Abnormal positioning of diencephalic cell types in neocortical tissue in the dorsal telencephalon of mice lacking functional Gli3

Citation for published version:

Digital Object Identifier (DOI):
10.1523/JNEUROSCI.2673-06.2006

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
Journal of Neuroscience

Publisher Rights Statement:
After six months distributed under the Creative Commons Attribution-Noncommercial-Share Alike 3.0 Unported License. Copyright © 2013 by the Society for Neuroscience

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Abnormal Positioning of Diencephalic Cell Types in Neocortical Tissue in the Dorsal Telencephalon of Mice Lacking Functional Gli3

Vassiliki Fotaki, Tian Yu,* Paulette A. Zaki,* John O. Mason, and David J. Price

Genes and Development Group, Biomedical Sciences, The University of Edinburgh, Edinburgh EH8 9XD, United Kingdom

The transcription factor Gli3 (glioma-associated oncogene homolog) is essential for normal development of the mammalian forebrain. One extreme requirement for Gli3 is at the dorsomedial telencephalon, which does not form in Gli3\(^{Xt/Xt}\) mutant mice lacking functional Gli3. In this study, we analyzed expression of Gli3 in the wild-type telencephalon and observed a high dorsal-to-low ventral gradient of Gli3 expression and predominance of the cleaved form of the Gli3 protein dorsally. This graded expression correlates with the severe dorsal-to-mild ventral telencephalic phenotype observed in Gli3\(^{Xt/Xt}\) mice. We characterized the abnormal joining of the telencephalon to the diencephalon and defined the medial limit of the dorsal telencephalon in Gli3\(^{Xt/Xt}\) mice early in corticogenesis. Based on this analysis, we concluded that some of the abnormal expression of ventral telencephalic markers previously described as being in the dorsal telencephalon is, in fact, expression in adjacent diencephalic tissue, which expresses many of the same genes that mark the ventral telencephalon. We observed occasional cells with diencephalic character in the Foxg1 (forkhead box)-expressing alon is, in fact, expression in adjacent diencephalic tissue, which expresses many of the same genes that mark the ventral telencephalon.

Key words: Gli3; mutant; development; telencephalon; diencephalon; eminentia thalami

Introduction

Gli3 (glioma-associated oncogene homolog), a zinc finger transcription factor (Ruppert et al., 1990), is an important component of the Sonic hedgehog (Shh) signaling pathway in mammals that resembles the hedgehog (Hh) signaling pathway in Drosophila (Ingham and McMahon, 2001). In the absence of Hh signal, cubitus interruptus (Ci), the fly homolog of mammalian Gli proteins (Hui et al., 1994), is cleaved to yield a transcriptional repressor, whereas in the presence of Hh, cleavage is repressed and the full-length isoform of Ci acts as a transcriptional activator (Aza-Blanc et al., 1997; Methot and Basler, 1999). It has been shown that Shh can similarly regulate Gli3 (von Mering and Basler, 1999; Aza-Blanc et al., 2000). Insight into the function of Gli3 in vivo has been gained with the study of the extratoes (Xt) mouse mutant, which has a 51.5 kb deletion in the Gli3 gene that includes the zinc-finger domain and is presumed to render it nonfunctional (Hui and Joyner, 1993; Maynard et al., 2002). Mice homozygous for the Xt\(^{f}\) mutation (Gli3\(^{Xt/Xt}\) mice) die perinatally with multiple phenotypic defects, including polydactyly and a high incidence of exencephaly, whereas non-exencephalic embryos display severe telencephalic abnormalities (Grove et al., 1998; Theil et al., 1999; Tole et al., 2000; Kuschel et al., 2003; Theil, 2005).

The telencephalic phenotype of the Gli3\(^{Xt/Xt}\) mutant includes a reduction in the size of the dorsal telencephalon, absence of olfactory bulbs, failure of the medial wall of the dorsal telencephalon to invaginate, and absence of the choroid plexus in the lateral ventricles (Hui and Joyner, 1993; Grove et al., 1998; Theil et al., 1999; Tole et al., 2000). Recently, Gli3 has been implicated in the maintenance of a proper laminar organization of the neocortex, as well as the apical/basal cell polarity of cortical precursors (Theil, 2005).

Several studies have reported ectopic expression of ventral telencephalic markers, such as Isl1, Dlx2 (distal-less homeobox), and Mash1 (mammalian achaete-schute homolog), in the dorsal telencephalon of the Gli3\(^{Xt/Xt}\) mutant (Tole et al., 2000; Rallu et al., 2002; Kuschel et al., 2003). However, the lack of...
dorsomedial telencephalon in these mice would result in an abnormal joining of the remaining dorsal telencephalon (neocortex) to the diencephalon, and previous studies might not have taken this abnormal forebrain anatomy into account when interpreting alterations in gene expression.

In this study, we performed a detailed analysis of the embryonic day 12.5 (E12.5) Gli3<sup>−/−</sup> forebrain and propose that some of the previously described ectopic dorsal expression of ventral markers in the Gli3<sup>−/−</sup> telencephalon actually reflects relatively normal gene expression in the diencephalon. We then focused on the development of the mutant neocortex. We present evidence that the telencephalic–diencephalic border in the E12.5 Gli3<sup>−/−</sup> mutants is compromised and that neocortical progenitors interspersed with diencephalic cells subsequently segregate into well-organized rosettes. Finally, we analyzed Gli3<sup>−/−</sup> forebrain younger than E12.5 and traced the likely origin of the clusters of misplaced diencephalic cells in the mutant neocortex to the presence of occasional cells of diencephalic character in the mutant dorsal telencephalon at E10.5.

