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Astrocyte pathology and the absence of non-cell autonomy in an induced pluripotent stem cell model of TDP-43 proteinopathy

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Glial proliferation and activation are associated with disease progression in amyotrophic lateral sclerosis (ALS) and frontotemporal lobar dementia. In this study, we describe a unique platform to address the question of cell autonomy in transactive response DNA-binding protein (TDP-43) proteinopathies. We generated functional astroglia from human induced pluripotent stem cells carrying an ALS-causing TDP-43 mutation and showed that mutant astroglial progenitors display an increased level of TDP-43, subcellular mislocalization of TDP-43, and decreased cell survival. We then performed coculture experiments to evaluate the effects of M337V astrocytes on the survival of wild-type and M337V TDP-43 motor neurons, showing that mutant TDP-43 astrocytes do not adversely affect survival of cocultured neurons. These observations reveal a significant and previously unrecognized glial cell-autonomous pathological phenotype associated with a pathogenic mutation in TDP-43 and show that TDP-43 proteinopathies do not display an astrocyte non-cell-autonomous component in cell culture, as previously described for SOD1 ALS. This study highlights the utility of induced pluripotent stem cell-based in vitro disease models to investigate mechanisms of disease in ALS and other TDP-43 proteinopathies.

transactive response DNA-binding protein (TDP-43) is the major component of ubiquitinated cytoplasmic and nuclear inclusions in neurons and astroglia in amyotrophic lateral sclerosis (ALS) and a subgroup of frontotemporal lobar degeneration (FTLD-TDP) (1–3). These pathological hallmarks provide a unifying description of a range of conditions defined as TDP-43 proteinopathies (4). At present, >30 mutations in the TDP-43 gene (TARDBP) have been linked to familial ALS (fALS) (5), strongly suggesting a causative role for TDP-43 in the pathogenesis of ALS.

Accumulating evidence from experimental systems implicating non-cell-autonomous mechanisms in ALS has highlighted the importance of the glial cellular environment to motor neuron (MN) degeneration (1, 5, 6–9). In two rodent models of ALS with lineage-specific SOD1 expression have particularly influenced our understanding of the nonneuronal contribution to disease progression. Glial expression of mutant SOD1 cannot initiate MN disease on its own, but is necessary for disease progression (6, 7). Furthermore, astroglia precedes MN degeneration in some animal models and is a dominant feature of all human ALS pathology (4, 6, 10). Collectively, these observations highlight the need to better understand the nature of astroglial pathology in ALS. Combining developmental neurobiological principles of cell fate determination with human induced pluripotent stem cell (iPSC) lines derived from patients carrying ALS disease-causing mutations may provide important insights into astroglia pathology.

We recently generated human MNs from iPSC lines derived from a fALS patient and demonstrated that the M337V TDP-43 mutation confers cell-autonomous toxicity to MNs (11). Moreover, quantitative Western blot and immunohistochemical analysis of M337V neuronal cultures revealed an increase in levels of soluble and detergent-resistant TDP-43 protein in the absence of any detectable changes in the nuclear TDP-43 immunohistochemistry, implicating cytoplasmic mislocalization of TDP-43 in disease pathogenesis (11). Given that key features of the TDP-43 proteinopathies could be detected in iPSC-derived neurons, and noting recent independent confirmation in multiple patient lines (12), we addressed the possibility that astroglial TDP-43 pathology could be investigated by the same approach.

Here we describe the pathological effects of mutant TDP-43 in isolated functional astrocytes, generated by a direct astrocyte specification protocol from patient-derived M337V iPSC lines. We then investigate the influence of mutant astrocytes on neurons to determine whether non-cell-autonomous toxicity can be detected in cell culture.

