



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

A computed tomographic study of the molar teeth of *Babyrousa* spp.

Citation for published version:

Macdonald, A, Ziehmer, B, Kitchener, A, Gelang, M, Åblad, B, Lintonsson, R, von Pückler, K, Schaub, S, Kiefer, I & Schwarz, T 2024, 'A computed tomographic study of the molar teeth of *Babyrousa* spp.', *Journal of Veterinary Dentistry*, vol. 41, no. 1, pp. 31-42. <https://doi.org/10.1177/08987564241248818>

Digital Object Identifier (DOI):

[10.1177/08987564241248818](https://doi.org/10.1177/08987564241248818)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Journal of Veterinary Dentistry

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



A Computed Tomographic Study of the Molar Teeth of *Babyrousa* spp.

Alastair A Macdonald, BSc, PhD, Dr Utrecht, MBA, FHEA¹, Bianca Ziehmer, BSc, MSc², Andrew C. Kitchener, BSc, PhD³, Magnus Gelang, PhD⁴, Björn Åblad, DVM⁵, Ruth Lintonsson, DSV, MI&F⁵, Kerstin von Pückler, Dr. med. vet., DECVDI⁶, Sebastian Schaub, Dr. med. vet., DECVD⁶, Ingmar Kiefer, Dr. vet. med.⁷ and Tobias Schwarz, MA, Dr. med. vet., FRSB, PgCAP, FHEA, DECVDI, DACVR, DVR, FRCVS¹

1. Royal (Dick) School of Veterinary Studies & The Roslin Institute, University of Edinburgh, Edinburgh, UK.
2. Department of Viticulture and Agriculture, Ministry of Economic Affairs, Transport, Agriculture and Viticulture Rhineland Palatinate (MWVLW), Mainz, Germany.
3. Department of Natural Sciences, National Museums Scotland, Edinburgh, UK. Institute of Geography, School of Geosciences, University of Edinburgh, Edinburgh, UK.
4. Göteborgs Naturhistoriska Museum, Goteborg, Sweden.
5. Blåstjärnans Djursjukhus, Bildenheten, Göteborg, Sweden.
6. Justus-Liebig-Universität Giessen, Klinik für Kleintiere, Radiologie, Justus-Liebig Universität, Gießen, Germany.
7. Universität Leipzig, Veterinärmedizinische Fakultät, Klinik für Kleintiere, Leipzig, Germany.

Abstract

A photographic and computed tomography (CT) scanning study was carried out on the molar teeth of 18 adult male *Babyrousa babyrussa* skulls and 8 skulls of *Babyrousa celebensis* including 7 adult males, 1 adult female and 1 subadult male. The occlusal

morphology of the adult maxillary and mandibular molar teeth of *B. babyrussa* was very similar to that of *B. celebensis*. Most *B. babyrussa* maxillary molar teeth had 6 roots, with small numbers of teeth having 4, 5 or 7 roots. A similar pattern was suggested in *B. celebensis*. Mandibular molar teeth had between 4 and 8 roots. Tooth roots of first and second maxillary and mandibular molar teeth were largely tapering, rod-like structures. The roots of the 111/211 teeth had a more complex arrangement; some inserted almost vertically into the maxilla; others orientated in a more distal direction. The mesial and distal roots were splayed in appearance. The 311/411 tooth roots retained elements of the open 'C' shape and were generally orientated distally. The pulp chambers were arched to fit under the main cusps in all molar teeth. Pulp canals were variable in number.

Key words

anatomy, wild pig, babirusa, *Babyrousa spp.*, tooth root, root canal

Introduction

The babirusa (genus *Babyrousa* Perry, 1811) is a suid endemic to the Indonesian island of Sulawesi and a number of smaller islands to the east.^{1,2} Illustrations of the molar teeth of the babirusa were published by Cuvier, Geibel and Stehlin.^{3,4,5} Babirusa molar teeth are brachydont, in which the root is longer than the crown, and these do not grow continuously. Physical evidence of an early interest in studying the roots of these molar teeth was indicated by the half-skull and mandible of a young adult male Sulawesi babirusa (*B. celebensis*) (Figure 1A); in 1860 this specimen was transferred from the Christ Church college collection into the main Oxford University Museum (OUM) collection - OUM 05266. Excavation of sections of the right lateral surface of the skull gave some indication of the arrangement of the lateral (buccal) roots of the molar teeth. Modern computed tomography (CT-scanning) techniques have offered the opportunity to examine the roots of babirusa teeth in a number of skulls without damaging the overlying bone (Figure 1B).

During studies of the anatomy of the babirusa stomach it was found that the ingested plant material had a 'granular appearance'.⁶ Research had also revealed that the mucosal surface lining of the of the cardiac gland region of the stomach was very much

larger than that of the *Sus* pigs^{7,8} and that it was uniquely covered in a layer of cells arranged in the form of a honeycomb.⁹ Examination of the wear patterns of babirusa molar teeth showed that there was heavy wear of maxillary and mandibular first molar teeth and a relative delay in the wear of the corresponding second and third molar teeth.⁶ It was also known that babirusa would select to eat fruits with hard seeds, and that these were cracked open by the molar teeth (Leus and Macdonald, 1995, unpublished observations). Recent studies have focused on the anomalous molar wear patterns found in a proportion of the (largely) North Sulawesi babirusa, *B. celebensis*, that seem to be regionally found in association with the presence of one particular type of wild banana, *Musa balbisiana* Colla.¹⁰ This wild banana species contains 150-170 very hard, small pea-sized seeds¹⁰.