Materials and Methods

**Animals.** Animal care was according to institutional guidelines. Gli3<sup>−/−</sup> CBA mice were mated, and the morning of the vaginal plug was defined as E0.5. Embryos were genotyped by PCR, as described previously (Maynard et al., 2002), fixed in 4% paraformaldehyde, and processed into paraffin blocks.

**Bromodeoxyuridine injections, immunohistochemistry, and immunofluorescence.** A 30 min pulse of bromodeoxyuridine (BrDU) (70 μg/g body weight, i.p.) was administered to pregnant dams, and E13.5 embryos were collected.

Sections were cut serially at 10 μm and reacted using standard protocols. Antigen retrieval was achieved by microwaving sections in 10 mM sodium citrate buffer. Mouse monoclonal antibodies were against BrdU (1:200; BD Biosciences, Oxford, UK), Mash1 (1:100; BD Biosciences), β-tubulin III (1:500; Sigma; Poole, UK), reelin (1:1000; Chemicon, Harrow, UK), Isl1/2 (1:50; Developmental Studies Hybridoma Bank, University of Iowa, Iowa City, IA), Lim1 and Lim2 (Lim is the three gene products Lin-11/Isl-1/Mec-3) (1:200); Developmental Studies Hybridoma Bank), nestin (1:100; Developmental Studies Hybridoma Bank), and Pax6 (paired box gene) (1:400; Developmental Studies Hybridoma Bank), Rabbit polyclonals were against calretinin and calbindin (1:2000; Swant, Bellizona, Switzerland), Foxg1 (forkhead box) (1:500; a gift from Y. Sasai, Kyoto University, Kyoto, Japan), pan-Dlx (1:100; a gift from J. Kozhtz, Northwestern University, Chicago, IL), Lhx2 (Lim homeobox) and Lhx9 (1:5000; a gift from T. Jessell, Columbia University, New York, NY), and Tbr2 (T-box brain gene) (1:200; a gift from R. Hevner, University of Washington, Seattle, WA). Binding of appropriate secondary antibodies (1:200; Vector Laboratories, Peterborough, UK) was revealed with the avidin–biotin–peroxidase system (Vector Laboratories) and diaminobenzidine (Sigma). For immunofluorescence, secondary antibodies were Alexa-488-conjugated goat anti-mouse or Alexa-568-conjugated goat anti-rabbit (Invitrogen, Paisley, UK). Fluorescent images were taken using a Leica (Nussloch, Germany) TCS NT confocal microscope. At least two to three wild-type and Gli3<sup>−/−</sup> embryos were used for each of the above antibodies.

In situ hybridization. In situ hybridizations on paraffin sections were performed as described by Nieto et al. (1996). A 611 bp fragment comprising nucleotides 560–1170 of the mouse Gli3 cDNA (a gift from T. Theil, Heinrich-Heine-University, Dusseldorf, Germany) was PCR amplified and subcloned into a pGEM-T Easy Vector (Promega, Southamp- ton, UK). The plasmid was linearized with Spel and transcribed with T7 RNA polymerase. A plasmid containing the 5’32P-labeled (a gift from J. Rubenstein, University of California, San Francisco, San Francisco, CA) was linearized with HindIII and transcribed with T3 RNA polymerase. RNA antisense probes were labeled using the digoxigenin RNA labeling kit (Roche, Welwyn Garden City, UK) according to the instructions of the manufacturer.

Western blotting. Protein was extracted from whole heads and from dissections of dorsal and ventral telencephalon of E12.5 wild-type and mutant embryos using standard methods. Tissues were homogenized with protease inhibitors and then lysed, sonicated, and boiled. Aliquots of each sample were used for protein quantification with the Pierce (Cramlington, UK) BCA protein assay, according to the instructions of the manufacturer. Equivalent amounts of protein were subjected to SDS-PAGE on a 3–8% gradient Tris-acetate gel (Invitrogen), and protein was blotted onto nitrocellulose membrane, which was incubated with rabbit polyclonal anti-Gli3 antibody (1:100; Santa Cruz Biotechnology, Heidelberg, Germany). After incubating with an HRP-conjugated anti-rabbit IgG secondary antibody (1:2000; DakoCytomation, High Wycombe, UK), signal was detected using ECL Plus detection (Amersham Biosciences, Little Chalfont, UK) according to the instructions of the manufacturer. Band intensity was measured using a densitometer and the Quantity One-4.0.3 software (Bio-Rad, Hemel Hempstead, UK).

