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A tolerogenic role for Toll-like receptor 9 is revealed by B-cell interaction with DNA complexes expressed on apoptotic cells

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Results

IL-10 Production by B Cells Requires Direct Contact with Whole ACs but Is Absent in B Cells Specific for Hen Egg Lysozyme. B cells secrete IL-10 in response to ACs, but only when they are able to make direct contact with whole ACs rather than with cellular debris (Fig. 1A and B). Little IL-10 protein is detectable when resting B cells are cocultured with ACs, although the majority of B cells die rapidly in culture in the absence of concomitant stimulation (28) (Fig. S1D). To address this in another way, we injected ACs into IL-10 reporter mice and 1 wk later analyzed IL-10 secretion from splenic MZB cells by FACS analysis. We found IL-10 expression in 5.15% of the MZB cells from mice that had received vehicle alone (Fig. 1A), and in 14.1% of the MZB cells in mice that had received ACs, indicating that B cells can respond to ACs in vivo in the absence of additional stimulation. In addition, short-term cocultures of MZB cells and ACs alone induced a fourfold increase in mRNA for IL-10 (Fig. 1B), again showing that B cells can respond to ACs in the absence of additional stimuli. The IL-10 protein level increased further when the B cells were activated by T cells or TLR ligands.

MZB and B1b B cells cocultured with ovalbumin (OVA)-specific T cells, OVA peptide, and ACs secreted significantly more IL-10 compared with those in cultures containing follicular B (FOB) cells (Fig. 1C and D and Fig. S1 C and D), which lack self-reactive BCRs (29). However, IL-10 secretion was not enhanced in FOBs and MZB cell populations drawn from BCR transgenic (MD4) animals carrying a single hen egg lysozyme (HEL) specificity (30) not expressed by the ACs (even when HEL was included in in vitro cultures) (Fig. 1C and D). To further clarify the role of the BCRs, we used SWHEL Ig knock-in mice that have a large proportion of B cells specific for HEL, but these can class switch, and these mice also contain populations of polyclonal B cells (31). MD4 and SWHEL mice have an increased number of MZB cells, but their B1 B cells maintain a FOB phenotype. When highly purified MZB cells from SWHEL mice were separated into polyclonal HEL− and HEL+ MZB cells, only polyclonal MZB cells produced significantly higher amounts of IL-10 in response to ACs and stimulation with the TLR ligands LPS and peptoglycan (PGN) (Fig. 1E). This suggests that MZB cells with a polyclonal BCR repertoire are required for the induction of IL-10 after the recognition of ACs. In WT mice, BiA1 B cells (i.e., CD5−) secreted more IL-10 in response to ACs compared with BiB1 B cells (CD5+) (Fig. 1F). However, in cocultures with ACs, activated splenic
CD1d<sup>hi</sup>CD5<sup>-</sup> and CD1d<sup>hi</sup>CD5<sup>-</sup> MZB cells responded similarly (Fig. 1G and Fig. S1F), suggesting that IL-10 secretion in response to AC is not restricted to a small subset of splenic B cells with defined cell surface markers.

**DNase Treatment Abolishes the Capacity of ACs to Enhance IL-10 Secretion and Inhibits Regulatory Responses in Vivo.** After apoptosis, chromatin complexes containing DNA are rapidly translocated to the AC surface (14). Given that recognition of CpG DNA motifs is a potent inducer of IL-10 in B cells (32), we reasoned that DNA-bearing molecular patterns on ACs are a potential candidate for recognition by Bregs. DNase treatment of ACs removed DNA from the surface of ACs (Fig. 2A and Fig. S2A) and abolished the AC-mediated enhancement of IL-10 production when these cells were used in cocultures with OVA-specific DO11.10 T cells, OVA peptide, and B cells (Fig. 2B). This effect was specific for DNA but not for RNA-containing complexes (Fig. S2B). DNase-treated ACs also offered no protection in the model of collagen-induced arthritis (CIA) in contrast to mice given untreated ACs (Fig. 2C), which was confirmed histologically (Fig. 2D). The level of protection were correlated with the amount of IL-10 secreted by spleen cells after in vitro restimulation with collagen (Fig. 2E). This provides clear evidence that DNA-bearing molecular patterns expressed on the surface of intact ACs are responsible for signaling the production of IL-10 by B cells.