These dietary observations prompted several questions. Could the numbers and the orientation of the roots of mandibular and maxillary molar teeth provide some form of insight into the function of babirusa molar teeth? Are the molar tooth roots anatomically arranged in some specific way to distribute the crushing force of a bite into the bony tissues of the mandibles? How might the structure of the corresponding number and arrangements of the maxillary tooth roots support the crushing and grinding motions of the occlusal surfaces against those of the mandibular teeth?

A search of the literature failed to reveal the results of any systematic study of the anatomy of the molar teeth of babirusa. As a consequence, we undertook a CT-scanning examination of babirusa skulls from four museum collections.^{a-d} The skulls selected represented the babirusa species from Buru and the Sula Islands (*B. babyrussa*) and from the island of Sulawesi (*B. celebensis*). Additional skulls were accessed for specific topographical information, e. g. Oxford University Museum, England (OUM).

The modified Triadan system of veterinary dental nomenclature¹¹, as illustrated for babirusa dentition¹², was adopted for this study to provide the description of dental location and orientation.¹³

Materials and Methods

Twenty-six babirusa skulls were studied. The primary research material comprised 13 adult male *B. babyrussa* skulls catalogued as from Buru. Five adult male *B. babyrussa* skulls were morphologically identified as coming either from Buru or the Sula Islands and 7 adult male and 1 adult female *B. celebensis* skulls from Sulawesi (Table 1). The total number of babirusa teeth available for evaluation was 295 (Table 2). One *B. babyrussa* skull had no 409 or 410 teeth and one *B. celebensis* skull had no 310 tooth. Two *B. babyrussa* skulls had no 109 teeth and one *B. celebensis* skull had no 109 tooth. The mandibular tooth measurements from one male *B. celebensis* (AAM355) were not recorded.

All specimens were photographed, including close-ups of the occlusal surfaces of the teeth. To enable anatomical clarity in the description of observations, the dental anatomical nomenclature of the babirusa teeth¹⁰ was derived from that published for Suidae.¹⁴ The main cusps on the maxillary molars are called paracone, protocone, metacone, tetracone, and pentacone (Figure 2A). The main cusps on the mandibular molars are called protoconid, metaconid, hypoconid, entoconid, and pentaconid. A smaller sixth cusp can be seen on the 311 tooth and it is named hexaconid (Figure 2B).

For curatorial and geographical reasons (Table 1) the skulls were CT scanned at four different institutions using four different CT scanner models.^{e-h}

Tooth length was measured from the highest point of the crown (the coronal end) to the apex (the extremity) of the tooth root. Tooth root length was measured from either the buccal or the lingual/ palatal cemento-enamel junction (depending on root orientation in the CT scan) to the apex of the root. The occlusal edges of the molar tooth were formed from the enamel cusps. The various ways in which the apical ends of the 111/211 more distal roots subdivided made comparative measurements unreliable. Tooth root width was measured across the mid-length of the tooth root.¹²

Statistical analysis of variance analysesⁱ were undertaken on the teeth of the adult babirusa.

Results

Occlusal morphology

The occlusal morphology of the adult maxillary molar teeth of *B. babyrussa* (Figure 2A) was very similar to that of *B. celebensis*. The paracone of each of the molar teeth was in the form of a buccally situated, rounded cone. On its distal surface lay the metacone. The paracone of 110/210 was bordered distally on its buccal side by a slightly smaller metacone. The protocone was situated on the mesiopalatal side of the crown and was of equivalent size to the paracone. Distal to it lay the tetracone. Distal to the protocone and the tetracone was the smaller pentacone.

Similarly, the occlusal morphology of the adult mandibular molar teeth of *B. babyrussa* (Figure 2B) was very similar to that of *B. celebensis*. The protoconid of each of the molar teeth was a conical, buccally situated, cusp. The adjacent, lingually situated, metaconid was approximately equivalent in size. The hypoconid and entoconid were situated distal to these, on the buccal and lingual sides respectively. The somewhat smaller pentaconid occupied the distal aspects of 309/409 and 310/410. In the 311/411 the smaller again hexaconid occupied the distolingual side of the tooth adjacent to the buccally situated pentaconid.

The morphology of the dentine underlying the enamel of the occlusal surface of the maxillary and mandibular molar teeth was as indicated in Figures 2C and 2D respectively. The irregular cone-shapes of the dentine corresponded, in reduced form, to the occlusal morphology of the overlying enamel layers of the maxillary and mandibular molar teeth.