Results

Gli3 expression in wild-type forebrain

We examined the expression of Gli3 mRNA in the mouse forebrain at midgestational stages. In situ hybridization showed that, in the telencephalon, Gli3 was expressed in the ventricular zone of the developing neocortex and dorsomedial telencephalon (Fig. 1A,B). No expression was detected in the choroid plexus (Fig. 1A,B), in agreement with previous studies (Grove et al., 1998). In
the ventral telencephalon, there was a high lateral-to-low medial gradient of expression of Gli3 through the ventricular zone of the lateral ganglionic eminence and medial ganglionic eminence (Fig. 1B). Gli3 was also expressed in the diencephalic ventricular zone, in a high-to-low gradient from epithalamus through dorsal thalamus to ventral thalamus (Fig. 1A) and in the hypothalamus at the level of the optic chiasm (Fig. 1B). Gli3 protein exists in two forms, a long 170 kDa full-length isoform and an 80 kDa isoform formed by cleavage of the full-length product (Aza-Blanc et al., 1997; Dai et al., 1999; Wang et al., 2000). To examine the spatial distribution of these two isoforms, we performed Western blots using an antibody against the N-terminus of Gli3. The results showed bands at ~170 and 80 kDa, in extracts from wild-type dorsal and ventral telencephalon that were absent in Gli3<sup>Xt/Xt</sup> tissue (Fig. 1C) and correspond to the previously described full-length and cleaved isoforms, respectively (Aza-Blanc et al., 1997; Dai et al., 1999; Wang et al., 2000). The antibody also detected non-specific bands of intermediate size and unknown identity, whose intensities varied in proportion to the total amount of protein loaded in each lane (Fig. 1, arrow). Total levels of Gli3 protein were approximately twofold higher in the dorsal compared with the ventral telencephalon, reflecting the pattern of Gli3 mRNA expression (Fig. 1C). We then quantified the ratio between the full-length and cleaved forms in these tissues. Dorsally, the cleaved form was present at 2.75 ± 0.45 times (mean ± SD; n = 7) the concentration of the full-length form, whereas ventrally the ratio was lower, at 1.33 ± 0.50 (mean ± SD; n = 8) and the difference between these two ratios was significant (Student’s t test; p < 0.0005). Overall, the highest concentration of Gli3 was of the cleaved form in the dorsal telencephalon (Fig. 1C).

**Identification of the limits of the dorsal telencephalon in Gli3<sup>Xt/Xt</sup> mutants**

We performed a detailed rostrocaudal morphological comparison of E12.5 wild-type and Gli3<sup>Xt/Xt</sup> forebrain sections immunolabeled with Pax6, which is highly expressed in the dorsal telencephalon and diencephalon (Walther and Gruss, 1991; Stoykova and Gruss, 1994; Mastick et al., 1997), and Mash1, which is expressed in the ventral telencephalon and diencephalon (Lo et al., 1991; Guillemot and Joyner, 1993; Porteus et al., 1994). Figure 2 depicts representative sets of wild-type and mutant forebrain sections in a caudal-to-rostral order. The most caudal diencephalic parts of the mutant forebrain (epithalamus, dorsal and ventral thalamus) appear relatively normal both anatomically and in their expression of Pax6 and Mash1 (Fig. 2A, A’, B, B’, E, E’, F, F’), although the ventricular space appears slightly enlarged in mutants. At a level in which caudal telencephalic lobes are clearly visible in wild-type sections (Fig. 2B, F), only the most caudal tips of the telencephalic lobes are observed in corresponding Gli3<sup>Xt/Xt</sup> sections (Fig. 2B’, F’). This may be because of the diminished size of the Gli3<sup>Xt/Xt</sup>lobes compared with wild type and/or their altered position. Moving rostrally, the dorsomedial telencephalon and choroid plexus, seen in wild types in Figure 2, C, D, G, and H, are absent in Gli3<sup>Xt/Xt</sup>embryos, and the connection between the third and the lateral ventricle is greatly enlarged (Fig. 2C’, D’, G’, H’). The intense labeling for Pax6 in the ventral thalamus of mutant embryos (Fig. 2B’) is continuous with strong staining for Pax6 in the neocortex (Fig. 2C’), because Pax6 low-expressing tissue that
would normally intervene (the dorsomedial telencephalon) (Fig. 2C) is missing in the mutant. In wild-type sections, Mash1 is expressed in the ganglionic eminences (Fig. 2G, H) and in the ventral thalamus (Fig. 2F, G) but is not observed in the neocortex (Fig. 2G, H, area of tissue between the lines). In comparable sections in the Gli3Xt/Xt mutant, Mash1 immunoreactivity is also present in the ganglionic eminences (Fig. 2G/H11032, H11032) and absent from the adjacent Pax6-positive area (Fig. 2, compare C/H11032, G/H11032). This suggests that the Mash1-negative region (Fig. 2G/H11032, H11032, area between the lines) is neocortical tissue, and the adjacent dorsal region of intense Mash1 staining most likely corresponds to diencephalic tissue [ventral thalamus and/or eminentia thalami (Fig. 2G/H11032, labeled VT&EmT)].

