Altered dark- and photoconversion of phytochrome B mediate extreme light sensitivity and loss of photoreversibility of the phyB-401 mutant

Citation for published version:
https://doi.org/10.1371/journal.pone.0027250

Digital Object Identifier (DOI):
10.1371/journal.pone.0027250

Link:
Link to publication record in Edinburgh Research Explorer

Document Version:
Publisher's PDF, also known as Version of record

Published In:
PLoS ONE

Publisher Rights Statement:
This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

General rights
Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.
Altered Dark- and Photoconversion of Phytochrome B Mediate Extreme Light Sensitivity and Loss of Photoreversibility of the phyB-401 Mutant

Éva Ádám1*, Andrea Hussong2*, János Bindics2, Florian Wüst2, András Viczián1, Marcus Essing2, Mátyás Medzhiradszky2, Stefan Kircher2, Eberhard Schäfer2,3, Ferenc Nagy1,4*

1 Institute of Plant Biology, Biological Research Centre, Szeged, Hungary, 2 Institut für Biologie II/Botanik, Universität Freiburg, Freiburg, Germany, 3 Centre of Biological Signalling (BIOSS), Universität Freiburg, Freiburg, Germany, 4 School of Biological Sciences, University of Edinburgh, Edinburgh, United Kingdom

Abstract

The phyB-401 mutant is 10^3 fold more sensitive to red light than its wild-type analogue and shows loss of photoreversibility of hypocotyl growth inhibition. The phyB-401 photoreceptor displays normal spectral properties and shows almost no dark reversion when exposed in yeast cells. To gain insight into the molecular mechanism underlying this complex phenotype, we generated transgenic lines expressing the mutant and wild-type phyB in phyB-9 background. Analysis of these transgenic lines demonstrated that the mutant photoreceptor displays a reduced rate of dark-reversion but normal Pfr to Pr photoconversion in vivo and shows an altered pattern of association/dissociation with nuclear bodies compared to wild-type phyB. In addition, we show (i) an enhanced responsiveness to far-red light for hypocotyl growth inhibition and CAB2 expression and (ii) that far-red light mediated photoreversibility of red light induced responses, including inhibition of hypocotyl growth, formation of nuclear bodies and induction of CAB2 expression is reduced in these transgenic lines. We hypothesize that the incomplete photoreversibility of signalling is due to the fact that far-red light induced photocconversion of the chromophore is at least partially uncoupled from the Pfr to Pr conformation change of the protein. It follows that the phyB-401 photoreceptor retains a Pfr-like structure (Pfr*) for a few hours after the far-red light treatment. The greatly reduced rate of dark reversion and the formation of a biologically active Pfr* conformer satisfactorily explain the complex phenotype of the phyB-401 mutant and suggest that amino acid residues surrounding the position 564 G play an important role in fine-tuning phyB signalling.

Introduction

Plants are sessile organisms, which have established a considerable plasticity of development to respond to changes in the natural environment. Light, a highly variable environmental factor, is used not only as the main energy source, but also as an environmental cue to regulate plant growth and development. To monitor these changes, plants have evolved several classes of photoreceptors: the UV-B photoreceptors [1,2], the blue/UV-A sensing cryptochromes controlling plant development [3], the phototropins controlling directional growth, chloroplast re-orientation and stomata opening [4,5] and the red/far-red absorbing phytochromes controlling plant development [6,7,8].

In Arabidopsis thaliana phytochromes are encoded by a small gene family of five members, PHYA to PHIE [9]. The phytochromes in vivo exist as homo and/or heterodimers of about 125 kDa subunits [10] and each subunit contains a covalently linked, linear tetrapyrrole chromophore. The phytochromes are synthesized in their R light absorbing, biologically inactive Pfr form in darkness. Absorption of R light converts them into their FR light absorbing biologically active Pfr form that again can be converted back into Pfr form upon absorption of FR light [7].

In addition to photoconversion, the Pfr form can also fall back into the Pfr form spontaneously, since the Pfr form has lower thermostability. This light-independent conformation change of the photoreceptor, also called dark reversion, was first suggested to function as an additional inactivation process by Eichenberg et al. [11]. It has been demonstrated by these authors that dark reversion beside being an intrinsic property of the molecule is also affected by cellular components and shows strong temperature dependence [12]. Whether the molecular dynamics of dark transition from Pfr to Pfr is identical to and mediated by the same intermediate states as the light induced Pfr to Pfr transition is not yet known.

Based on their mode of action and the light stability of the proteins, phytochromes can be divided into two classes. Type I phytochromes controlling the Very Low Fluence Responses (VLFR) and the far-red High Irradiance Responses (HIR) show rapid degradation of the Pfr form. Type II phytochromes are light
stable and control Low Fluence Responses (LFR) and red light High Irradiance Responses. Using mutant analysis, it was shown that type I phytochromes are encoded by the PHA4 gene and type II phytochromes by the PHYB, PHYC, PHYD and PHYE genes [6,8,13].

Physiological experiments showed that all these phytochromes sense not only the quality but also the quantity of light. Light quality sensing is explained by the formation of different amounts of the active Pfr-form and for phyA, a light quantity sensing mechanism can also be envisaged: In this model the fast proteolytic degradation of phyA [14] and the EID1 dependent degradation of additional signal transduction components are likely to play major roles. EID1 is an F-box protein and mutations of the EID1 gene lead to light hypersensitivity and a shift of the action spectrum from the far-red peak to a red peak [15,16,17]. In the case of phyB the capacity to monitor light quantity is less obvious [18], as phyB-mediated responses cannot be provoked by a single R light pulse that establishes saturating Pr levels, whereas continuous red light elicits strong responses [19,20]. Recent observations, however, suggest that dark-reversion [21], proteolytic degradation of the active phyB Pfr form [22] and formation of phyB-containing nuclear bodies [23] are important factors in mediating light quantity measurement by this photoreceptor.