Number of tooth roots

The numbers of the measurable roots found on each molar tooth are summarised in Table 2. Small bumps and short ridges were also present on the undersides of some teeth. The majority of *B. babyrussa* maxillary molar teeth had 6 roots, with small numbers of teeth having 4, 5 or 7 roots. The 4 main roots were positionally designated mesiobuccal, mesiopalatal, distobuccal and distopalatal and the orientation of the additional roots was as illustrated in (Figure 3A). Despite the smaller sample number of *B. celebensis* maxillary teeth, a similar pattern may pertain in this species also. The orientation of the mandibular roots was indicated by the pattern of root alveoli in the

exposed mandible (Figure 3B). The majority (60%) of *B. babyrussa* 309/409 and 310/410 teeth had 4 roots (Figures 3C & 3D) positionally designated mesiobuccal, mesiolingual, distobuccal and distolingual, with the remaining 40% having 5 roots, the fifth root designated as central. None of the 311/411 teeth in that species had 4 roots, with the majority (38.9%) having 7 roots, including those designated as central, buccal and lingual. The 309/409 and 310/410 teeth of *B. celebensis* had 4 or 5 roots, while the 311/411 teeth had between 6 and 8 roots (Table 2).

Tooth root structure

The 109/209 tooth roots were largely tapering, rod-like structures (Figure 1B). These had an irregular, somewhat oval cross-section with many showing a semi-closed or open 'C' shaped construction (Figure 3A). The distobuccal root insertion exhibited a slight distal tilt from vertical in 75% of these teeth in *B. babyrussa* and in 50% of *B. celebensis*. The mesiobuccal roots were almost vertically inserted into the maxillae (Figure 1).

The data for the 109/209 tooth lengths, tooth root lengths and tooth root thicknesses are summarized in Table 3. In both *B. babyrussa* and *B. celebensis* the four main roots (mesiobuccal, mesiopalatal, distobuccal and distopalatal) had longer tooth and tooth root lengths than those of the mesiomedial, mesial and distomedial roots ($P < .001$). These four main roots in *B. babyrussa* were thicker than those of the mesiomedial, mesial and distomedial roots ($P < .001$). In *B. babyrussa* the mesiopalatal and distopalatal roots were thicker than the corresponding mesiobuccal and distobuccal roots ($P < .001$). In *B. celebensis* the mesiopalatal roots were thicker than the corresponding mesiobuccal roots ($P < .05$), and the distobuccal roots were thicker than the distomedial roots ($P < .01$).

The 309/409 tooth roots were also tapering, rod-like structures (Figure 1B). The majority, 64% of the *B. babyrussa*, and 50% of the *B. celebensis* 309/409 mesiobuccal and distobuccal roots inserted almost vertically into the bone (Figure 1B). The data for the 309/409 tooth lengths, tooth root lengths and tooth root thicknesses are summarized in Table 4. In both species the central tooth and root lengths were shorter,

and in *B. babyrussa* thinner, than each of the 4 main roots ($P < .001$). In *B. babyrussa* the tooth lengths of the four main roots were not significantly different from one another. However, the mesiolingual roots were longer than those of the other three main roots ($P < .01$) and thicker than the mesiobuccal roots ($P < .001$). In *B. celebensis* the mesiolingual tooth and root lengths were longer than those of the other three main roots ($P < .001$). The thickness of the roots of these teeth was not significantly different from one root to another. There were no 309/409 buccal or lingual roots in either species.

The 110/210 tooth roots were tapering, rod-like structures that were somewhat blunt-ended (Figure 1B). A slight distal tilt from vertical of the distobuccal root was also seen in the 111/211 tooth roots, where 72% of the *B. babyrussa* and 67% of the *B. celebensis* distobuccal roots inserted at an angle distally (Figure 1B). The variety of patterns of oval and closed 'C' roots in these teeth was note-worthy, and in particular the open 'C' shaped distopalatal roots (Figure 4). The data for the 110/210 teeth are summarized in Table 3. In both species the 4 main roots had similar lengths but shorter mesial tooth lengths ($P < .001$). In *B. babyrussa*, the mesiopalatal root was thicker than the other mesial roots ($P < .001$), and the diagonally opposite distobuccal root was thicker than the other distal roots ($P < .001$). The mesiopalatal and mesial root thickness of the *B. babyrussa* were larger than the corresponding distopalatal and distal roots ($P < .001$), whereas the converse was true for the mesiobuccal and distobuccal root thickness ($P < .001$). Only one central root was found in *B. babyrussa* (Table 3). In *B. celebensis* the mesiopalatal and distobuccal root thicknesses are greater than that of the distopalatal root ($P < .01$).