To confirm that Mash1-expressing tissue adjacent to the neocortex is diencephalic and not telencephalic in origin in Gli3Xt/Xt embryos, we performed immunohistochemistry using an antibody against Foxg1. Foxg1 is expressed in the dorsal and ventral telencephalon and is absent from the dorsal and ventral diencephalon (Tao and Lai, 1992; Hanashima et al., 2002). As shown in Figure 3, A and A’, Foxg1 expression is found in the telencephalon of both wild-type and Gli3Xt/Xt embryos and is absent from the adjacent Mash1-positive tissue in the mutant (Fig. 2G’, labeled VT&EmT), confirming that this tissue is not diencephalic. These results clearly delineate the neocortical region and confirm that the Mash1-positive region dorsal to the Foxg1-positive neocortical region is diencephalic.
To provide additional evidence for this, we examined the pattern of expression of Islet1 and Dlx in Gli3Xt/Xt E12.5 embryos (Fig. 3B’,C’). These genes are normally expressed in the ventral telencephalon and the ventral thalamus (Fig. 3B, C), and previous reports have suggested that they are ectopically expressed in the dorsal telencephalon of the Gli3Xt/Xt mutant (Tole et al., 2000; Rallu et al., 2002; Kuschel et al., 2003). As in wild types, the expression of Islet1 and Dlx is confined to (1) the ventral telencephalon and (2) the ventral thalamus and/or eminentia thalami region in Gli3Xt/Xt embryos (Fig. 3B’,C’). That this latter region corresponds to the mutant equivalent of ventral thalamus and/or eminentia thalami is further supported by the expression pattern of Shh transcript in the zona limitans intrathalamica (ZLI) (Echelard et al., 1993; Marti et al., 1995), which defines the border between the dorsal and ventral thalami (Fidgert and Stern, 1993; Rubenstein et al., 1994; Kiecker and Lumsden, 2004). Shh mRNA expression is present in both wild-type and Gli3Xt/Xt sections (Fig. 3D,D’), showing clearly that the Islet1, Dlx-positive region just below the ZLI corresponds to ventral thalamus in both wild types and mutants. Furthermore, the region that corresponds to the neocortex (Fig. 3B’,C’, area of tissue between the lines) does not express either Islet1 or Dlx. Altogether, our results show that the previously described ectopic expression of Mash1, Islet1, and Dlx in the dorsal telencephalon of Gli3Xt/Xt mutants (Tole et al., 2000; Rallu et al., 2002; Kuschel et al., 2003) is actually expression in the diencephalon (ventral thalamus and/or eminentia thalami).

We examined Pax6 and Mash1 expression patterns during later stages of Gli3Xt/Xt forebrain development by immunohistochemistry on caudal-to-rostral sets of sections from E13.5 to E15.5. At most levels, Pax6 immunoreactivity was confined mainly to the dorsal telencephalon and ventral thalamus, and Mash1 immunoreactivity was confined to the ventral telencephalon and ventral thalamus in both wild types and Gli3Xt/Xt mutants (Fig. 4A,A’,B,B’), similar to results at E12.5. Results for both Mash1 and Pax6 expression in E14.5 and E15.5 wild-type and Gli3Xt/Xt tissue were also similar (data not shown), although their interpretation at these stages became more complex because of the high degree of disorganization of the putative Gli3Xt/Xt neocortex (see below).

The Gli3Xt/Xt neocortex contains clusters of cells with characteristics of the eminentia thalami

High-power images of Foxg1 immunostaining at E12.5 revealed patchy expression of Foxg1 in the Gli3Xt/Xt neocortex (Fig. 5A’), which was not observed in the wild type (Fig. 5A). A similar patchy staining was also observed with an antibody against Lhx2 (Fig. 5B’), which is normally expressed throughout the neocortex in a high-dorsal-to-low-lateral gradient (Fig. 5B) (Monuki et al., 2001).

We examined the nature of the Foxg1-negative cells observed in the Gli3Xt/Xt neocortical area. Results described above suggest their possible identity. They are immunonegative for Mash1, Islet1, and Dlx (Figs. 2G’,H’, 3B’,C’) (supplemental data, available at www.jneurosci.org as supplemental material), indicating that they do not share properties with neuronal progenitors or postmitotic cells from the ventral telencephalon and/or ventral thalamus. However, the tissue negative for Foxg1, Mash1, Islet1, and Dlx is positive for Pax6 (Fig. 2C’,D’) (supplemental data, available at www.jneurosci.org as supplemental material). Pax6 is expressed not only in the dorsal telencephalon and ventral thalamus but also in the ventricular zone of the eminentia thalami (Fig. 2C) (Puelles et al., 2000), which forms part of the rostral boundary between the diencephalon and the telencephalon (Rubenstein et al., 1994; Puelles and Rubenstein, 2003). We examined whether the Foxg1-immunonegative patches could have an eminentia thalami identity and used as a marker calretinin, which labels eminentia thalami postmitotic cell somata and fibers in wild-type mice (Fig. 5C,D) (supplemental data, available at www.jneurosci.org as supplemental material) (Abbott and Jacobsen, 2003). As in wild types, the expression of Foxg1, Lim1/2, Calretinin, and Lhx2 was confined to the neocortex (Fig. 5D,G’). Double immunofluorescence with Foxg1 (red) and Lim2 (green) labels the neocortical region and eminentia thalami, respectively, in the wild type (F). In the mutant, Foxg1-immunonegative patches are immunopositive for Lim2 (F’). Pax2 immunostaining reveals the presence of a small population of positive cells at the most dorsolateral tip of the eminentia thalami (EmT) of both wild types (C, D) and mutants (C’, arrowhead in D’). In the mutants, calretinin-positive clusters localize in the vicinity of the Foxg1-immunonegative patches (compare A’, C’). Similarly, Lim2 is expressed in the eminentia thalami in both wild type (E) and mutant (E’) and reveals the presence of Lim2-positive clusters in the vicinity of the Foxg1-immunonegative patches in the mutant (E’). Double immunofluorescence with Foxg1 (red) and Lim2 (green) labels the neocortical region and eminentia thalami, respectively, in the wild type (F). In the mutant, Foxg1-immunonegative patches are immunopositive for Lim2 (F’). Pax2 immunostaining reveals the presence of a small population of positive cells at the most dorsolateral tip of the eminentia thalami, close to the choroid plexus (ChP) in the wild type (G). In the mutant, a similar Pax2-immunopositive population is observed in a region that is Lim2 positive and Foxg1 negative (arrow). However, additional Pax2-positive cells are observed among the Foxg1-immunonegative patches (arrowheads). C and C’ are magnifications of the boxed areas in B and B’, respectively. Sections in C, C’, D, and D’ are counterstained with cresyl violet. In F and F’, rostral is to the left. Scale bars: A’, A’, B’, C’, 50 μm; B, E, E’, F’, G, G’, 100 μm; C, F, 150 μm; D, D’, 400 μm. dmT, Dorsomedial telencephalon; GE, ganglionic eminentia.