Accordingly, expression of phytochrome cDNA in yeast cells and reconstitution of the holo-photoreceptor by addition of the chromophore have shown that all tested phytochromes exhibit partial dark reversion of Pr to P, after phototransformation of P to P [24]. Recently, a slower but almost complete dark reversion of phyB has been measured in vivo, in phyB over-expressing lines in a phyA null background [21], whereas Oka et al. [25,26] reported that the N-terminal PHY domain of phyB is involved in regulating dark reversion of the photoreceptor in transgenic plants. Interestingly, it has been demonstrated [21,27] that dark reversion of phyB is at least partially regulated by its interaction with the response regulator ARR4. Thus it follows that regulation of dark reversion of phyB is likely to be a target for hormone-induced signalling pathways.

In etiolated seedlings, phyB is predominantly localized in the cytosol [28]. After transfer to light, nuclear import of phyB and formation of intra-nuclear speckles, also termed as nuclear bodies (NBs) have been described [28]. Surprisingly, these reactions show a strong light quality and quantity dependence, specific for the individual photoreceptors [20,23,28,29]. Although the precise biological function of the different types of phyB-associated NBs [29–32] is not fully understood, it is evident that the majority of phyB signalling mutants display aberrant NB formation [20,29,31]. Moreover, recent data described by Rausenberger et al. [23] suggest that, in contrast to phyB Pr localized in the nucleoplasm, the NB-associated phyB Pr conformers are prevented from fast dark-reversion.

The phyB-401 mutant isolated by Kretsch et al. [33] shows extreme hypersensitivity to red light and displays strongly reduced reversibility by far-red light. The same authors reported that the phyB-401 photoreceptor exhibits a nearly complete lack of dark reversion but normal photo-reversibility in yeast cells. The reduced dark reversion could, in principle, explain the extreme hypersensitivity to R light, but it is not sufficient to account for the poor far-red light reversibility of the inhibition of hypocotyl growth. To interpret the complex phenotype of the phyB-401 mutant at molecular level we raised transgenic lines expressing the wild-type PHYB:GFP/YFP and mutant PHYB-401:GFP/YFP fusion proteins in phyB-9 background. Next we measured (a) the rate of dark reversion and light induced photoconversion of the mutant phyB-401 and wild type phyB fusion proteins in planta and (b) analysed far-red reversibility of cellular and molecular events related to phyB action. Our data suggest that the complex phenotype of phyB-401 can be best explained by assuming that the G to E amino acid change at position 564 inhibits dark reversion and at least partially uncouples photoconversion of the chromophore from the Pr-Pr conformational change of the holoprotein.

Results

We have generated several independent transgenic lines expressing the 35S:PHYB-401:YFP and 35S:PHYB-401:GFP fusion proteins in phyB-9 background. The primary transgenic plants were selfed, homozygous segregants were selected and lines displaying stable phenotype were multiplied for additional experiments. Next, the expression level of the PHYB-401:YFP or GFP fusion proteins was determined and compared to PHYB:GFP and PHYB:YFP in phyB-9 and endogenous PHYB in WT backgrounds, respectively. The transgenic lines expressing the 35S:PHYB:GFP or YFP fusion protein used in this study had been isolated previously, they display stable phenotypes and fully complement the phyB-9 mutant [20,28]. We performed a series of western-blot hybridization experiments and selected one line for further experiments from each of the 35S:PHYB:401:YFP, 35S:PHYB:401:GFP, 35S:PHYB:GFP and 35S:PHYB:YFP lines. The selected transgenic lines over-express the fusion proteins 3–8 times as compared to endogenous phyB (for details see Table S1).

Fluence rate response analysis of hypocotyl growth inhibition

Figure 1 shows the fluence rate response curves obtained for WT (Col), phyB-9 and transgenic PHYB-401:YFP and PHYB:YFP seedlings in cR. This figure illustrates that the transgenic line expressing the PHYB-401:YFP fusion protein displays extreme hypersensitivity to cR, since these seedlings reach saturation of the response already at approximately 0.001 μmol m⁻² s⁻¹. In contrast, the transgenic PHYB:YFP and the WT (Col) seedlings start responding at ~0.01 μmol m⁻² s⁻¹ and show saturation around 1 and 10 μmol m⁻² s⁻¹, respectively. Transgenic lines expressing the PHYB-401:GFP fusion protein displayed similar hypersensitivity to R (Figure 1 and Figure S1A). We note that all seedlings expressing PHYB-401 and PHYB fusion proteins displayed normal etiolated phenotype as compared to WT seedlings (Figure S2). Taken together, these fluence rate measurements are in good agreement with the data reported by Kretsch et al. [33] and indicate that the PHYB-401:YFP and GFP fusion proteins affect R induced inhibition of hypocotyl elongation as expected.

In vivo dark reversion kinetics of PHYB-401:YFP after saturating red light treatment

Data reported by Kretsch et al. [33] demonstrated that the phyB-401 photoreceptor exhibits a reduced dark reversion in vitro. To confirm and extend this observation, we measured the dark reversion kinetics of the PHYB-401:GFP fusion protein in planta. To this end, 4-day-old seedlings were irradiated for 3 h with R (3 μmol m⁻² s⁻¹) and the amount of Pr was measured directly with a dual-wavelength ratio spectrophotometer (RatioSpec) during the following dark period. For detailed information about the RatioSpec measurement see Materials and Methods. Figure 2 shows that in vivo dark reversion of PHYB:GFP has a half life time of around 1 h, which is in good agreement with previously
published data [21,23]. As expected, dark reversion of the PHYB-401:GFP expressing line displayed a different, strongly reduced reversion kinetics (Figure 2). This figure illustrates that after 12 h of darkness about 50%, whereas after 24 h darkness about 40% of the original amount of PHYB-401:GFP Pfr is still detectable. These data unambiguously demonstrate that the PHYB-401:GFP fusion protein, like the PHYB-401 protein in vitro, shows a strongly reduced dark reversion in vivo. This in turn results in an increased stability of the PHYB-401 Pfr form, which is especially important in continuous weak red light or after light pulses. Under these conditions, where only a low percentage of PHYB Pfr/Ptot homodimers are generated [6], the enhanced stability of the Pfr form of the PHYB-401:YFP fusion protein can indeed result in an increased sensitivity, but it has little to do with the loss of reversibility response described by Kretsch et al. [33].