The 310/410 tooth roots were rod-like structures. In both species, 67% of the 310/410 mesiobuccal and distobuccal roots inserted almost vertically into the bone (Figure 1B). Many of the 310/410 roots showed semi-closed or open 'C' shaped structures (Figure 5B). In addition, 29 (80.6%) of the *B. babyrussa* and 10 (71.4%) of the *B. celebensis* 310/410 teeth had a ridge of dentine, orientated mesiodistally, descending from a central position into the bone (Figure 5). Comparable ridges were less frequently seen in 309/409 teeth (in 16 (44.4%) *B. babyrussa* and in 4 (28.6%) *B. celebensis*) and in 109/209 teeth (in 10 (27.8%) *B. babyrussa* and 5 (35.7%) *B. celebensis*) and 110/210 teeth (in 6 (16.7%) *B. babyrussa* and in 8 (57.1%) *B. celebensis*). Underlying ridges of

tooth dentine were also seen in a central position in 111/211 teeth (in 19 (52.8%) of the *B. babyrussa* and in 10 (62.5%) of the *B. celebensis*) and to a lesser extent in 311/411 teeth (in 11 (30.6%) of the *B. babyrussa* and in 2 (12.5%) of the *B. celebensis*).

The measurement data for the 310/410 teeth are summarized in Table 4. In both species the mesiolingual tooth and root measurements were longer than those of the other roots ($P < .001$). In *B. babyrussa* the distolingual roots were thicker than the other roots ($P < .001$). In both species the central roots were about half the length of the 4 main roots ($P < .001$). The central roots were also thinner than the 4 main roots ($P < .001$). There were no 310/410 buccal or lingual roots in either species.

The structures of 111/211 tooth roots were complex. The robust and tapering rod-like mesiobuccal roots inserted almost vertically into the maxilla when viewed laterally (Figure 6). The distobuccal roots, also robust and tapering rods, were more variable with some inserting almost vertically into the maxilla (Figure 6B) and others orientated in a more distal direction (Figures 6A,C,D). The distobuccal, distal and distopalatal roots were orientated distally into the maxilla (Figures 6 & 7A&B). The mesial and distal roots were splayed in appearance when viewed in the longitudinal section (Figures 7D & E). The cross-sectional shapes of the mesial and distal roots were oval, the latter appearing more elongated in a mesiodistal direction. The cross-sectional shapes of these distal roots were variable and difficult to define. The apices of the distal roots were usually indistinct (Figures 6CD, 7ABC) making systematic measurement unreliable. The measurable data for the 111/211 teeth are summarized in Table 3. In *B. babyrussa* the mesiobuccal tooth lengths were longer than the mesial and mesiopalatal lengths, and the distobuccal tooth lengths were longer than the distal and distopalatal lengths ($P < .001$). However, the mesial tooth root lengths were longer than those of the mesiobuccal and mesiopalatal roots ($P < .05$). The distal root lengths were longer than those of the distopalatal roots ($P < .001$). The *B. babyrussa* 111/211 teeth had no central roots (Table 3). In *B. celebensis* the mesiobuccal root was longer than the mesiopalatal root and the distal root was longer than both distobuccal and distopalatal roots ($P < .001$) as well as the central and mesial roots ($P < .05$). In both species the mesiopalatal root thickness was greater than those of the mesial and distopalatal tooth

roots ($P < .001$), and the distobuccal root thickness was greater than those of the distopalatal and distal roots ($P < .05$).

The 311/411 tooth roots retained elements of the open 'C' shape (Figure 8A) and their insertions were generally blunt-ended and orientated distally (Figures 8B,D,E). The cross-sectional shapes of these roots were generally elongated oval (Figure 8A) and the apices were often bowed (Figure 8B) or splayed lingually and buccally (Figure 8C) into the bony matrix. The shapes of the distal roots were rod-like with distobuccal and distolingual roots being identified (Table 4). The data for the 311/411 teeth are summarized in Table 4.

Within the two species, the lingual, distolingual and distobuccal teeth have similar tooth lengths. In *B. babyrussa* the mesiolingual tooth and root lengths are greater than that of the mesiobuccal ($P < .001$). The central root is less than half the lengths of the other roots; its root thickness is also thinner than those of the mesobuccal and mesolingual roots ($P < .001$). Buccal and lingual roots are thicker than distobuccal and distolingual roots ($P < .001$).

In *B. celebensis* the mesiolingual tooth and root lengths are longer than those of the mesiobuccal roots ($P < .01$). There are no significant differences in tooth and root lengths between the other roots. However, the mesiolingual roots were thicker than lingual and distolingual roots ($P < .01$). Central roots were thinner ($P < .01$).

Root canal system

The pulp chamber of the 109/209 tooth was arched slightly in 4 places to fit under the thick dentine layer which itself was raised to fit under the 4 molar cusps; paracone, protocone, metacone and tetracone (Figures 2A & 2C). In each of the maxillary molar teeth the floor of the pulp chamber rested quite flat on the overlying dentine layer. The number of pulp canals detected in each 109/209 tooth root was 1. However, *B. babyrussa* skulls had 2 pulp canals in two mesiobuccal roots and in three distopalatal roots. In *B. celebensis* skulls, 2 pulp canals were detected in each of one mesiopalatal root, two distobuccal roots and five distopalatal roots, and 3 pulp canals were detected in one distopalatal root.

The pulp chamber of the 110/210 tooth was more obviously arched in 4 places to fit under the thick dentine layer which itself was raised to fit under the 4 main molar cusps (Figures 2A & 2C). The number of pulp canals detected in each 110/210 tooth root was 1. However, *B. babyrussa* skulls had 2 pulp canals in eleven distopalatal roots, 3 in eleven distopalatal roots and 4 in four distopalatal roots. In *B. celebensis* skulls, 2 pulp canals were detected in each of two mesiopalatal roots, one distobuccal root and four distopalatal roots; 3 pulp canals were found in eight distopalatal roots; 4 pulp canals were found in one distopalatal root.