Figure 5. Dorsal telencephalic and eminentia thalami marker analysis in wild-type (Gli3Xt/Xt) (A–G) and Gli3 mutant (Gli3Xt/Xt) (A’–G’) coronal (A–E, G, and A’–E’, G’) and sagittal (F, F’) sections at E12.5. Foxg1 immunolabeling reveals the presence of patches lacking Foxg1 staining in the mutants (A’) that are never observed in corresponding wild-type sections (A). Lhx2 immunolabeling, which presents a high-dorsal-to-low-lateral gradient in the wild-type neocortex (nctx) (B), reveals the presence of immunonegative patches in the mutant neocortex (B’). Calretinin expression is detected in the eminentia thalami (EmT) of both wild types (C, D) and mutants (C’, arrowhead in D’). In the mutants, calretinin-positive clusters localize in the vicinity of the Foxg1-immunonegative patches (compare A’, C’). Similarly, Lim2 is expressed in the eminentia thalami in both wild type (E) and mutant (E’) and reveals the presence of Lim2-positive clusters in the vicinity of the Foxg1-immunonegative patches in the mutant (E’). Double immunofluorescence with Foxg1 (red) and Lim2 (green) labels the neocortical region and eminentia thalami, respectively, in the wild type (F). In the mutant, Foxg1-immunonegative patches are immunopositive for Lim2 (F’). Pax2 immunostaining reveals the presence of a small population of positive cells at the most dorsolateral tip of the eminentia thalami, close to the choroid plexus (ChP) in the wild type (G). In the mutant, a similar Pax2-immunopositive population is observed in a region that is Lim2 positive and Foxg1 negative (arrow). However, additional Pax2-positive cells are observed among the Foxg1-immunonegative patches (arrowheads). C and C’ are magnifications of the boxed areas in B and B’, respectively. Sections in C, C’, D, and D’ are counterstained with cresyl violet. In F and F’, rostral is to the left. Scale bars: A’, A’, B’, C’, 50 μm; B, E, E’, F’, G, G’, 100 μm; C, F, 150 μm; D, D’, 400 μm. dmT, Dorsomedial telencephalon; GE, ganglionic eminentia.
In Gli3<sup>XO/Xt</sup> mutants, the eminentia thalami region, as revealed by intense calretinin staining, lies above the dorsal limit of the neocortical area and displays a thinner postmitotic layer compared with the wild type (Fig. 5C, D<sup>′</sup>) (supplemental data, available at www.jneurosci.org as supplemental material). Adjacent to this area, small, calretinin-positive cell clusters were observed in the mutant neocortex (Fig. 5C, D<sup>′</sup>). To provide additional evidence about the eminentia thalami nature of these clusters, we examined the expression pattern of the transcription factor Lim2, also known as Lhx5, which specifically labels the eminentia thalami and ventral thalamus in E12.5 wild types (Fig. 5E) (Sheng et al., 1997). The antibody used recognizes both Lim1 (Lhx1) and Lim2 (Lhx5) proteins, but Lim1 mRNA expression is very weak in these tissues at this age (Sheng et al., 1997). In the Gli3<sup>XO/Xt</sup> mutant neocortex, Lim2 immunostaining presented a patchy expression (Fig. 5E<sup>′</sup>), similar to that observed with calretinin and complementary to the Foxg1-negative patches (Fig. 5, compare A, C, E, E<sup>′</sup>). To confirm this, we performed double immunofluorescence for Foxg1 and Lim2 in sagittal E12.5 wild-type and Gli3<sup>XO/Xt</sup> sections. In wild types, Lim2 immunostaining was confined in the eminentia thalami, whereas Foxg1 specifically labeled the dorsal and ventral telencephalon (Fig. 5F<sup>′</sup>). In Gli3<sup>XO/Xt</sup> mutant, Lim2 immunostaining was found in the Foxg1-positive region in the form of patches, and the two markers did not colocalize (Fig. 5F′). Tbr1, another marker of the eminentia thalami (Puelles et al., 2000), also revealed the presence of clusters in the vicinity of the Foxg1-immunonegative patches (data not shown).

Finally, we observed that Pax2, a well described marker of the hindbrain, optic chiasm, and optic stalk (Nornes et al., 1990; Puschel et al., 1992), labels a distinct population of eminentia thalami cells, found in close proximity to the choroid plexus in wild-type E12.5 sections (Fig. 5G). In the Gli3<sup>XO/Xt</sup> mutants, Pax2 immunostaining revealed dispersed patchy-like expression (Fig. 5G′) that was similar to that revealed with the calretinin and Lim1/2 antibodies. Similar results were also obtained in a sagittal plane for all of the eminentia thalami markers examined (calretinin, Tbr1, and Pax2) (data not shown).