Results

Generating Functional Astrocyte Populations from iPSC Lines. Astroglial populations from two TDP-43 M337V and two control (CTRL) iPSC lines were derived from neural precursors (NPCs) (Fig. L4), generated as described (11). NPCs were cultured in suspension as neurospheres in medium containing epidermal growth factor (EGF) and leukemia inhibitory factor (LIF) for 4–6 wk. In the absence of fibroblast growth factor 2 (FGF2), coaplication of EGF and LIF in prolonged culture promoted astroglial specification of NPCs (13, 14) and efficiently selected for neurospheres with a high content of astroglial progenitors. After this enrichment phase, neurospheres were expanded in EGF- and FGF2-containing medium before enzymatic dissociation to single cells. The resulting populations were positive for vimentin and nuclear factor 1A, two markers of astrocyte progenitor cells (APCs) (refs. 15 and 16; Fig. 1B), and could be propagated as a monolayer culture with EGF and FGF2. Less than 30% of cells in early passage APC cultures were positive for the astrocytic marker glial fibrillary acidic protein (GFAP). Subsequent differentiation over 14 d in the presence of ciliary neurotrophic factor (CNTF) increased the proportion of GFAP-positive cells with >90% of the cells coexpressing the astrocytic markers GFAP and S100β (Fig. 1C and D). Fewer than 2% of cells were positive for


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the neuronal marker βIII tubulin, confirming the efficiency of this protocol in generating highly enriched APC populations.

No difference was observed in the efficiencies of astrocyte differentiation in TDP-43 M337V and CTRL iPSC lines (Fig. 1F). The differentiated populations of iPSC-derived astrocytes were positive for the metabolotropic glutamate transporter EAAT1 (Fig. S1A) and could uptake t-glutamate from the medium in a time-dependent fashion (Fig. 2A). t-glutamate uptake was blocked by the glutamate transporter inhibitor l-transpyrrolidine-2,4-dicarboxylic acid or by removing sodium from the medium, consistent with the mechanism of action of EAAT1. No difference was observed in the t-glutamate clearance properties of mutant and CTRL astrocytes (Fig. 2B and C).

We next sought to determine whether the iPSC-derived astrocytes promoted the formation of mature synapses. Astrocytes from either CTRL or mutant iPSC lines were cocultured with differentiating CTRL iPSC-derived ventral-spinal cord patterned neurons (11) for a period of 3 wk. Cultures were stained with antibodies against the presynaptic protein synaptophysin 1 (PSY) and the postsynaptic density protein PSD-95 to quantify the number of mature synapses in cocultures and isolated neuronal cultures (17–19). No difference between mutant and CTRL astrocytes was observed in promotion of PSY+/PSD-95+ synaptic puncta on CTRL neurons (Fig. 2 D and E). Moreover, similar numbers of PSY+/PSD-95+ synaptic puncta were found in M337V and WT neurons when cocultured with M337V or WT astrocytes, suggesting that synaptogenesis is independent of the genetic background of the neurons and the glia (Fig. 2F). Both genotypes of iPSC-derived astrocytes exhibited the potential to propagate calcium waves upon mechanical stimulation. (Fig. 2G and Movie S1). Calcium waves were also elicited when astrocytes were stimulated by local application of ATP (Movie S2), as described (20–22). This ATP-evoked increase in cytosolic calcium was abolished by application of 2-aminoethoxydiphenyl borate, an inhibitor of IP3-dependent calcium release (22) (Movie S3). These findings confirm functional equivalence between TDP43 mutant and CTRL astrocytes.