The pulp chamber of the (111/211) tooth was similar to that of the (110/210) tooth in that the pyramidal arching under each of the 4 main molar cusps (Figures 2A & 2C) was clearly defined (Figures 7A & 7D). The number of pulp canals detected in each 111/211 tooth root was generally 1. However, *B. babyrussa* skulls had 2 pulp canals in each of five mesiopalatal roots, two distobuccal roots, one distopalatal root, one additional buccal associated distal root and four palatal associated distal roots; 3 pulp canals were found in one mesiopalatal root. In *B. celebensis*, 2 pulp canals were detected in each of nine mesiopalatal roots, four distobuccal roots, four distopalatal roots and two palatal associated distal roots; 3 pulp canals were found in one mesiopalatal root and in three palatal associated distal roots.

The pulp chamber of the 309/409 tooth was arched slightly in 4 places to fit under the thick dentine layer which itself was raised to fit under the 4 main molar cusps (Figures 2B & 2D). In each of the mandibular molar teeth the floor of the pulp chamber rested quite flat on the underlying dentine layer. The number of pulp canals detected in each *B. babyrussa* 309/409 tooth root was 1. In *B. celebensis* each tooth root contained 1 pulp canal except for 2 of them found in two mesial roots.

The pulp chamber of the 310/410 tooth was more obviously arched in 4 places to fit under the thick dentine layer (Figures 5C & 5D), which itself was raised to fit under the 4 main molar cusps (Figs 2B & 2D). The number of pulp canals detected in each 310/410 tooth root was 1 with the exception in *B. babyrussa* skulls where 2 were found in two distolingual roots. In *B. celebensis* each tooth root contained 1 pulp canal with the exception of 2 pulp canals found in each of two distobuccal roots and nine distolingual roots; 3 pulp canals were found in one distolingual root.

The pulp chamber of the 311/411 tooth was somewhat similar to that of the 310/410 tooth in that the pyramidal arching under each of the main molar cusps (Figures 2B & 2D) was clearly defined. The dentine layer overlying it was relatively thick (Figures 8B, 8C & 8E). The number of pulp canals detected in each 311/411 tooth root was 1. However, *B. babyrussa* skulls revealed 2 in each of one central root, twenty-six buccal roots, twenty-one lingual roots, nine distobuccal roots and six distolingual roots; 3 pulp canals were found in four buccal roots, as well as in three lingual roots and two distobuccal roots. In *B. celebensis* each tooth root contained 1 pulp canal. However, 2 pulp canals were found in each of one central root, seven buccal roots, nine lingual roots, six distobuccal roots, and one distolingual root; 3 pulp canals were found in one buccal root and in one distobuccal root; 5 pulp canals were found in one distobuccal root.

Discussion

Recent studies have described the deciduous and permanent premolar teeth of babirusa (*Babyrousa* spp.).^{12,15} The current investigation is the first since Stehlin^{5,16} to have closely examined the structure of the maxillary and mandibular molar teeth in the adult babirusa (*Babyrousa* spp.). It has extended, by illustration and measurement, his observed descriptions of those teeth and has provided additional detail regarding the anatomical variation of their roots. It has confirmed that for both species of *Babyrousa* the roots of these teeth are more complex than simple rod-like structures (Figures 7 & 8). It has also quantified the various numbers of roots associated with each molar tooth (Table 2).

Studies of *Sus domesticus* by Habermehl¹⁷ and wild *Sus scrofa* by Gabriel¹⁸ have made available detailed comparative information on the numbers of tooth roots and their frequency of occurrence in these species. Additional tooth root information was provided for the mandible of the 'Clawn' strain of miniature Japanese *Sus domesticus*¹⁹ and the European wild pig, *Sus scrofa*.²⁰ During development the first and second mandibular molar teeth of *Phacochoerus africanus* each receive 4 separate roots which are thereafter completely eroded.¹⁶

One possible explanation for the relatively short lengths of the first molar roots of the babirusa may be that these permanent teeth erupt relatively early (~6-8 months) when the depths of bony support provided by the growing maxilla and mandible are structurally shallow.¹⁵ In *Sus scrofa* these first permanent teeth, first maxillary and mandibular molars, are usually present from about 5 months of age.²¹ In babirusa the first molar teeth provide functional masticatory support to the 3 upper and lower deciduous cheek teeth until about 20-24 months of age. In *Sus domesticus* the maxillary and mandibular first molar teeth are present before the replacement of the deciduous teeth by the permanent premolar teeth.²² At about 20-24 months of age the deciduous cheek teeth of the babirusa are replaced by 2 relatively long-rooted upper and lower permanent premolar teeth. By this time the second, long-rooted, molar teeth had taken up their positions.^{12,15} The third molar teeth fully erupt early in the third year, at the end of the pre-adult stage of life.²³