The above results strongly support an eminentia thalami identity for the cell clusters observed in the neocortical region of the Gli3<sup>XO/Xt</sup> mutants. However, because calretinin, Lim2, and Tbr1 also label Cajal-Retzius cells in the marginal zone (del Rio et al., 1995; Super et al., 1998; Hevner et al., 2001, 2003; Yamazaki et al., 2004), it was possible that the calretinin-positive, Lim2-positive, and Tbr1-positive clusters in the Gli3<sup>XO/Xt</sup> neocortex comprised this cell type. To examine this possibility, we used immunostaining with reelin, which is also found in Cajal-Retzius cells (Alcantara et al., 1998), and calbindin, which is not normally observed in this cell population (Hevner et al., 2003; Jimenez et al., 2003). We did not observe any reelin-positive cell clusters in the Gli3<sup>XO/Xt</sup> mutant (Fig. 6A<sup>′</sup>, B<sup>′</sup>). In fact, the number of reelin-positive cells was significantly lower than in wild type (Fig. 6A, A<sup>′</sup>, B, B<sup>′</sup>), which is in agreement with recently published data (Theil, 2005). Calbindin immunostaining was detected in the ventral telencephalon in both wild type and mutant (Fig. 6C, C<sup>′</sup>), as described previously (Davila et al., 2005). However, it also labeled lightly eminentia thalami neurites in the wild type and mutant (Fig. 6D, D<sup>′</sup>) and was also detected in the vicinity of calretinin-positive clusters in the neocortex (Fig. 6C<sup>′</sup>, D, D<sup>′</sup>) (compare Figs. 6D<sup>′</sup>, 5C<sup>′</sup>). These results indicate that the calretinin-positive, Lim2-positive, and Tbr1-positive clusters observed among the Foxg1-negative patches are not Cajal-Retzius cells.

A few dispersed reelin-positive cells in the Gli3<sup>XO/Xt</sup> neocortex (Fig. 6B<sup>′</sup>, arrows) were found in the same region as the calretinin/calbindin-positive ectopic clusters (data not shown), suggesting that a few Cajal-Retzius cells (Fig. 6B) may contribute to these clusters of displaced eminentia thalami cells. However, it is likely that these reelin-positive cells are derived from the eminentia
thalamus, because reelin also labels a population of eminentia thalami cells (Fig. 6A).

Altogether, our results clearly show that the neocortex of Gli3<sup>XO/Xt</sup> embryos contains not only neocortical cells but also cells of eminentia thalami identity, indicating that the border between the telencephalon and diencephalon is compromised in this mutant.

Eminentia thalami clusters are first observed in the Gli3<sup>XO/Xt</sup> neocortex at E11.5

To gain insight into the developmental stage at which the eminentia thalami clusters start to form in the neocortical region of Gli3<sup>XO/Xt</sup> embryos, we studied the expression of eminentia thalami markers in sagittal sections at E11.5 (Fig. 7). Sections through wild types (Fig. 7A–C) are at, or close to, the plane marked (I) in supplemental data A (available at www.jneurosci.org as supplemental material); sections through mutants are at the planes marked in supplemental data A′ (available at www.jneurosci.org as supplemental material) as either (II) (Fig. 7A′–C′) or (III) (Fig. 7A″–C″). E11.5 is the stage at which wild-type calretinin expression is first observed in the eminentia thalami (Abbott and Jacobowitz, 1999). In wild types, the eminentia thalami lacks Foxg1 (Fig. 7A), expresses calretinin in its differentiating cells (Fig. 7B) and Pax2 in a collection of cells close to the choroid plexus (Fig. 7C), and is located caudal to the dorsomedial telencephalon, which does not express Foxg1 (Fig. 7A) (Dou et al., 1999). In mutants, the Foxg1 nonexpressing eminentia thalami is directly caudal to the dorsal telencephalon, which is immunopositive for Foxg1 (Fig. 7A′, A″, a). As in wild types, its differentiating cells are detected by their expression of calretinin (Fig. 7B′, B″). Pax2 is expressed mainly in dorsal eminentia thalami of mutants (Fig. 7C′, C″). In Gli3<sup>XO/Xt</sup> mutants, calretinin-positive and Pax2-positive cell clusters were observed in close proximity to the eminentia thalami and within the Foxg1-positive telencephalic region (Fig. 7B′, B″, b′, c′, c″, e′, arrowheads). Clusters that were positive for both calretinin and Pax2 were more numerous in the most lateral parasagittal sections of the Gli3<sup>XO/Xt</sup> mutants (Fig. 7B′, b′, c′, c″), in which the Foxg1-immunonegative dorsal telencephalic patches are also more abundant (Fig. 7A′, a). These results show that, at E11.5, eminentia thalami cells are already present in the Gli3<sup>XO/Xt</sup> neocortex in the form of clusters.

At E10.5, the choroid plexus and eminentia thalami have not differentiated yet into distinct morphological structures (Sturrock, 1979; Abbott and Jacobowitz, 1999). In addition, markers that specifically label the eminentia thalami at later development stages are either not expressed yet (calretinin and Tbr1) or label a broader region (Lim2) (Abbott and Jacobowitz, 1999). The only informative marker for the study of the developing eminentia thalami at E10.5 was Pax2.