Characterization of TDP-43 in iPSC-Derived Astrocytes. We then sought to investigate the consequence of M337V mutation on astrocyte mRNA expression, protein levels, and subcellular localization of TDP-43. Quantitative RT-PCR (qRT-PCR) analysis revealed no significant difference in the expression levels of TARDBP and HDAC6 (which is transcriptionally regulated by TDP-43 itself; ref. 23) (Fig. 3A). M337V and CTRL cultures also showed no difference in expression levels of astrocytic markers GFAP and S100β, further confirming that both genotypes can give rise to comparable astrocyte populations (Fig. 3A). Having established that the level of TDP-43 transcripts do not differ between M337V and CTRL astrocytes, we investigated whether the mutant astrocytes exhibit a cellular and biochemical signature (24) similar to that observed in MN cultures derived from the same iPSC lines (11). Immunoblot analysis showed that M337V astrocytes have significantly more soluble TDP-43 (Fig. 3B) (11). Quantitative RT-PCR (qRT-PCR) analysis revealed no significant difference in the expression levels of TARDBP and HDAC6 (which is transcriptionally regulated by TDP-43 itself; ref. 23) (Fig. 3A). M337V and CTRL cultures also showed no difference in expression levels of astrocytic markers GFAP and S100β, further confirming that both genotypes can give rise to comparable astrocyte populations (Fig. 3A). Having established that the level of TDP-43 transcripts do not differ between M337V and CTRL astrocytes, we investigated whether the mutant astrocytes exhibit a cellular and biochemical signature (24) similar to that observed in MN cultures derived from the same iPSC lines (11). Immunoblot analysis showed that M337V astrocytes have significantly more soluble TDP-43 (Fig. 3B) (11). Quantitative RT-PCR (qRT-PCR) analysis revealed no significant difference in the expression levels of TARDBP and HDAC6 (which is transcriptionally regulated by TDP-43 itself; ref. 23) (Fig. 3A). M337V and CTRL cultures also showed no difference in expression levels of astrocytic markers GFAP and S100β, further confirming that both genotypes can give rise to comparable astrocyte populations (Fig. 3A). Having established that the level of TDP-43 transcripts do not differ between M337V and CTRL astrocytes, we investigated whether the mutant astrocytes exhibit a cellular and biochemical signature (24) similar to that observed in MN cultures derived from the same iPSC lines (11). Immunoblot analysis showed that M337V astrocytes have significantly more soluble TDP-43 (Fig. 3B) (11).
cytoplasmic fluorescence. We noted a strong influence of expression levels on TDP-43 localization. Elevated expression levels were directly related to cytoplasmic mislocalization of TDP-43 (Fig. 4D), as reported (28). However, whereas WT TDP-43 displayed a dynamic range of localization correlating with expression levels, M337V TDP-43 showed an inherent bias toward cytoplasmic localization (Fig. 4C), even at low expression levels (Fig. 4D). These results confirm that the observed changes in localization of TDP-43 in iPSC-derived astrocytes are due to the presence of the M337V mutation on TARDBP.

**Survival Analysis on iPSC-Derived Astrocytes.** We next asked whether the accumulation of soluble and cytoplasmic TDP-43 was accompanied by a reduction in survival as assessed by longitudinal live fluorescence microscopy as described (11, 28). M337V and CTRL iPSC-derived astrocytes were transfected with a plasmid that constitutively expresses enhanced green fluorescent protein (EGFP) and imaged every 24 h for 10 d. Astrocyte death was marked by loss of fluorescence or dissolution of the cell itself (Fig. S4). These criteria are at least as specific as traditional markers of particular cell death pathways (29), have the advantage of detecting all forms of cell death in a single assay, and are therefore more sensitive. Kaplan–Meier survival analysis was used to plot cumulative hazard curves depicting the risk of death for M337V and CTRL astrocytes (Fig. S4). Cox proportional hazards analysis was then applied to calculate the relative risk over four independent experiments, demonstrating a cumulative hazard ratio (HR) of 2.5 (P = 2 × 10−16; log-rank test) for M337V astrocytes, indicating a 2.5-fold greater risk of death associated with the TDP-43 M337V mutation in comparison with CTRLs.

Having established that the M337V mutation reduces the survival of astrocytes under basal conditions, we then investigated the mechanism of this toxicity. To determine whether the observed cell death was due to increased apoptosis, the effect of pan-caspase inhibitor QVD-oph was measured (30). Treatment with the pan-caspase inhibitor decreased the cumulative HR of M337V and CTRL astrocytes. QVD-treated CTRL astrocytes showed a cumulative HR of 0.33 (P = 5.07 × 10−11; log-rank test) compared with the vehicle-treated group. On the M337V background, the QVD-treated group showed a HR of 0.72 (P = 0.0145; log-rank test, compared with vehicle-treated CTRL astrocytes), and the vehicle-treated M337V group showed a HR of 2.66 (P = 8.13 × 10−7; log-rank test, compared with vehicle-treated CTRL astrocytes) (Fig. 5D). Therefore, the presence of caspase inhibitor decreased the risk of death more than threefold in both genotypes. However, when the M337V and WT QVD-treated groups were compared directly, the M337V astrocytes still displayed a significantly greater HR than WT astrocytes (2.19; P = 2.32 × 10−12; log-rank test). The comparison of HRs between M337V and CTRL astrocytes with QVD or vehicle-only treatment revealed that caspase inhibition failed to significantly lower mutant TDP-43 specific toxicity (Fig. 5C). Furthermore, survival analysis performed within the transfected astrocyte populations described earlier revealed that cytoplasmic localization of mutant TDP-43 was associated with a 225% increase in the risk of death (HR = 2.23; P = 0.0002). These results suggest that cytoplasmically mislocalized M337V TDP-43 significantly increases the risk of death of astrocytes.