**B-Cell TLR9 Signaling Is Required for Immunosuppressive Responses to ACs.** After endocytosis, DNA is sensed by several innate receptors (33). One of these, the MyD88-dependent receptor TLR9, recognizes mammalian DNA in endosomes and requires acidification of that compartment for signaling to occur (34). Chloroquine, which inhibits endosomal acidification, prevents signaling through TLR7 and TLR9 but not through TLR1, TLR2, or TLR4 (Fig. S3A). When B cells were stimulated with TLR2 (PGN) and TLR4 (LPS), chloroquine inhibited the AC-induced enhancement of IL-10 secretion (Fig. 3A). B cells deficient in TLR9 or its signaling adapter (but not in TLR2 or TLR4) also were unable to induce a significant rise in IL-10 secretion in cocultures with T cells (TLR9/MyD88-deficient DO11.10 plus pOVA) when exposed to ACs (Fig. 3B and Fig. S3B). To confirm that this result is a direct effect of TLR9 signaling on B cells, TLR9-deficient B1 cells were stimulated with TLR4 or TLR2 with and without ACs; the cells failed to respond to ACs by secreting IL-10 (Fig. 3C).

Clearly, to substantiate an immunosuppressive, regulatory role for B-cell TLR9, we needed to examine in vivo the effects of TLR9 deficiency on AC suppression of autoimmunity. We used myelin oligodendrocyte glycoprotein (MOG) peptide to induce EAE in TLR9<sup>-/-</sup> and TLR9<sup>−/−</sup>/mice, with ACs given at the time of disease induction. Here ACs were able to confer protection only in the WT mice, confirming a crucial role for TLR9 in mediating protection (Fig. 3D). Restimulation of spleen cells from the mice with EAE on day 12 demonstrated that TLR9-deficient mice made significantly less IL-10, but more proinflammatory IL-6 and IL-17 (Fig. S3C). Importantly, when TLR9-deficient mice were injected with WT B cells (but not TLR9<sup>−/−</sup>/B cells) along with the ACs at the time of EAE induction, the protective phenotype of ACs was restored (Fig. 3D and Fig. S3D).