Sagittal sections through the telencephalic–diencephalic boundary region of wild-type E10.5 embryos showed Pax2-positive cells in a similar relative position to that at E11.5 (Fig. 8A, a). They are within a region identified as diencephalic on the basis of morphology, intense Pax6 immunoreactivity (Fig. 8A, a) (Mastick et al., 1997), and absence of Foxg1 immunoreactivity (data not shown). These data indicate that, as at E11.5, the Pax2-expressing cells are on the diencephalic side of the diencephalic–telencephalic boundary at E10.5, in close proximity to the hippocampal primordium, which is immunonegative for Foxg1 (Dou et al., 1999).

In the E10.5 mutant, Pax2-positive cells are mainly within the diencephalic region, which presents intense Pax6 immunostaining similar to that observed in the wild type (Fig. 8A′, a′). This tissue is joined directly to Foxg1/Pax6-immunopositive tissue corresponding to the Gli3<sup>XO/Xt</sup> dorsal telencephalon (Fig. 8A″, a″), because the intervening Foxg1-immunonegative hippocampal primordium is absent in the Gli3<sup>XO/Xt</sup> embryos, in accordance with published data (Grove et al., 1998; Theil et al., 1999; Tole et al., 2000). A few, isolated Pax2-positive cells were observed in the Foxg1/Pax6-positive dorsal telencephalon (Fig. 8A′, a′, a″, a‴, arrowheads, B‴, asterisk). These results show that, at E10.5, individual Pax2-immunopositive cells from the Gli3<sup>XO/Xt</sup> diencephalon are present ectopically in the adjacent dorsal telencephalon of the mutants.

In the diencephalon, we observe a mixed population of diencephalic cells that either express Pax2 but not Pax6 (Fig. 8B, B‴,
Discussion

The severe reduction in size of the dorsal telencephalon and the lack of dorsomedial telencephalon in the Gli3Xt/Xt mouse have been reported (Grove et al., 1998; Theil et al., 1999; Tole et al., 2000). However, previous studies neither precisely defined the extent of the remaining dorsal telencephalon nor showed how this disrupted structure is joined to its neighboring region, the diencephalon. Our study allowed us to identify the area of the Gli3Xt/Xt forebrain corresponding to neocortical tissue (summarized in supplemental data, available at www.jneurosci.org as supplemental material). By studying the expression patterns of Foxg1, which is expressed in developing telencephalon but not diencephalon (Tao and Lai, 1992; Hanashima et al., 2002), Pax6, a well-characterized marker of the dorsal telencephalon and diencephalon (Walther and Gruss, 1991; Stoykova and Gruss, 1994; Mastick et al., 1997), and Mash1, which is expressed in both the ventral telencephalon and ventral thalamus (Lo et al., 1991; Guillemot and Joyner, 1993; Porteus et al., 1994), we were able to distinguish the dorsal and ventral limits of the residual Gli3Xt/Xt dorsal telencephalon. Based on this analysis, we demonstrated that the Mash1-, Dlx-, and Islet1-immunopositive regions found
at the dorsal end of the Gli3<sup>−/−</sup> neocortex correspond to the ventral thalamus. This is in contrast to previous studies that have reported that these ventral telencephalic markers are ectopically expressed in the dorsal telencephalon of Gli3<sup>−/−</sup> mice (Tole et al., 2000; Rallu et al., 2002; Kuscht et al., 2003). This discrepancy can be attributed to the fact that previous studies did not define the Gli3<sup>−/−</sup> telencephalic limits relative to the diencephalon and did not consider that many ventral telencephalic markers are also expressed in the developing diencephalon.

**Correspondence between the severity of the forebrain defects in Gli3<sup>−/−</sup> forebrain and the expression of Gli3**

Our expression analysis of Gli3 mRNA in the developing mouse telencephalon reveals high expression dorsally and a high/lateral-to-low/medial gradient ventrally, as described previously (Grove et al., 1998), and is in accordance with the severe phenotypic defects of the Gli3<sup>−/−</sup> mutants dorsally and the absence of gross alterations ventrally. Furthermore, it agrees with our observation that levels of Gli3 protein are higher in the dorsal than in the ventral telencephalon.

Estimates of the relative amounts of the long and short isoforms of Gli3 in the developing telencephalon showed that there is significantly more of the short than the long isoform in the dorsal telencephalon, whereas there are almost equivalent levels of both isoforms ventrally. The ratio of the cleaved to the full-length isoform of Gli3 in the dorsal and ventral telencephalon are analogous to those described in the anterior and posterior limb bud, respectively (Wang et al., 2000; Litingtung et al., 2002; Chen et al., 2004). Litingtung et al. (2002) proposed that the ratio of the two Gli3 forms is crucial for digit number and identity. Similarly, regional differences in this ratio may be important for dorsoventral patterning of the telencephalon. Relatively high expression of the cleaved Gli3 repressor form in the anterior limb domain has been correlated with absence of Shh signaling (Wang et al., 2000), in a similar manner to that described in the Drosophila anterior wing bud (Methot and Basler, 1999). In the dorsal telencephalon, absence of Shh (Sussel et al., 1999; Nery et al., 2001) may account for the high levels of the processed Gli3 isoform, which might act as a repressor of the Shh signaling pathway.