In vivo photoconversion of PHYB and PHYB-401 Pfr to Pr
Kretsch et al. [33] demonstrated that photoconversion of phyB-401 does not significantly differ from that of the wild type phyB in vitro. To evaluate whether this statement is valid in vivo we measured far-red light induced photoconversion of PHYB:GFP and PHYB-401:GFP fusion proteins in planta. Photoconversion of PHYB-401:GFP and PHYB:GFP Pfr to Pr was determined as follows. 4-day-old etiolated seedlings were treated with red light for 3 h (40 μmol m⁻² s⁻¹) to convert all phyB Pr into Pfr form and simultaneously to degrade phyA. During the last 15 min of red light treatment seedlings were placed on ice and afterward all additional steps were carried out on ice to avoid dark reversion of the generated phyB Pfr. After completing the red light treatment seedlings were irradiated with 6 μmol m⁻² s⁻¹ far-red light (720 nm) for 30 s, 60 s or 135 s and the Pfr and total phy levels were measured in dual-wavelength ratio spectrophotometer (Ratiospect). Figure 3 demonstrates that photoconversion of PHYB:GFP and PHYB-401:GFP Pfr to Pr is identical in vivo.

These data suggest that aberrant/impaired photoconversion of the phyB-401 Pfr to Pr cannot be accounted for the loss of far-red reversibility of phyB-401 mutant regulated responses.

Light quantity dependent nucleo-cytoplasmic partitioning of the photoreceptor:YFP fusion proteins in seedlings grown in cR
We examined the subcellular localization pattern of the PHYB-401:YFP and PHYB:YFP fusion proteins in transgenic seedlings grown for 4 days in darkness or at 0.03 μmol m⁻² s⁻¹ and 22 μmol m⁻² s⁻¹ intensities of cR. It is estimated that under...
these conditions the biologically active Pfr conformer represents ~20 and >80% of the total phyB amount, respectively (for a detailed explanation for the calculation of the Pfr/Ptot ratio see Material and Methods). PHYB:YFP displayed diffuse staining and was detectable in the nuclei and cytoplasm in dark-grown seedlings (Figure 4A). Figure 4B shows that illumination of PHYB:YFP seedlings with weak cR induces, in a low percentage of cells, the formation of a few small NBs. Irradiation with higher light intensity clearly promotes association of the PHYB:YFP fusion protein with NBs (Figure 4C). PHYB:YFP was still detectable in the cytoplasm after illumination with weak cR (Figure 4B), whereas the absence of cytoplasmic YFP signal in seedlings irradiated with high-intensity R light (Figure 4C) indicates an increased level of nuclear import of the PHYB:YFP. Intracellular distribution of PHYB:401:YFP was clearly different from that of PHYB:YFP. Figure 4E and F demonstrate that irradiations with low and/or high intensity cR uniformly induced formation of numerous large PHYB:401:YFP NBs, whereas, as shown in Figure 4D–F, cytoplasmic YFP fluorescence was detectable only in etiolated material. Taken together, these data indicate that nuclear import and formation of large PHYB:401:YFP associated NBs display increased sensitivity in cR as compared to PHYB:YFP.

**Figure 4. Subcellular distribution of PHYB:YFP and PHYB:401:YFP differs in seedlings irradiated with cR.** Localization of PHYB:YFP (A, B, C) and PHYB:401:YFP (D, E, F) fusion proteins in 4-day-old etiolated seedlings grown in darkness (A, D), low-intensity (0.03 μmol m⁻² s⁻¹) (B, E) or high-intensity cR light (22 μmol m⁻² s⁻¹) (C, F) is shown. Below the fluorescence pictures of cells (A–F) the corresponding differential interference contrast microscopic images are also shown. doi:10.1371/journal.pone.0027250.g004

Kinetics of the formation, stability and photoreversibility of R pulse induced nuclear bodies containing PHYB:401:YFP and PHYB:YFP

First we tested the effectiveness of a single R pulse to induce the formation of phyB NBs. To this end we monitored cellular distribution of PHYB:YFP and PHYB:401:YFP in 4-days-old dark-grown seedlings that had been treated with a short R pulse (30 s, 70 μmol m⁻² s⁻¹) and then returned to darkness. In etiolated seedlings both PHYB:YFP (Figure 5A, D) and PHYB:401:YFP (Figure 5B, C) showed weak, diffuse nuclear staining. The applied high-intensity R pulse induced rapid formation of numerous, small NBs associated with PHYB:YFP or PHYB:401:YFP already within 2 minutes after the treatment. These NBs were short-lived and transient, as they were not detectable later than 1 hour after the R pulse (Figure 5A and B). As for PHYB:YFP, we could not detect NBs in samples monitored 5 h, 16 h, 24 h and 48 h after the treatment: the PHYB:YFP fusion protein remained uniformly distributed in the nucleoplasm. In contrast, the PHYB:401:YFP-associated NBs reappeared in the nuclei within 5 h after the R pulse (Figure 5B). These newly formed, late NBs differed from the transient NBs detected shortly after the light treatment, i.e. they were less numerous but larger and, surprisingly, they were detectable and got even larger up to 48 h of darkness (Figure 5B). As only the Pfr conformer of phyB is known to be able to associate with NBs, these observations imply that the Pfr form of PHYB:401:YFP is more stable than that of PHYB:YFP. Furthermore the disappearance of early speckles and appearance of late NBs after 2–3 h in complete darkness, indicates that PHYB:401:YFP remains in the Pfr state in darkness for long time and that formation of PHYB containing late NBs takes several hours even in the continuous presence of Pfr.