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**Fig. 1.** Regulatory B cells from the MZB cell and B1a cell subsets recognize whole ACs via the BCRs and secrete IL-10. (A) IL-10-GFP reporter mice were injected with 20 × 10<sup>6</sup> ACs, and MZB cells were harvested 1 wk later. IL-10 expression was determined by FACS analysis. (B) MZB cells were stimulated with ACs for up to 4 h in the absence of further stimulation and mRNA for IL-10 measured by quantitative PCR. (C) B cells from WT or transgenic B cells from mice specific for HEL (MD4) were separated into FOB and MZB cells. After stimulation for 72 h in the presence of OVA-specific transgenic T cells and OVA peptide with or without ACs, supernatants were collected and IL-10 measured by ELISA. (D) MZB cells from MD4 and WT mice stimulated as in C, but with the cognate protein HEL added to some of the cultures. (E) Polyclonal MZB cells (HEL<sup>+</sup> and HEL-specific MZB cells (HEL<sup>−</sup>) derived from the same SW<sub>HEL</sub> mice were stimulated with the TLR4 ligand LPS or the TLR2 ligand PGN for 72 h with and without ACs, and IL-10 was measured. *Indicates a significant difference between IL-10 in stimulated B cells cocultured with ACs and HEL<sup>+</sup> and HEL<sup>−</sup> MZB cells. (F) Peritoneal-derived (PEC) CD5<sup>−</sup> and CD5<sup>+</sup> CD19 B cells and splenic MZB and FOB cells were stimulated as in C. (G) MZB cells were sorted into CD1d<sup>hi</sup>CD5<sup>-</sup> or CD1d<sup>hi</sup>CD5<sup>-</sup>, <sup>-</sup> subsets and stimulated for 72 h in C (OVA) or with various TLRs, including PGN (TLR2), and PGN (TLR7), and IL-10 was measured after 72 h. Each figure is representative of at least three experiments performed with three mice per group. Error bars represent SEM. ***p < 0.0004; **p < 0.004; *p < 0.04. The threshold for detection of IL-10 was 25 pg/mL.
B cells secreted significantly more IL-10 than those from mice treated with DNase (p < 0.04). B cells respond to ACs by secreting IL-10, but stimulation by the ACs pretreated with DNase (DNase-AC) or untreated ACs (AC) was injected into mice at the time of immunization with type II collagen (CII) in complete Freund's adjuvant (CFA), and arthritis was assessed clinically. (D) Photomicrographs of H&E-stained sections through the knee joints of mice taken from Fig. 3C. Both the DNase-treated mice and the PBS-treated mice exhibited active synovitis associated with a fibroinopurulent exudate within the joint space, with less inflammation in the AC-treated mice. (E) Splenocytes from DNase-treated mice (filled squares), DNase-treated mice (open squares), and WT mice (filled circles) obtained at the end of the experiment shown in Fig. 3C were restimulated in vitro with CII for 3 d, and IL-10 levels were measured. The CIA experiment is representative of two experiments with eight mice per group. The remaining figures are representative of at least three experiments performed with three mice per group. Error bars represent SEM. **P < 0.0004; ***P < 0.0004; *P < 0.04.

Human B Cells Generate an IL-10 Response After Interaction with ACs That Is Prevented by DNase Treatment. Finally, we asked whether human B cells also secreted IL-10 in response to DNA-bearing molecular patterns on ACs. We cocultured peripheral blood B cells (derived from healthy volunteers) with ACs and IL-4 to prevent B-cell apoptosis (28). CD27+ B cells secreted significantly more IL-10 (Fig. 4A). This increase did not occur when chloroquine (Fig. 4C) or DNase (Fig. S4A) was included in the cocultures, suggesting that both human and mouse Bregs are dependent on DNA-containing complexes expressed on ACs to secrete IL-10.

Discussion

Healthy mice and humans exhibit significant autoreactivity to self-antigens expressed on ACs, especially within the MZB and B1 B-cell repertoires (12, 13, 35, 36). The mechanism by which tolerancy is maintained in these peripheral B-cell populations has remained obscure, although these cells are able to contact and be selected by intracellular antigens expressed by ACs (37). Likewise, the mechanism by which regulatory function can be imparted to B cells by interaction with ACs is not known. In this paper, by way of explanation, we find that DNA-containing complexes on the surface of whole ACs are sensed by Bregs and delivered to TLR9-containing endosomes, where they induce IL-10 secretion. This provides a means for tolerizing self-reactive B and T cells, as well as modulating the severity of ongoing immune responses. For full-memory T-cell responses to occur, T cells must interact with both dendritic cells (DCs) and antigen-specific B cells (38, 39). Indeed, the role of B cells in shaping the T effector response through antigen presentation, costimulation, and cytokine production is being increasingly recognized (40). However, DCs that ingest infected ACs are able to up-regulate costimulatory molecules and to effectively present antigens derived from the ACs (41). Therefore, after an infectious insult, DCs are at risk of activating self-reactive T cells. Of note, our data clearly show that Bregs secrete IL-10 in response to ACs despite the presence of activating stimuli, and thus they are able to mediate a dominant tolerogenic signal despite an inflammatory milieu, which should ensure that self-reactive T cells that have been primed by DCs are induced to become regulatory IL-10–secreting cells when they contact Bregs.

