schematic Model for Potential Function of Cdc13 Threonine 308 Phosphorylation

WT

T308A

Stn1 ΔC199

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overelongation of telomeres. The results using the stn1-JC199 mutant also argue against the cdc13-T308A mutation affecting the processing of single-strand G-rich overhangs. If cdc13-T308A caused an end processing phenotype, we should have seen a telomere length difference between CDC13 and cdc13-T308A yeast strains in the stn1-JC199 background. In addition to Cdk1-dependent phosphorylation, we suggest that the preferential recruitment of Tel1 to short telomeres and subsequent phosphorylation of Cdc13 (at serine 249 and serine 255 or other unidentified phosphorylation sites) by Tel1/Mec1 on the short telomere may preferentially synergize with the cdk1 phosphorylation to promote the recruitment of telomerase complex (likely through preferential binding to Est1) to that short telomere in a cell-cycle-dependent manner (Figure 7).

Previous data have shown that loss of Stn1 results in significant telomere elongation, while Stn1 overexpression results in telomere shortening (Chandra et al., 2001; Dahlised et al., 2003; Grossi et al., 2004). It was therefore proposed that the interaction of the Stn1-Ten1 complex with Cdc13 could compete with the recruitment of telomerase complex by Cdc13, thereby inhibiting telomerase-dependent telomere elongation (Chandra et al., 2001; Pennock et al., 2001). Consistent with this hypothesis, we found that the telomere shortening induced by loss of Cdk1-dependent phosphorylation of Cdc13 is fully alleviated in stn1-JC199 yeast, in which the interaction of Stn1 and Cdc13 is disrupted. Hence, since Cdc13 can no longer interact with Stn1, phosphorylation of Cdc13 threonine 308 is no longer necessary to promote the competing interaction with Est1. Using ChiP analysis, we also detected recruitment of Stn1 to telomeres during late S to G2 phases of the cell cycle in Cdc13 or cdc13-T308A yeast. Such cell-cycle-dependent recruitment of Stn1 to telomeres was recently reported by others (Puglisi et al., 2008). Interestingly, the magnitude of Stn1 telomere binding is independent of telomere length (Puglisi et al., 2008). Our data also suggest that the interaction of both Cdc13-3HA and Cdc13-T308A-3HA with overexpressed 13myc-Est1 is stronger than with 13myc-Stn1, so that even with the Cdc13 threonine 308 to alanine mutation, Est1 can still recruit the telomerase complex in the presence of Stn1-Ten1 competition, albeit less efficiently. This can explain why we only see a moderate telomere shortening in the presence of Stn1-Ten1 competition, albeit less efficiently. The results show an average of three experiments. Statistically significant differences in data sets were established by using a Student’s t test. The error bars represent standard deviation.

**Southern Blot Analysis and Telomere Length Measurement**

Yeast genomic DNAs were digested with XhoI, separated by 0.8% agarose gel electrophoresis, and transferred to Hybond-XL membrane (Amersham). The blot was probed for telomeric C13-A sequence with the 32P-end-labeled oligonucleotide (TGTGGTGTGGGTGTTGTTGTTG). The in-gel native hybridization was previously described (Dionne and Wellinger, 1996).

**Chromatin Immunoprecipitation**

Chromatin was isolated as previously described (Taggart et al., 2002). Primer sets were specific for amplification of 114 base pair sequence at XI-L Y’ subtelomeric sequence (TEL) and 372 base pair ARO1 sequence (ARO1, as negative control). Enrichment of telomere sequence is expressed as the fold of Y’ subtelomeric DNA immunoprecipitated per reaction from tagged strain relative to that from the nontagged strain after normalization to total input (TEL1test input/TEL1input)(TEL1no tag input/TEL1no tag). The enrichment of ARO1 mRNA was also calculated in the same way and used as control. The results show an average of three experiments. Statistically significant differences in data sets were established by using a Student’s t test. The error bars represent standard deviation.

**SUPPLEMENTAL DATA**

Supplemental Data include Supplemental Experimental Procedures, one table, and 11 figures and can be found with this article online at http://www.cell.com/S0092-8674(08)01052-X.

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