This slow process can either be due to the slow modification of pre-existing NBs to allow interaction with PHYB Pfr or a slow modification of PHYB Pfr to facilitate interaction with preexisting NBs.

We also tested far-red light reversibility of this process. For these assays we employed a similar experimental setup, i.e. 4-day-old etiolated seedlings were treated with a short R pulse (30 s, 70 μmol m⁻² s⁻¹) directly followed by a 2 min RG9 pulse (40 μmol m⁻² s⁻¹) and transferred to darkness. We found that the RG9 treatment prevented the appearance of R-induced early, transient PHYB:YFP NBs (Figure 5D). In contrast, Figure 5C demonstrates that the formation of R-induced PHYB:401:YFP-associated NBs, early/transient as well as late/stable ones, was only partially affected by RG9. These nuclear complexes remained clearly detectable, although RG9 irradiation lowered the number of the early, transient and the newly appearing late PHYB:401:YFP NBs. As only the Pfr conformer of phyB is known to be able to associate with NBs, these experiments suggest that RG9-induced photoconversion of PHYB:401:YFP Pfr holoprotein to Pp is significantly impaired as compared to PHYB:YFP. We conclude this despite the fact that in zero spectroscopic measurements of the Pfr to Pp, photoconversion -monitoring the photoconversion of the chromophore - does not show any significant alteration (Figure 3).

Characterization of the photoreversibility of cR-induced PHYB:YFP and PHYB:401:YFP nuclear bodies

Rausenberger et al. [23] demonstrated that light treatments reverting Pfr back to the Pp form led to a fast depletion of stable phyB NBs. Thus we concluded that monitoring the depletion kinetics of cR-induced PHYB:401:YFP NBs after an RG9 pulse provides an alternative method for testing photoreversibility of these nuclear complexes. To perform these studies, 3-day-old
etiolated seedlings were irradiated with cR light (3 \(\mu\)mol m\(^{-2}\) s\(^{-1}\)) for 24 h and then either transferred to darkness, or irradiated with an additional RG9-pulse (2 min, 40 \(\mu\)mol m\(^{-2}\) s\(^{-1}\)) prior to transfer to darkness. Localization of the PHYB-401:YFP fusion protein was analyzed immediately after the light treatments and during the following dark period, up to 6 days. Figure 6 (left panel) shows that without the RG9 pulse the PHYB-401:YFP NBs are stable in darkness, since their number and size do not change significantly up to 72 h. This figure also shows that treatment with RG9 pulse did not deplete the PHYB-401:YFP NBs up to 48 h after the pulse but led to the appearance of smaller, more numerous NBs. These observations are in sharp contrast to data reported by Rausenberger et al. [23], who showed that cR-induced PHYB-YFP NBs were no longer detectable after 9 h in darkness and RG9 treatment significantly accelerated depletion of these nuclear structures that became undetectable within 1 h after the RG9 treatment. Finally, we note that the RG9-pulse applied during these assays did not induce NB formation or any physiological response in seedlings expressing the PHYB-401:YFP fusion protein. Taken together, these and the data shown in Figure 5C indicate that the RG9-generated P\(_r\) form of PHYB-401:YFP is still capable of associating with NBs, this P\(_r\) form is relatively stable and its properties resemble that of the biologically active P\(_a\) form of PHYB-YFP.
Figure 6. RG9 treatment is ineffective in depleting cR-induced PHYB-401:YFP NBs in darkness. Seedlings expressing PHYB-401:YFP or PHYB:YFP were grown for 3 days in darkness and on day 4 they were irradiated with cR (3 μmol m$^{-2}$ s$^{-1}$). After the 24 h R irradiation the seedlings were transferred to darkness (–RG9) or were treated with a RG9 pulse (40 μmol m$^{-2}$ s$^{-1}$) prior to transfer to darkness (+RG9). Subcellular localization of the PHYB-401:YFP and the PHYB:YFP fusion proteins were monitored after the light treatments. Sampling time (h) after the light treatments is indicated in the left panels. Scale bar is shown.

doi:10.1371/journal.pone.0027250.g006
Comparison of the signalling efficiency of PHYB:GFP and PHYB-401:GFP

Beside the decreased rate of dark reversion, the G to E change at position 564 may influence interaction of the mutant photoreceptor with components of the signal transduction cascade and thereby contribute to the dramatic hypersensitivity of the phyB-401 mutant and the PHYB-401:GFP seedlings as compared to WT (Figure 7). Since it has been shown that the total amount of phyB is the limiting factor for maximal signal strength [23], comparison of fluence rate responses of lines containing different levels of total phyB can provide additional information concerning the mechanism underlying the hypersensitive phenotype of the phyB-401 mutant. To achieve this, we measured fluence rate dependent inhibition of hypocotyl growth in lines containing different amounts of phyB (in phyA-201 and 35S:PHYB:GFP in phyB-9, respectively) and mutant phyB (in phyB-401/phyA-201 and 35S:PHYB-401:GFP in phyB-9 backgrounds, respectively). Over-expression levels are shown in Table S1. Figure 7 shows the relation between the amount of the photoreceptor and the maximal response/signal for the mutant phyB (in phyB-401/phyA-201 and PHYB-401:GFP in phyB-9) and wild-type phyB (in phyA-201 and PHYB:GFP in phyB-9). This figure demonstrates that under saturating light conditions (10 µmol m⁻² s⁻¹ red light) the amount of phyB or phyB-401 Pfr neither in phyA-201 nor in phyA-201/phyB-401 background is sufficient to induce maximal physiological response in contrast to the transgenic lines expressing the PHYB:GFP and PHYB-401:GFP in phyB-9 background. Because at 10 µmol m⁻² s⁻¹ fluence rate the amounts of phyB and phyB-401 and PHYB:GFP and PHYB-401:GFP Pfr are pairwise identical we conclude that the phyB-401 Pfr molecule is not more efficient than the wild-type photoreceptor.