**Analysis of MN–Astrocyte Cocultures.** We next sought to determine whether mutant M337V astrocytes exerted a non-cell-autonomous toxic effect on MNs, similar to that described for mutant SOD1 rodent astrocytes (1, 8, 9, 31, 32). To this end, we first verified the utility of survival analysis by longitudinal microscopy for detecting non-cell-autonomous toxic effects of glia on WT MNs derived from human iPSCs by coculturing HB9:GFP-transfected WT MNs on murine primary astroglia overexpressing either hSOD1 WT or hSOD1 G93A. This experiment, as predicted from previous studies (8), revealed an increased toxic non-cell-autonomous effect exerted by hSOD1 G93A glia compared with hSOD1 WT counterparts on WT MNs (Fig. S5), confirming that a longitudinal microscopy-based survival analysis approach is capable of detecting astrocyte toxicity in human iPSC-derived MN cocultures.
We next determined the survival of mutant M337V MNs in astrocyte coculture experiments. First, we confirmed that mutant MNs cultured in isolation demonstrated greater cell-autonomous vulnerability than CTRL MNs, as previously shown (Fig. 5E) (11). Using real-time survival analysis, we found that mutant astrocytes did not exert non-cell-autonomous toxic effects on MNs carrying the M337V TDP-43 mutation (Fig. 5F). Indeed, when cocultured with astrocytes of either genotype, the previously observed cell-autonomous vulnerability of isolated mutant MNs was no longer evident. These results suggest that, independent of the genetic background of glia, astrocyte coculture rescues the survival difference associated with the TDP-43 M337V mutation in motor neuronal cultures and that iPSC-derived TDP-43 mutant astrocytes do not exert an in vitro toxic effect on neurons.

Discussion

The present study describes a platform to study the glial component of TDP-43 proteinopathies and its effect on cocultured neurons. We efficiently generated near-homogenous populations of astrocytes from both CTRL and mutant iPSC lines and cocultured astrocytes and MNs from different genetic backgrounds to study potential non-cell-autonomous contributions to ALS pathology.

We exploited developmental gliogenic signaling pathways to generate enriched and scalable astroglial progenitors. Propagating NPCs with LIF and EGF, but without FGFR2, promoted glial commitment and expansion and resulted in efficient APC fate specification within 6 wk (33–36). A number of methodologies derive astroglia from pluripotent cells with varying levels of function and purity (37–40). The protocol described here achieves functional astrocyte differentiation without serum supplementation, presenting a more chemically defined and comparably faster alternative to other methods (37). The resulting astroglial populations have low numbers of neuronal cells (<2%), enabling the study of astroglial survival, function, and biochemistry in near-homogenous populations. This homogeneity has a clear benefit over methodologies that yield glial populations with greater neuronal differentiation (38) or those that require longer periods of time. Highly enriched astrocyte populations can also be generated by prolonged culture (3–9 mo) of neural progenitors with gradual temporal loss of neurogenic potential, similar to the neurogenic–gliogenic switch during development (39, 40).