It is likely that relative differences between the upper and lower jaw bones with regard to the amount of structural bone available to support molar tooth roots contribute to the observed differences in tooth root lengths. In *Sus domesticus* the densities of the bone in the maxilla and mandible differ significantly, being lower in the maxilla.^{24,25} This may partly explain why the maxillary molar teeth of babirusa possess more roots than those of the mandible; to increase the stability of maxillary teeth in bone of lower density. The tooth root transmits and balances occlusal forces through the periodontium to the alveolar bone.²⁶ In the babirusa the cheek tooth wear seen⁶ suggested that the first molar teeth had been taking most of the chewing activity with less wear seen in the second and third molar teeth until later in the life of the animal. This pattern was different from that shown for *Sus scrofa*.^{27,28,29} In *Sus scrofa* there was a relative evenness of progressive wear shown by the first 2 molar teeth and a lot more wear of the third molar tooth as that suited aged. The babirusa's mandibular molar teeth appeared to be rooted to function in part as 'anvils' for the breaking and crushing of fruits and nuts.³⁰ The relative pattern of wear of the occlusal surfaces of the mandibular teeth when compared with those of the maxilla lends support to this hypothesis.^{6,10} The slight splaying of the babirusa tooth roots would contribute towards support of the molar teeth. Both male and female babirusa in zoological collections have been observed to use the molar teeth to crack open the very hard Kenari (*Canarium*

amboinense Hochr.) nutshell and then consume the nut contained within (Leus and Macdonald, 1995, unpublished observations).

The distal orientation of the distal roots of the third mandibular molar teeth was observed in the results of the radiographic study on the Large White breed of pig.²¹ This was also seen in Japanese wild pig (*Sus scrofa leucomystax*).³¹ In this arrangement it appeared to be acting as a 'brace' to the tooth to prevent the occlusal surface from being pushed distally. Some preliminary observations of mandibular yaw movements by the feeding babirusa have been recorded.³² The dynamics of mastication have been examined in more detail in *Sus domesticus*^{33,34,35}, and subsequent research has shown the findings to have wider mammalian application.³⁶ The closure of the jaw was primarily due to the activity of the masseter and pterygoid muscles on the non-chewing side, and the temporalis muscle on the chewing side. The masseter muscles increased the crushing force on the food bolus, which was positioned in a slightly lateral position, but they also then contributed to the movement of the bolus over the mandible lingually. The anatomy of the cranial muscles supporting babirusa jaw movements has been described.³⁷ Further studies of their function during mastication are required.

Despite the considerable amount of work in vitro and in vivo with mice and rats³⁸, it remains unknown what determines the number of roots per tooth and the direction of root formation, although they are likely to be influenced by the Hertwig's epithelial root sheath (HERS). Research results indicate that an array of growth and transcription factors was expressed during the initiation of root formation, and these activators and inhibitors appear to work together to achieve a balanced signalling outcome and produce the proper patterning, number and length of dental roots during later stages of tooth morphogenesis.³⁸ There is some evidence that the cervical tongue, a tongue-shaped epithelial process extending from the tooth's cervical loop, is involved.³⁹ In rodent studies, the elongation and contact between these cervical tongues form discrete regions which resulted in the furcation zone of roots. As a consequence, it has been suggested that the pattern of cervical tongues plays an important role in the determination of the numbers, lengths, and shapes of the tooth roots.^{39,40} Despite these and other findings, the overall mechanisms regulating dental root formation remain poorly understood.

Interspecies tooth root comparisons have been made for primates, bats and carnivores.^{41,42,43} In carnivore species, those that feed on hard food objects have larger tooth roots than those that eat soft or tough foods.⁴³ In bats with diets of different hardness there was a linear relationship between root size and crown size, indicating that the roots were not expanded disproportionately; instead, the entire tooth was larger in the hard diet species.⁴⁴ Given that the babirusa on the northern peninsula of Sulawesi seem to have larger 311/411 teeth than those from the Togian Islands, and that a proportion of these Sulawesi babirusa appear to be attracted to the banana (*Musa balbisiana*) which has as many as 150-170 hard seeds within it, perhaps a lengthy dietary relationship between (*Musa balbisiana*) and babirusa has contributed towards relative molar tooth size.¹⁰ The babirusa on the Togian islands are thought to have reached there from east central Sulawesi where this species of banana does not appear to grow.^{45,46}

Conclusions

The relative structure of the molar teeth in babirusa suggests that the main role they play is food grinding and crushing. The first molar teeth appear to provide most of the functional masticatory support in this way to the three upper and lower deciduous cheek teeth until about 20-24 months of age. Their small roots appear to be related to the shallow availability of bony material in the jaws of young babirusa. Differences in bone density and the requirement for tooth stability may partly explain why the maxillary molar teeth of babirusa possess more roots than those of the mandible. The slight splaying of the babirusa tooth roots would contribute towards support of the tooth. The rooting pattern of babirusa mandibular molar teeth appeared to function in support of the teeth acting as 'anvils' for the breaking and crushing of fruits and nuts.