However, the extreme hypersensitivity for red light induced physiological responses displayed by the phyA-201/phyB-401 and PHYB-401:GFP seedlings makes it difficult to assess the signalling efficiency under non-saturating conditions. To determine this by using the hypocotyl growth inhibition as a read-out we have to know the exact amount of phyB and phyB-401 Pfr present. This parameter we can estimate only with large uncertainty because of the altered dark reversion and finding shown in Figure 6 which indicates that the cycled Pfr form of phyB-401 may display biological activity. To overcome this difficulty we compared signalling efficiency of the wild type and the mutant phyB-401 photoreceptors by other experimental approaches described below.

Kinetics of signalling initiated by PHYB:YFP and PHYB-401:YFP

As RG9 pulses were shown to revert hourly R pulses in WT but not in the phyB-401 mutant [33] we tested the reversibility of daily applied R pulses. Figure 8A shows that under these conditions seedlings expressing the wild-type PHYB:YFP display a weak hypocotyl growth inhibition response to daily applied R pulses, whereas the PHYB-401:YFP seedlings show an almost saturated growth inhibition. We note that the observed strong effect of daily red light pulses is in good agreement with the reduced dark reversion kinetics of PHYB-401 and clearly demonstrates the physiological consequence of the increased stability of the Pfr form. The same figure, however, also shows that (i) the effect of daily R pulses on hypocotyl growth of the PHYB-401:YFP seedlings can be reverted (to about 50%) by an additional RG9 pulse (ii) which alone was not inductive. We also determined the kinetics of the reversibility of PHYB-401:YFP-controlled inhibition of hypocotyl growth. To this end we grew PHYB-401:YFP seedlings as described above but included 4 and 8 h dark intervals between the inductive R and the reverting RG9 pulses. Figure 8B illustrates that the PHYB-401:YFP-induced response can be partially reversed by far-red pulses applied leading to 60% and 50% reduction of hypocotyl length, respectively. These data fit well the kinetics of the loss of reversibility of wild-type phyB published recently by Hennig et al. [34]. Taken together, we conclude that (i) the kinetics of signal transduction initiated by the mutant phyB-401 photoreceptor and wild-type phyB do not differ significantly, and that (ii) the loss of photoreversibility of the R pulse induced hypocotyl growth described by Kretsch et al. [33] is not caused by an accelerated signal transduction operating in the phyB-401 seedlings.

Fluence rate dependence of light pulse induced accumulation of CAB2 mRNA

To compare the sensitivity of PHYB and PHYB-401 photoreceptors at the molecular level we tested the CAB2 mRNA accumulation after an inductive far red light pulse. 4-day-old etiolated seedlings were irradiated for 5 min with far red light of variable fluence rates to establish fluences between 104 and 13350 µmol m⁻². Figure 9 shows that these fluences were not inductive in phyA-201 but they caused significant elevation of CAB2 mRNA levels in phyA-201/phyB-401. This figure also shows that maximal induction of CAB2 mRNA was detected at 3418 µmol m⁻² fluence. These data demonstrate the hypersensitivity of the PHYB-401 because at least at the higher fluences the same Pfr levels should be established for PHYB and PHYB-401. The unexpected fluence dependence of the CAB2 mRNA induction by a far-red light pulse indicates that the photochemical cycling of the PHYB-401 photoreceptor produces an active signalling component.

Discussion

The intragenic phyB-401 point mutant reported by Kretsch et al. [33] displays extremely enhanced light sensitivity and loss of
photoreversibility of hypocotyl growth inhibition to hourly R light pulses. This phenotype is unique among the numerous phyB intragenic mutants isolated to date yet it is still unclear how the G to E amino acid substitution at position 564 modifies phyB signaling at the molecular level. Here we report about a broad array of experiments performed with PHYB-401:YFP and GFP expressing transgenics that allowed us to confirm and extend results reported by Kretsch et al. [33]. We show that the 35S:PHYB-401:YFP and GFP transgenic lines mimic the phenotype of the original phyB-401 mutant, display extreme hypersensitivity to cR (Figure 1 and Figure S1A) and to cFR (Figure S1B). We also demonstrate that the PHYB-401:YFP fusion protein has a reduced dark reversion rate in vivo (Figure 2), whereas its in vivo photoconversion rates (Figure 3), its degradation in cR (Figure S3) and signalling efficiency under saturating light conditions (Figure 7) are identical to PHYB:YFP. Importantly, we found that hypocotyl growth inhibition induced by daily applied R pulses (in contrast to hourly R pulses) is photoreversible in PHYB-401:YFP seedlings (Figure 8A), which in turn allowed us to document, by increasing the interval between the inductive R and the inhibitory RG9 pulses, that signaling launched by the mutant photoreceptor is not faster than that initiated by wild-type phyB (Figure 8B).