M337V-expressing astrocytes displayed cytoplasmic mislocalization of TDP-43 with elevated levels of soluble TDP-43 protein, which was not due to an increase in TARDBP mRNA. There was no increase in detergent-resistant TDP-43 in M337V mutant astrocytes, but their survival was significantly reduced under basal conditions. The effects on survival were replicated in CTRL iPSC-derived astrocytes transiently transfected with M337V mutant TDP-43. These features are comparable to earlier findings in isolated MN cultures (12) and, together with the increased stability of mutant TDP-43 reported in isogenic stable cell lines (41), suggest that the M337V mutation affects TDP-43 protein stability regardless of expression level and membrane localization.
Survival analysis of M337V iPSC-derived astrocytes revealed a significantly greater risk of death compared with WT astrocytes under basal conditions. This result represents a unique report of cell-autonomous astrocyte toxicity in a patient-derived iPSC model. These findings are consistent with the idea that—at least initially or in part—reactive astrogliosis observed in ALS might not be simply a response to neuronal injury but a consequence of direct mutation-mediated astrocyte toxicity. Treatment of mutant and CTRL astrocytes with pan-caspase inhibitor resulted in a three to fourfold improvement in cell survival on both genetic backgrounds. However, when QVD-treated M337V and CTRL astrocytes were compared directly, they still had a significant difference in their survival, as measured by Cox proportional hazards analysis. This result suggests that the difference in survival between M337V and CTRL astrocytes can be attributed to caspase-independent mechanisms. Indirect support for this possibility is based on the absence of TDP-43 aggregates or significant increase of TDP-43 cleavage products in mutant astrocytes, both of which are associated with caspase activity (46, 47). Although further studies are required to gain a better understanding of the mechanism of M337V TDP-43 toxicity on astrocytes, the ectopic M337V and WT TDP-43 expression studies on isogenic WT astrocytes reported here demonstrate that the M337V mutation confers an inherent bias toward cytoplasmic localization. In turn, the observation that cytoplasmic localization is associated with an increased risk of death offers an interesting correlation between the increased cytoplasmic TDP-43 of iPSC-derived astrocytes and their lower survival, compared with CTRLs. Importantly, these experiments involved isogenic human astrocytes, so the differences in mislocalization and toxicity can be attributed specifically to the presence of a disease-associated mutation in TDP-43, and not to potential influences of differences in genetic background.

Recent in vitro and in vivo models convincingly demonstrated that astrocytes mediate, at least in part, disease progression in SOD1 fALS models (1, 8, 9, 31, 32). This finding, in turn, led to the concept of non-cell-autonomous neurodegeneration. However, SOD1 fALS cases comprise a great majority of sporadic ALS (sALS) and fALS cases, do not show TDP-43 pathology and are not therefore classified as TDP-43 proteinopathies (26). Understanding whether the TDP-43 mutation carrying astrocytes have a comparable neurotoxic effect is thus of great interest. Our previous iPSC-based study showed that the M337V mutation impairs survival in isolated MNs cultured under basal conditions (11). In this study, we used the same longitudinal microscopy–based approach to examine the survival of MNs cocultured with either M337V or CTRL astrocytes. We detected no significant difference in survival of CTRL MNs plated on mutant astrocytes and equivalent MNs cocultured with CTRL astrocytes. Importantly, we show, using the same longitudinal microscopy-based methodology, that murine glia overexpressing hSOD1 are toxic to human iPSC-derived MNs, confirming the ability of this experimental system to address cellular autonomy in iPSC-based models as has previously been shown in other xeno-cultures (1, 8).

Notwithstanding the necessity of further validation of our results in independent human iPSC lines, our results suggest a potential difference in the consequence of astroglial–neuronal interaction, at least when modeled under basal conditions in vitro, between SOD1 and TDP-43 iALS. Interestingly, astrocytes derived from postmortem sALS and SOD1 iALS patients have a non-cell-autonomous toxic effect on mouse ES cell-derived MNs, extending the concept of toxicity mediated by sporadic astroglia to include sporadic MN disease (9). Reactive astrogliosis is defined by a number of phenotypic changes in astrocytes (7), and the extent of mechanistic overlap between the SOD1 iALS and SALS astrogliosis-mediated MN toxicity remains to be independently confirmed. It is also important to note that non-cell-autonomous toxicity of human sporadic or, indeed, SOD1 iALS iPSC-derived astrocytes has, to date, not been reported. One potential explanation is that in vitro differentiation of astrocytes from human iPSCs, as in our study, in the absence of degenerating neurons does not capture some of the as-yet-uncharacterized mutation-specific or more general reactive properties of in vivo astroglia. Nevertheless, the platform described here is ideal to