Our finding that HEL-specific MZB cells from MD4 and SWHEL mice are refractory to the regulatory effects of ACs leads us to speculate that self-reactive BCRs are responsible for the recognition and uptake of these chromatin complexes. Both of these subsets are replete with self-reactive BCRs (42, 43) and AC DNA, which is more hypomethylated than DNA from viable cells (44), can activate TLR9 (45), even though TLR9’s endosomal location was previously thought to prevent its “accidental” stimulation by self-DNA (46). Despite this, the regulatory responses of B cells to ACs were ablated after treatment with chloroquine, which is known to prevent TLR7 or TLR9 signaling in the endosomal compartment, but not signaling through other TLRs (Fig. S34). Although CpG is able to gain access to the B cells independent of the BCRs and can induce B cells to secrete cytokines and proliferate (32), recent studies have demonstrated that B cells cannot take up longer lengths of DNA found physiologically, unless they are initially internalized through the BCRs (47). This will prevent large-scale activation of TLR9+ memory B cells in vivo and restrict this activation to self-reactive B cells. In fact, ligation of BCRs has been shown to control the subcellular distribution of TLR9+ in B cells, allowing relocation of TLR9 from the endoplasmic reticulum to the endosomal compartment, where interaction with antigen internalized through the BCRs can occur (48, 49). This again adds substance to the hypothesis that regulatory responses to AC-expressed chromatin complexes occurs via the BCRs.

Human CD27+ B cells respond to ACs by secreting IL-10, but only in the presence of DNA-containing chromatin complexes on the ACs, which also may help maintain peripheral tolerance in humans. The conclusion that TLR9 is involved in preventing the development of potentially damaging autoreactive responses fits with the observations from mouse disease models. Treatment of mice with CpG to stimulate TLR9 has alleviated disease severity in colitis, arthritis, and diabetes (50–53). In addition, lupus-related renal disease is exacerbated in TLR9-deficient autoimmune prone mice (20–23), and recent reports suggest that TLR9 is required to prevent pathological responses that result from TLR7-mediated signaling (24, 25). Other studies, however, show that TLR9 stimulation has an adjuvant effect driving Th1 responses allied with IgG2a antibodies, thereby potentially exacerbating autoimmune disease (19, 54–57). In addition, hydroxychloroquine is used to good effect in patients with systemic lupus erythematosus and rheumatoid arthritis, which suggests that in the rheumatic diseases, autoreactive B cells, which do not have regulatory activity, dominate the immune response.

Thus, it seems likely that TLR9 can mediate both proinflammatory and immunoregulatory signals, depending on the context in which the DNA is sensed. What factors might lead to changes in the balance between regulatory and inflammatory responses? The experiments that ascribe an inflammatory response of AC-derived DNA to TLR9 (19, 54, 55), in which DNA-antibody (IgG2a) complexes are taken up by rheumatoid factor-expressing B cells, may represent one of the situations that leads to autoimmunity, disease, for instance, when one or more of these low-affinity B cells receives signals that allow affinity maturation and drive antibody
B cells respond to ACs by secreting IL-10. The threshold for detection of IL-10 was 25 pg/mL. CD27 and CD27 is representative of at least three experiments performed with three mice per group. Error bars represent SEM. ***P ≤ 0.0004; **P ≤ 0.004; *P ≤ 0.04. The threshold for detection of IL-10 was 25 pg/mL.