Monitoring light-induced changes of the nucleo/cytoplasmic distribution of PHYB and PHYB-401:YFP and/or GFP provided essential information for identifying a novel feature of the mutant photoreceptor and for suggesting a plausible explanation for the appearance of the unique phenotype. Our data clearly demonstrate that PHYB-401:YFP NBs are more stable in darkness (Figure 5B and Figure 6), RG9 light is ineffective to prevent appearance of R-induced PHYB-401:YFP NBs (Figure 5C) and/or to induce rapid depletion of PHYB-401:YFP NBs formed in cR.
photoreceptor. Under such conditions the Pr* will be accumulated
red light is mediated by photochemical cycling of the PHYB-401
applied at 24 h intervals.
a significant, about 70% reversibility when R/RG9 pulses are
hypocotyl growth inhibition of PHYB-401:YFP seedlings displays
constant after the photoequilibrium is reached. Data obtained by
be longer than one hour as hourly red light pulses are not
Pfr. In contrast to this the in vivo spectroscopic measurements show
that photoversion of the chromophore - as measured by in vivo
spectroscopy - is normal. This P* form is biologically active
(Figure 9 and Figure S1B); it persists for some hours (Figure 5B
and Figure 6) and it slowly falls back to P by a mechanism not yet
elucidated. The kinetics of this process cannot be precisely estimated, however, our data show that the life time of P* must
be longer than 8 hours. The precise calculation of the life time
is not possible as the signaling efficiency of P* compared to P is
unknown.

Our data demonstrate that such a P* signaling competent can
also be produced by far-red light pulses. The level of P* will be
elevated with increasing fluence, whereas the P level will remain
constant after the photoequilibrum is reached. Data obtained by the
mRNA induction by far-red light pulses supports the hypothesis
of the formation of a signaling P*. The hypersensitivity of inhibition
phyB-401:YFP under cFR also cannot be explained by simply altered dark reversion of the mutant
photoreceptor, because even in the absence of any dark reversion
far-red light can only produce P level up to 2–3%. (Figure S1B).
The strong fluence rate dependency of this response suggests that
this unexpected sensitivity of phyB-401 mutant to continuous far-
red light is mediated by photochemical cycling of the PHYB-401
photoreceptor. Under such conditions the P will be accumulated
in a fluence rate dependent manner.

This hypothesis and the reduced dark reversion rate of the
phyB-401 protein satisfactorily explain the extreme light sensitivity
in weak cR and the loss of reversibility of hypocotyl growth
inhibition to hourly R/FR pulses [33]. In weak cR, dark reversion
of P is the major regulator of P levels. Under this condition only
a low percentage of P/P phyB homodimers are generated [6],
thus the low dark reversion rate leads to an enhanced stability of
the PHYB-401:YFP P, which then results in an increased sensitivity. We note that dark reversion of PHYB-401:YFP could
also generate a P-like P*, conformer, which then would further enhance the probability of the formation of the presumably active
P*/P dimers. This will also contribute strongly to the extreme
efficiency of PHYB-401:YFP or GFP to very low red light fluence rates under continuous irradiation (Figure 1 and 7).

As far as the loss of photoresponsiveness of hypocotyl growth
inhibition of the phyB-401 mutant is concerned [33], formation of
the phyB-401:YFP P* (Figure 5C, Figure 6) readily explains why hourly given RG9 pulses are ineffective in blocking hourly R-induced hypocotyl growth inhibition and why the phyB-401 does respond strongly to
far-red light (Figure 9 and Figure S1B). However, we note that the proposed P* form is likely to be less stable than the P form. Data
shown in Figure 5B support this conclusion by showing that hypo-
cotyl growth inhibition of PHYB-401:YFP seedlings displays a
significant, about 70% reversibility when R/RG9 pulses are
applied at 24 h intervals.

The phenotype displayed by phyB-401 mutant is sharply
different from that of the constitutively active PHYB mutant
described by Su and Lagarias [35] and Hu et al. [36]. These
authors showed that the universally conserved GAF domain Tyr
residue, with which the chromophore is intimately associated,
performs a critical role in coupling light perception to signal
transduction. In other words, this particular amino acid substit-
ution confers a conformation that mimics the photoactivated P* form of phyB in the absence of light in a chromophore-dependent
manner. Consequently, the PHYB protein mutant displays de-
ciated phenotype in dark and fluorescence independent
hypocotyl growth inhibition in light, whereas the PHYB-401:
seedlings display normal etiolated phenotype (Figure S2).

It is of interest to understand at the structural level why the G to
E amino acid change at position 564 in the PHY domain affects
photoversion of the P form to P, so drastically. Crystal
structure analysis of the Synechocystis phytochrome (Cph1) [37]
showed that the PHY domain contains an additional unique feature, a tongue-like protrusion that is present in all phytochrome
classes. According to Essen and colleagues [37] the tongue lies on the protein surface, it interacts with the GAF domain, seals the
chromophore pocket and thereby ensures close packing of the
chromophore and stabilization of the photoactivated P state of
phytochromes. The authors postulate that the very close packing
of the chromophore may play a role in passing the chromophore’s
conformational change onto the protein, leading to the light-
driven conformational change of phytochrome proteins and the
transduction of the light signal. A sequence comparison of
Arabidopsis phytochrome B and Cph1 shows that the G to E
mutation of phyB-401 lies exactly in the conserved WGG motif of
tongue. Thus one can speculate that replacing a small G residue with a bulky polar E residue could disturb exactly that part
of the phytochrome structure that couples the photoconversion of
chromophore and protein. Thus it is possible that the mutation
changes the conformation of the tongue-like protrusion in such a
way that the first light switch from the inactive P to the active P
leads to a change in the holoprotein structure that reduces the
dark reversion as well as the light-dependent conformational change
back to the inactive state. If the chromophore’s conformational change is uncoupled from the protein after the first switch, then
the phyB-401 mutant can hardly be switched off again by light.
After reverting light treatments the mutant photoreceptor stays in
an active form with the chromophore in its P state, while the
protein possesses a P-like structure, thus it is still capable of light
signalling. Only a slow reversion back to the real inactive P state
(Figure 8B) can switch off the signalling again.