protein levels via a posttranslational mechanism. It is also of interest that in the astrocyte populations analyzed, TDP-43 does not seem to negatively autoregulate its own mRNA, as reported in established stable lines (42). Mutant astrocytes, unlike normal MNs derived from the same lines, do not show increased levels of detergent-resistant TDP-43 levels. The reason for this finding is unclear but suggests different cell-specific processing of protein accumulation. Along with accumulation of soluble TDP-43, mutant astrocytes displayed inherent mislocalization of cytoplasmic TDP-43. Increased cytoplasmic TDP-43 levels were not accompanied by a corresponding depletion of nuclear TDP-43, further supporting the idea that the additional TDP-43 is due to increased stability and/or slower clearance. Interestingly, the loss of TDP-43 nuclear localization has been linked to neuronal degeneration in TDP-43 proteinopathies in animal models (43) and cellular systems (44, 45). The absence of TDP-43 nuclear clearance in M337V iPSC-derived MNs (11) and astrocyte cultures suggests that loss of nuclear TDP-43 is a later event in the disease process.

Fig. 5. Survival analysis on iPSC-derived astrocytes and MN-astrocyte cocultures. (A) Real-time survival analysis of M337V iPSC-derived astrocytes and CTRLs. Mutant astrocytes showed increased cumulative risk of death associated with M337V TDP-43 under basal conditions (HR = 2.5, P = 2 × 10−10, CTRL taken as baseline; ***P < 0.001). (B) Real-time survival analysis of M337V and CTRL iPSC-derived astrocytes in the presence of 10 μM QVD-oph. The inhibition of caspase activation reduced the risk of death in M337V and CTRL astrocytes (with CTRL vehicle as a reference, in M337V + vehicle, HR = 2.61, P = 8.13 × 10−12; M337V + QVD, HR = 0.72, P = 0.0145; CTRL + QVD, HR = 0.33, P = 5.07 × 10−11; ***P < 0.001). (C) Comparison between cumulative HR of M337V astrocytes in QVD- and vehicle-treated groups. The error bars represent 95% confidence intervals. (D) Real-time survival analysis of WT iPSC-derived MNs plated on either M337V or CTRL iPSC-derived astrocytes. Mutant astrocytes were toxic to cocultured WT MNs (with WT MNs on WT astrocytes as a reference, in either M337V or CTRL iPSC-derived astrocytes. Mutant astrocytes were not toxic to cocultured WT MNs. M337 astrocytes had no detectable toxicity to cocultured MNs. Both WT and M337V astrocytes improved the survival of M337V MNs. M337 astrocytes had no detectable toxicity to cocultured MNs. All graphs represent pooled data from three independent experiments.

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study both the reactive properties of in vitro-generated astrocytes and the potential differences between astrocytes generated from sALS and other TALS iPSC lines.

In conclusion, this study established a platform to address the biological consequence of somatic mutations on human iPSC-derived astroglia and to dissect the contribution of glial–neuronal cross-talk in neurodegenerative processes. The application of the platform to study the non-cell-autonomous component of TDP-43 proteinopathies and the description of a previously unrecognized cell-autonomous TDP-43 toxic effect on astrocytes further highlights the potential of using patient-derived iPSCs to gain a deeper understanding of the molecular pathogenesis of neurodegenerative disorders.

Materials and Methods

Generation of Astrocytes from iPS Cell Lines. Generation of neurospheres from iPSC lines and their patterning to acquire MN progenitor identity were as described (11). Neurospheres were mechanically chipped at the beginning of the enrichment phase and cultured in NSCR EL20 medium for 2–4 wk.

After enrichment, the spheres were propagated in EGF and FGFI2 containing medium and passed mechanically by chipping them every 2 wk. Details on establishment of monolayer cultures, dissociation of spheres, media composition, and functional characterization of the astrocytes are provided in SI Materials and Methods.

Survival Analysis. For survival analysis, differentiated astrocytes were plated on 96-well Matrigel-coated plates at a density of 2 × 10³ cells per well and transfected with pGW1-mApple (Fig. 2G) or cotransfected with pGW1-EGFP and pGW1-TDP43-mApple (Fig. 2 H and J) with Lipofectamine 2000 (Invitrogen). For analysis of cocultures, dissociated MNs from patterned neurospheres were plated on 96-well Matrigel-coated astrocyte platform. Transfection was performed as described (11). A robotic microscope was used to perform the imaging (28, 29). Following transfection, cells were imaged at 24-h intervals for 10 d, and survival was determined by using algorithms developed in MatLab and ImageJ. Details on transfections, survival analysis software, and procedures used are provided in SI Materials and Methods.

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