**Neocortical cells form rosettes intermingled with eminentia thalami cells in Gli3<sup>−/−</sup> mutants**

Having defined the limits of the presumptive neocortex in the E12.5 Gli3<sup>−/−</sup> mutant, we evaluated its development at later stages. Our analyses led to the observation of two striking features. First, starting at E13.5, we noticed the formation of rosettes, which consisted of cells surrounding a lumen. These became more numerous as development proceeded. They were composed of neocortical neural progenitors, and the relative position of S-phase cells around the lumen was similar to that of S-phase cells around the ventricle in wild types. Tbr2 expression in cells surrounding the outermost surface of the rosettes resembled that observed overlaying the ventricular zone in the neocortex of wild types (Englund et al., 2005). It appears that the rosettes comprise well organized neocortical progenitors that segregate from surrounding cells, many of which express markers of the eminentia thalami.

Previous work described the formation of aberrant structures in the Gli3<sup>−/−</sup> dorsal telencephalon but did not characterize the cell types involved (Theil et al., 1999). Rosette-like structures forming close to the ventricle have also been described in mice with loss-of-function mutations in the genes for the membrane-associated protein Lgl1 (lethal giant larvae homolog 1) (Klezovitch et al., 2004) and the myosin II-B heavy chain (Tullio et al.,...
2001). Their formation has been attributed to alterations in the adhesive properties of the neuroepithelial cells and in their apical–basal cell polarity, defects that have been described recently in Gli3lox/lox neocortical tissue (Theil, 2005) and may be the primary cause for the rosette formation in this mutant.

The second important observation in the E12.5 Gli3lox/lox dorsal telencephalon was the presence of patches of Foxg1-negative cells that expressed markers of the neighboring eminentia thalami among the Foxg1-positive neocortical cells. Calretinin, which labels postmitotic cells and fibers found in the eminentia thalami (Abbott and Jacobowitz, 1999), was expressed by cells in these patches that were located near the marginal zone. Additional evidence for the origin of these clusters was their expression of Lim2 and Tbr1, known markers of eminentia thalami (Sheng et al., 1997; Puelles et al., 2000), and of Pax2, a newly described marker of an eminentia thalami population found close to the choroid plexus in wild types.

In mammals, the eminentia thalami is a transitional developmental structure (Keyser, 1972), is proposed to act as an organizer for the diencephalon, and appears at approximately E11 (Abbott and Jacobowitz, 1999). It is found in the rostralmost diencephalic area (prosomere 3) (Puelles and Rubenstein, 2003) and forms part of the rostral boundary between the diencephalon and the telencephalon (Trujillo et al., 2005). We found that eminentia thalami clusters first appear among the dorsal Gli3lox/lox telencephalon at E11.5. At E10.5, when eminentia thalami cells have not yet started to differentiate and the medial walls of the dorsal telencephalon have not invaginated, we observed a few dispersed Pax2-positive cells within the Foxg1/Pax6-positive dorsal telencephalic area, close to the presumptive mutant diencephalic–telencephalic boundary. These Pax2-positive cells are most likely the precursors of the Pax2-positive clusters observed 1 day later. The most straightforward hypothesis to explain the aberrant presence of cells of eminentia thalami identity in neocortical tissue in Gli3lox/lox mutants is that the medial wall of the dorsal telencephalon normally prevents eminentia thalami cells reaching the neocortex. If the dorsomedial telencephalon is lost, as in Gli3lox/lox mutants, the lifting of this restriction on the movement of eminentia thalami cells might allow them to mix with neocortical cells. The cortical hem, a Bmp/Wnd (bone morphogenetic protein/wingless-type MMTV integration site family)-rich tissue found in the dorsomedial telencephalon, is suggested to act as a signaling center (Grove et al., 1998; Grove and Tole, 1999) and is missing in Gli3lox/lox mutants (Grove et al., 1998; Theil et al., 1999; Tole et al., 2000). This tissue might be the source of molecular signals that prevent mislocation of diencephalic cells in the telencephalon. It is interesting that cells characteristic of eminentia thalami mislocated in the dorsal telencephalon at E10.5 are not respecified to the fate of the majority of their neighbors, suggesting a strong commitment to an eminentia thalami fate. The fact that some of these Pax2-mislocated cells do not express Pax6 provides additional evidence as to their eminentia thalami origin, because Pax6 is expressed by all dorsal telencephalic precursors at this developmental stage (Walther and Gruss, 1991).

Other factors might contribute to the abnormalities described here. For example, a well studied diencephalic boundary, the ZLI, which separates the dorsal and ventral thalamus (Figdor and Stern, 1993; Rubenstein et al., 1994; Kiecker and Lumsden, 2004), is a source of Shh (Hashimoto-Torii et al., 2003; Kiecker and Lumsden, 2004) and is considered a secondary organizer of the diencephalon (Echevarria et al., 2003; Vieira et al., 2005). Although Shh expression in the presumptive ZLI is maintained in the Gli3lox/lox mutants, the characteristic wild-type shape of the ZLI, perpendicular to the main axis of the neural tube (Kiecker and Lumsden, 2004) (Fig. 3D), is altered. This change in ZLI orientation might affect the signaling properties that the ZLI experts on the developing diencephalon and might directly or indirectly influence the formation of an intact diencephalic–telencephalic boundary.

In summary, the present study challenges the concept of widespread ventralization in the dorsal telencephalon of Gli3lox/lox mutants and shows that the absence of functional Gli3 results in abnormal localization of cells of diencephalic identity in the mutant neocortical region. Our results highlight the importance of considering the relative position of the diencephalon to the telencephalon when analyzing the telencephalic defects of this mutant.

References