We have attempted to obtain direct biochemical evidence for
demonstrating existence of the hypothesized phyB-401 P* conformer. As our data indicated above the P* conformer is
clearly capable of signalling. Accordingly, we argued that this
putative conformer should also be capable of binding to PIF3. To
this end we have determined and compared binding of PIF3 to
WT phyB and mutant phyB-401 P and P proteins by yeast two-
hybrid assays after different light treatments as shown in Figure S4.
The ultimate goal of these studies was to demonstrate in vitro that
FR treatment leads to the production of phyB-401 P* thus
reversion of PIF3 binding to the P conformer of phyB-401, in
contrast to that of phyB, shall not be complete. However, Figure S4 shows that capacity of P forms of phyB and phyB-401 to bind
PIF3 is approximately identical and that FR irradiation applied
after the inductive R pulse completely abolishes binding of PIF3.
Judicious interpretation of the data however suggests that these
results should not be considered as decisive proof for the non-
existence of the putative phyB-401 P* conformer. There are two
major caveats of this experimental approach. First we perform these
assays under saturating conditions and the sensitivity of such
an assay is obviously limited. Second, we do not have reliable information about the stability of the hypothetical P_{f1}^{*} in vitro as compared to in vivo. This latter one is especially a critical factor as the kinetics of accumulation of the hypothetical P_{f1}^{*} conformer is likely to be different in the cellular environment in planta and yeast two-hybrid assays.

Taken together, data presented in this paper and elsewhere [24,25] adequately support the importance of the PHY domain in regulating stability, i.e. dark reversion of the P_{f1} form. The ultimate proof for the mechanism by which it mediates passing the conformational change of the chromophore onto the protein could, however, only be obtained after the resolution of the 3D structure of the full phytochrome-B.

Materials and Methods

Plant material and growth conditions

Seeds were surface sterilized and sown on Petri-dishes (9 cm diameter) containing 4 layers of filter paper (Macherey-Nagel, Germany) and 4.7 ml distilled water. After stratification (three days at 4°C in darkness) uniform germination was induced by a 4 h white light treatment at 22°C. Afterwards seedlings were grown at 22°C in darkness for four days or subjected to various light treatments as described in the text. Transgenic plants expressing the 35S::PHYB:YFP and 35S:PHYB-401:YFP and GFP transgenes were generated in phyB-9 [38] backgrounds, as described [19,30]. The phyB-401 mutant was isolated in Landsberg erecta background [33].

Light treatments and hypocotyl length measurement

Dark-grown seedlings were handled under dim-green safelight [19]. For light pulse treatments and fluence rate response curves modified Leitz Prado 500-W universal projectors (Leitz, Wetzlar, Germany) were used. Red light was obtained by using KG65 filters (λ_{max} = 650 nm; Blazers, Liechtenstein) and far-red light was generated by filtering with an RG9 filter (λ_{max} = 775 nm; Schott, Germany) which absorbs 99.9% P_{r} [33]. Unless otherwise noted, all other red light treatments were done in a red light field (3 or 22 μmol m^{-2} s^{-1}) of fluorescent tube lamps (Philips TL 40 W/15) filtered with Plexiglas 501/3 (Rohm and Haas, Darmstadt, Germany). Hypocotyl length of 4-day-old seedlings treated with various light programs was measured by using ImageJ software (http://rsb.info.nih.gov/ij/).

Epifluorescent and light microscopy

For epifluorescent and light microscopy, seedlings were transferred to glass slides under dim-green safelight and analyzed with an Axioskop microscope (Zeiss, Oberkochern, Germany). Excitation and detection of YFP were performed with specific YFP filter (AHF Analysetechnik, Tubingen, Germany). Each experiment was repeated at least three times using at least five seedlings in which we monitored minimum 200 nuclei in each single experiment. Representative cells were documented by photography with a digital Axiocam camera system (Zeiss). Photographs were processed for optimal presentation using the Photoshop 7.0 (Adobe Systems Europe, Edinburgh, UK) and MS Office 97 (Microsoft, Redmond, WA) software packages.

In vivo spectroscopy

Dark reversion and photoconversion of transgenic PHYB-GFP and PHYB-401:GFP was measured in intact seedlings in a dual-wavelength ratio spectrophotometer (Ratiocam) [23,39]. Seedlings were irradiated with saturating 3 h R to create maximal P_{f1} levels and to degrade light-labile phyA. The complete loss of phyA signal after this treatment was confirmed in phyB-9 mutant seedlings (data not shown). For measurements of dark reversion, seedlings were either directly analyzed in the Ratiocam or transferred to darkness for measurement of P_{f1} levels in prolonged dark incubation. For photoconversion analysis, seedlings were placed on ice during the last 15 min of the R treatment and all later steps to avoid dark reversion. After R irradiations and before measurements seedlings were subjected to 30 s, 60 s or 135 s far-red light pulses (720 nm; 6 μmol m^{-2} s^{-1}).

The total amount of phytochrome (P_{tot}) was measured by six times alternate irradiation with far-red and red light followed by absorption measurements that gave the relative amount of phototranslatable phytochrome Δ(ΔA) of the sample. The absorption difference in the sample between the baseline at the beginning of spectroscopy and the first far-red light treatment 25 in the Ratiocam gave the P_{f1} level of the sample relative to P_{tot}. For each measurement around 300 seedlings were used and fresh weight was determined directly before the measurement to normalize Δ(ΔA) between the samples. Each time point was measured at least three times, error bars indicate standard error.

Calculation of P_{fr}/P_{tot} ratio under continuous irradiation

Jabben et al. [40] reported that the P_{fr}/P_{tot} ratio measured in etiolated mustard seedlings is fluence rate dependent. To account for this phenomenon they hypothesized that an unknown reaction inducing conversion of P_{fr} to P_{r} in dark must exist. This process called dark-reversion was shown to enhance conversion of P_{fr} to P_{r}, especially at low light. The ratio P_{fr}/P_{tot} is k_{1}/(k_{1}+k_{2}+k_{r}) whereby k_{1} and k_{2} are the rate constants for P_{fr} to P_{r} and P_{fr} to P_{r}, photoconversion, respectively. k_{r} is the rate constant for dark reversion. From Figure 3 k_{1} and k_{2} is estimated to be 0.008 s^{-1} and 0.002 s^{-1} at 6 μmol m^{-2} s^{-1}. k_{r} is calculated from Figure 2 (k_{r} = 0.00013 s^{-1}) if a first order dark reversion reaction is assumed.