Materials and Methods

Cell Stimulation and Treatments. Cells were treated with the following: DNase, 50 μg/mL (Roche); Rnase, 10 μg/mL (Sigma-Aldrich); chloroquine, 2 μg/mL (Sigma-Aldrich); TLR1/2 (PAM3CSK4), 0.2 μg/mL (InVivoGen); TLR4 ligand LPS, 2 μg/mL (Sigma-Aldrich); TLR3 ligand (poly I:C), 25 μg/mL (InVivoGen); TLR2 ligand peptidoglycan, 10 μg/mL (InVivoGen); TLR7 ligand R848, 0.1 μg/mL (InVivoGen); and TLR9 ligand CpG (ODN 1826) (InVivoGen), 1 μg/mL except in one experiment in which a dose of 25 μg/mL was used (Fig. S1D). Biotinylated HEL protein was used at 5.8 μg/mL.

Fig. 3. B-cell TLR9 signaling is required for the immunosuppressive response to ACs. (A) B cells were stimulated with the TLR ligands PGN (TLR2) and LPS (TLR4) alone or in the presence of ACs or ACs and chloroquine (AC Chloro) for 72 h, after which IL-10 in the supernatants was measured. (B) WT, MyD88-deficient (MyD88), and TLR9-deficient (TLR9) B cells were cocultured with ACs, OVA-specific T cells, and OVA peptide in vitro. After 72 h, IL-10 secretion was measured by ELISA. (C) PEC B1 cells were stimulated with the same TLR ligands as in Fig. 1A with and without AC for 72 h, and IL-10 was measured. (D) WT and TLR9-deficient mice were immunized with MOG/CFA, and EAE was scored. A single i.v. injection of ACs also was administered on day 0. Some TLR9-deficient mice also received an injection of 10 × 10⁶ CD19 B cells on day 0. The EAE experiment is representative of three experiments with five mice per group. The remaining figures are representative of at least three experiments performed with three mice per group. Error bars represent SEM. ***P ≤ 0.0004; **P ≤ 0.004; *P ≤ 0.04. The threshold for detection of IL-10 was 25 pg/mL.

Fig. 4. Human CD27⁺ B cells respond to ACs by secreting IL-10. (A) Human B cells isolated from healthy volunteers were separated into CD27⁻ and CD27⁺ fractions and cultured in the presence of IL-4 for 3 d with AC (+AC) or without AC, after which IL-10 in the supernatants was measured. (B) B cells were stimulated with ACs and IL-4 in the presence of increasing concentrations of chloroquine for 3 d, and IL-10 secretion was measured by ELISA. A shows data collected from 14 healthy volunteer blood donors; B is representative of at least two separate experiments. Error bars represent SEM. ***P ≤ 0.0004; **P ≤ 0.004; *P ≤ 0.04. The threshold for detection of IL-10 was 25 pg/mL.
Mice. DO.11.10 TcR transgenic mice (H2A−restricted, OVA peptide 323−339-specific) (63), MD4 HEL-specific BCR transgenic mice (30), IL−10−GFP mice (64), TLR2−/− mice, (56) and MyD88−/− mice (65) were bred and maintained under specific pathogen-free conditions in the animal facilities at the University of Edinburgh. The IL−10−GFP mice were kindly provided by Dr Richard Flavell (Yale University, New Haven, CT), and the TLR2−/−, TLR4−/−, TLR9−/−, and MyD88−/− mice were all generously provided by Prof. S. Akira (Hyogo College of Medicine, Nishinomiya, Japan). The SWtH mice were a kind gift from Dr. Robert Brink (Garvan Institute, Darlinghurst, NSW, Australia) (31). Male DBA1 mice were purchased from Harlan, and C57BL6 and BALB/c mice were bred in house. Mice were used at 8−12 wk of age and were age- and sex-matched in experiments. All experiments were covered by a project license granted by the Home Office under the Animal (Scientific Procedures) Act of 1986. Locally, this license was approved by the University of Edinburgh’s Ethical Review Committee.