Materials

- a. Senckenberg Naturhistorische Sammlungen Dresden, Germany (SNSD)
- b. National Museums Scotland, Edinburgh, Scotland (NMS)
- c. Naturmuseum Senckenberg, Frankfurt am Main, Germany (SMF)

- d. Göteborgs naturhistoriska museum, Göteborg, Sweden (GNM)
- e. The University of Edinburgh: Helical scan mode, collimator pitch of 1, 0.4 mm slice width, 0.2 mm slice interval, 120 kVp, 111 mAs, ultra-high resolution bone image reconstruction kernel (proprietary name *U75u*), 192 mm diameter display field of view, 512² image matrix (64-slice CT, Somatom® Definition AS Siemens, Erlangen, Germany).
- f. The University of Giessen: Helical scan mode, collimator pitch of 0.75, 0.65 mm slice width, 0.32 mm slice interval, 140 kVp, 190 mAs, bone image reconstruction kernel (proprietary name *D*), 176 mm diameter display field of view, 512² image matrix (16-slice CT, Brilliance® 16, Philips Medical Systems, Eindhoven, the Netherlands).
- g. The University of Leipzig: Helical scan mode, collimator pitch of 0.75, 0.7 mm slice width, 0.35 mm slice interval, 140 kVp, 193 mAs, bone image reconstruction kernel (proprietary name *D*), 100mm diameter display field of view, 512² image matrix, g. (6-slice CT, Brilliance® 6, Philips Medical Systems, Eindhoven, the Netherlands).
- h. Blåstjärnans Djursjukhus Göteborg: Helical scan mode, collimator pitch of 0.53125, 0.625 mm slice width, 0.312 mm slice interval, 120 kVp, 200 mAs, bone image reconstruction kernel (proprietary name *Bone Plus*), 182 mm diameter display field of view, 512² image matrix (16-slice CT, GE Revolution® EVO CT, GE Healthcare Japan Corporation, Tokyo, Japan).
- i. JASP Team (2020) JASP (Version 0.14.1)

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors received no financial support for the research, authorship and/or publication of this article.

Acknowledgements

We are thankful for the kind hospitality and assistance provided by Clara Stefen and Jens Jakobitz of the Senckenberg Naturhistorische Sammlungen Dresden, Germany, to Irina Ruf and Katrin Krohmann of the Senckenberg Forschungsinstitut und Naturmuseum Frankfurt am Main, Germany and to Ines Merseburger of the Universität Leipzig, Veterinärmedizinische Fakultät, Klinik für Kleintiere, for her technical assistance. We are also grateful for the technical support and access to the mammal collections in each of the National Museums Scotland, Edinburgh, Scotland, the Senckenberg Naturhistorische Sammlungen Dresden, Germany, the Senckenberg Forschungsinstitut und Naturmuseum Frankfurt am Main, Germany and the Göteborgs naturhistoriska museum, Göteborg, Sweden. The assistance of Masaaki Ito with respect to the Japanese literature was also much appreciated. We are grateful to the University of Edinburgh and the Balloch Trust for academic support.

References

- 1 **Macdonald AA**. 2018. Sulawesi Babirusa *Babyrousa celebensis* (Deninger, 1909). Chapter 6, In **Melletti M, Meijaard E**, eds. Ecology, Evolution and Management of Wild Pigs and Peccaries. Implications for Conservation. Cambridge; Cambridge University Press; 2018:59-69.
- 2 **Sheherazade Hesdianti E, Indrawan M**. Moluccan babirusa *Babyrousa babyrussa* (Linnaeus, 1758). In: **Melletti M, Meijaard E**, eds. Ecology, Evolution and Management of Wild Pigs and Peccaries. Implications for Conservation. Cambridge; Cambridge University Press; 2018:70-75.
- 3 **Cuvier F**. Des Dents des mammifères considérées comme caractères zoologiques. Strasbourg: F.G. Levrault. 1825.