Protein extraction, protein gel blotting and immunodetection

To analyze eR-induced degradation of phyB 3-day-old dark-grown seedlings were irradiated up to 24 h in 3 μmol m^{-2} s^{-1} red light and harvested together with a non-irradiated control at the end of irradiation period. Protein extraction and protein gel blotting was performed as described [30]. Immunodetection of phyB was performed using the monoclonal phyB antibody B6-B3 [41]. Horseradish-peroxidase-coupled anti mouse antiserum (Vector Laboratories) was used as secondary antibody. Development of the blot was done with the Phototrope Star Detection Kit (New England BioLabs). Quantification of phyB signals was carried out using the ImageJ software (http://rsbweb.nih.gov/ij/) and the gel analysis tools (described in the user manual: http://rsbweb.nih.gov/ij/docs/user-guide.pdf). Linearity of the signal was ensured by the use of internal standards.

Analysis of RNA accumulation by real-time RT-PCR

Far-red light treatments were carried out in a far-red light field for 5 min with varying intensities to obtain the aimed total fluences. Total RNA from Arabidopsis seedlings was isolated with Plant RNA reagent (Invitrogen) according to the manufacturer’s protocol. Purification and on-column DNaseI digestion was performed by using the RNeasy mini Kit from Qiagen (Hilden, Germany). After first-strand cDNA synthesis, the RT-PCR was accomplished by using an ABI Prism7300 (Applied Biosystems). FAM- and JOE-reporter dyes (5’end) were used for the TaqMan probes (see below). A standard curve method was used for the
calculation of relative transcript quantities [42]. Data were normalized to the corresponding ACTIN1 mRNA levels; phyB-401/phyA-201 relative dark levels were set to one. Each time point was measured at least in two biological replicates, error bars indicate standard error of technical replicates.

Sequences of primers and probes used for Real-Time RT-PCR were as follows: ACTIN1 (forward: 5'- GGCTCCAGGACGAT-GAAG-3'; reverse: 5'- ACCCTGAACTCCAGAGAAGTTT- 3'); probe: 5'JOE-CAAAAGTCGGTTGCCCTTCGAGAGG-3' BHQ1) and CAB2 (forward: 5'- GAGAGGGCCGAGCTGGC-TT-3'; reverse: 5'- CTCTGGGTCGGTAGCACCAC-3'; probe: 5'FAM-ACCCCGGTGAGTTTCGACC-3'TAMRA).

Yeast two-hybrid assay

Quantitative yeast two-hybrid assay was performed as previously described [43].

Supporting Information

Figure S1 Fluence rate dependent hypocotyl growth inhibition of seedlings expressing PHYB and PHYB-401 fusion proteins. Wild-type Col-0 (filled diamonds), phyB-9 mutant (empty squares) and transgenic seedlings expressing PHYB:GFP or PHYB:YFP (empty triangle) and PHYB-401:GFP or PHYB-401:YFP (empty circle) fusion proteins in phyB-9 (empty circle) were grown for 4 days under different fluence rates of cR (A) or cFR (B) light. Hypocotyl lengths were measured and relative hypocotyl lengths are shown. Each time point was measured at least three times, error bars indicate standard error.

(Fig)

Figure S2 Overexpression of the PHYB-401:YFP fusion protein does not affect skotomorphogenesis of the transgenic seedlings. Arabidopsis seedlings were germinated according to the standard protocol (after 3 days stratification at 4°C seeds were imbibed and treated with 4 h white light) and then grown for 4 days in darkness. Phenotypes of seedlings expressing the PHYB:YFP (A) and PHYB-401:YFP (B) fusion proteins in phyB-9 background and those of the wild-type Col-0 (C) and phyB-401/phyA-201 (D) double mutant are shown.

(Fig)

Figure S3 Degradation kinetics of the mutant and wild-type phyB do not differ significantly in cR. Accumulation levels of phyB and phyB-401 in phyA-201 background were analyzed by western blot hybridization using the monoclonal antibody B3B6 [41]. Degradation of the native phyB (A) and mutant phyB-401 protein (B) is shown in phyA-201 seedlings that were grown 4 days in darkness (d) and then irradiated with R light for 0, 3 and 24 h. The actin signals illustrate loading, the diagram (C) shows quantification of several western blots, error bars indicate standard error.

(Fig)

Table S1 Comparison of the expression levels of PHYB and PHYB-401 fusion proteins to the endogenous phyB and phyb-401 photoreceptors. Accumulation levels of phyB, phyB-401 and those of the PHYB and PHYB-401 fusion proteins were analyzed by western blot hybridization using the monoclonal antibody B3-B6 [41]. Western blot hybridization assays were quantified (as described in Materials and Methods section) and expression levels of the PHYB and PHYB-401 fusion proteins in phyB-9 background, the native phyB in Col-0 and phyb-201 (L. erecta) backgrounds as well as the phyb-401 protein in phyb-201 (L. erecta) background were compared. Fold expression levels normalized to endogenous phyB in Col-0 are shown.

(Fig)

Acknowledgments

We thank Peter Quail for providing the monoclonal phyB antibody B6-B3 and Erzsébet Fejes, László Kozma-Bognár for critical reading of the manuscript.

Author Contributions

Conceived and designed the experiments: ES FN. Performed the experiments: EA AH JB FW AV ME MM SK. Wrote the paper: ES FN. Created figures and handled manuscript for submission: EA AV SK FN.

References