Cell Isolation and Culture. Human mononuclear cells were extracted from the peripheral blood of healthy volunteers or from blood donor buffy coats using dextran sedimentation and a Percoll gradient as described previously (66) with Lothian Research Ethics Committee approval (LREC/2001/4/456). CD19+ B cells were isolated in accordance with the manufacturer’s instructions using a negative selection kit (Miltenyi Biotech). Viable CD19+ B cells were further sorted into CD27−, CD27+, IgM−, and IgM+ subsets using a FACSaria cell sorter (BD Biosciences) to generate highly purified populations. Human B cells (3 × 107) were cocultured with ACs (1 × 106) in the presence of 10 ng/mL of IL-4 for 72 h in RPMI medium supplemented with FCS. CD4+ T cells and CD19+ B cells from single-cell suspensions of spleen or lymph nodes were separated using CD4 and CD19 microbeads, respectively (Miltenyi Biotech), in accordance with the manufacturer’s instructions. In some experiments, viable CD19+ B cells were further separated into FOB and MZB B cells after staining with anti-CD21 and anti-CD23 and sorting with a FACSAria. For experiments with SWtH mice, viable CD19+CD23−CD21− B cells (MZB cells) were further stained with biotinylated HEL protein and sorted to high purity to yield HEL− and HEL+ specific B cells.

Cytokine Quantification. Single-cell spleen suspensions were cultured at ≥5 × 104 cells/mL with serial dilutions of CI, OVA peptide (OVA233−266, Albachem), or MyD88+/- mice in volume of 200 μL/cell in round-bottom plates (Costar; Corning, Living Sciences). For T- and B-cell cocultures, T and B cells were sorted to >98% purity using anti-CD4 and anti-CD19 magnetic beads (Miltenyi), and then ≥2 × 106 B cells were pulsed with peptide and incubated with 1 × 105 DO.11.10 CD4+ T cells. Cytokines were quantitated using standard ELISA (R&D Systems). Transwell plates were purchased from Corning Life Sciences. All experiments were performed in triplicate. Using the optimized kits (R&D Systems), the threshold for detection was 25 pg/mL for IL-10, 7.8 pg/mL for IL-6, and 3.9 pg/mL for IL-17.

Generation of ACs. Apoptotic mouse thymocytes were generated by incubating cells for 4 h with 0.5 μM dexamethasone, followed by extensive washing. For in vivo experiments, ACs were injected i.v. For in vitro cultures, ACs were added to B cells at a 5:1 ratio and then left in the wells for the duration of the assay.

Induction of CIA. CIA was induced as described previously (8). The first signs of arthritis appeared between day 21 and day 35, with a prevalence of 60−90% of immunized mice. Each arthritis-positive murine articular immune infiltrate was classified in accordance with the treatment regimen as 0, normal; 1, erythema or swelling in a single digit; 2, erythema and swelling in two or more joints; or 3, swelling of whole paw, including the hock joint. The scores of all four joints were added together, and the sum was taken as a measure of the degree of arthritis, with a maximum possible score of 12 (i.e., 4 × 3). For groups, the mean of this score was calculated.

Induction and Assessment of EAE. EAE was induced by s.c. injection of 100 μg of MOG peptide (35−55) (MEGVGWRFSKRVSGLNLIGM) emulsified in CFA containing 500 μg of heat-killed Mycobacterium tuberculosis H37RA (Sigma-Aldrich). Mice were given 200 ng of pertussis toxin (Spewey Pharmaceuticals) on days 0 and 2, and 20 × 106 AcS i.v. on day 0. Clinical signs of EAE were assessed daily on a scale of 0 to 6: 0, no signs; 1, flaccid tail; 2, impaired righting reflex and/or gait; 3, partial hindlimb paralytic; 4, total hindlimb paralytic; 5, hindlimb paralysis with partial forelimb paralysis; 6, moribund or dead.

Histology. Hindlimbs were prepared as described previously (16). Sections were analyzed blind by a histopathologist (D.S.).

Statistics. Data are expressed, when appropriate, as mean ± SEM. Significance was assessed using unpaired t tests, and P values <0.04 are considered significant.

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