- 4 **Giebel CG.** Odontographie. Leipzig: Verlag von Ambrosius Abel. 1855.
<https://www.biodiversitylibrary.org/item/52039#page/11/mode/1up> Accessed 24 02 2021.
- 5 **Stehlin HG.** Ueber die Geschichte des Suidae Gebisses. Zweiter Teil Abh Schweizer palaontolog Gesell. 1900:27:337–527. Available from <https://archive.org/stream/StehlinH.g.1899-UeberDieGeschichteDesSuiden-gebisses/StehlinH.g.1900-UeberDieGeschichteDesSuiden-gebissesPart2#page/n183/mode/2up> [accessed 24 February 2021].
- 6 **Macdonald AA.** Cheek tooth erosion in male babirusa (genus *Babyrousa*). C R Biologies. 2019:342(5–6):199–208. doi:10.1016/j.crv.2019.07.005. PMID:31474523.
- 7 **Leus K, Goodall GP, Macdonald AA.** Anatomy and histology of the babirusa (*Babyrousa babyrussa*) stomach. C R de l'Acad des Sci, Série III - Sciences de la Vie. 1999:322:1081-1092.
- 8 **Leus K, Macdonald AA, Goodall G, Veitch D, Mitchell S, Bauwens L.** Light and electron microscopy of the cardiac gland region of the stomach of the babirusa (*Babyrousa babyrussa* – Suidae, Mammalia). C R Biologies. 2004:327:735-743.
- 9 **Macdonald AA, Mitchell S, Signorella A, Leus K.** Ultrastructural characterization of the epithelium that constitutes the cardiac gland epithelium 'honeycomb' in the stomach of the babirusa (*Babyrousa babyrussa*). C R Biologies. 2008:331:32-41.
- 10 **Macdonald AA.** Anomalous erosion patterns on the cheek teeth of babirusa (*Babyrousa* Perry 1811.). Can J Zool. 2021a:99:1-8.
- 11 **Floyd MR.** The modified Triadan system: nomenclature for veterinary dentistry. J Vet Dent. 1991:8:18-19.
- 12 **Macdonald AA, Ziehmer B, Kitchener AC, Gelang M, Åblad B, Lintonsson R, von Pückler K, Schaub S, Kiefer I, Schwarz T.** The premolar teeth of *Babyrousa* spp. – a CT study. J Vet Dent. 2023:40: in press.
- 13 **AVDC.** American Veterinary Dental College: Nomenclature and Numbering of Teeth. <https://avdc.org/avdc-nomenclature/> Accessed 30 March, 2021.

- 14 **van der Made J.** Listriodontinae (Suidae, Mammalia), their evolution, systematics and distribution in time and space. *Contrib Tert Quatern Geol.* 1996:33(1-4):3-254. Available from <http://natuurtijdschriften.nl/record/521620> [accessed 26 July 2020].
- 15 **Macdonald AA.** Molar tooth emergence in *Babyrousa* spp. *Suiform Sound.* 2021b:20:12-19.
- 16 **Stehlin HG.** Ueber die Geschichte des Suidae Gebisses. Erster Teil Abh Schweizer palaontolog Gesell. 1899:26:1-336. Available from <https://archive.org/details/StehlinH.g.1899-UeberDieGeschichteDesSuiden-gebisses/StehlinH.g.1899-UeberDieGeschichteDesSuiden-gebissesPart1/mode/2up> [accessed 24 February 2021].
- 17 **Habermehl KH.** Über das Gebiß des Hausschweines (*Sus scrofa dom.* L.) mit besonderer Berücksichtigung der Backenzahnwurzeln. *Zent Vetmed.* 1957:4:794-810.
- 18 **Gabriel P.** Kopfdarm und Schlund des Wildschweines (exkl. Mundboden).VIII. Beitrag zur Anatomie von *Sus scrofa* L. und zum Domestikationsproblem. *Zeitschr Anat Entwickl.* 1934:102:521-571.
- 19 **Ide Y, Nakahara T, Nasu M, Matsunaga S, Iwanaga T, Tominaga N, Tamaki Y.** Postnatal mandibular cheek tooth development in the miniature pig based on two-dimensional and three-dimensional X-ray analyses. *Anat Rec.* 2013:296:1247-1254.
- 20 **Gubler R.** Die Mundbodenorgane des Wildschweines. *Anat Anz.* 1933:77:129-168.
- 21 **Briedermann L.** Schwarzwild: Neuausgabe bearbeitet von Burkhard Stöcker. Kosmos; Stuttgart. 2009.
- 22 **Tonge CH, McCance RA.** Normal development of the jaws and teeth in pigs, and the delay and malocclusion produced by calorie deficiencies. *J Anat.* 1973:115:1-22.
- 23 **Macdonald AA, Shaw D.** Maxillary tooth growth in the adult male babirusa (genus *Babyrousa*). *C R Biologies.* 2018:341(4):235-244. doi:10.1016/j.crvi.2018.04.002. PMID:29752201.
- 24 **Devlin H, Horner K, Ledgerton D.** A comparison of maxillary and mandibular bone mineral densities. *J Prosthet Dent.* 1998:79:323-327.

- 25 **Verdonck HWD, Meijer GJ, Nieman FH, Stoll, C, Riediger D, de Baat C.** Quantitative computed tomography bone mineral density measurements in irradiated and non-irradiated minipig alveolar bone: an experimental study. *Clin Oral Impl Res.* 2008;19:465–468.
- 26 **Wu X, Hu L, Li Y, Li Y, Wang F, Ma P, Wanh J, Zhang C, Jiang C, Wang S.** SCAPs regulate differentiation of DFSCs during tooth root development in swine. *International J Med Sci.* 2018;15:291-299.
- 27 **Grant A.** The use of tooth wear as a guide to the age of domestic ungulates, in: **Wilson B, Grigson S, Payne S.** eds. *Ageing and sexing animal bones from archaeological sites*, Oxford, British Archaeological Reports, British Series. 1982.
- 28 **Desbiez ALJ, Keuroghlian A.** Ageing feral pigs (*Sus scrofa*) through tooth eruption and wear, *Suiform Sound.* 2011;9:48–55.
- 29 **Lemoine X, Zeder MA, Bishop KJ, Rufolo SJ.** A new system for computing dentition-based age profiles in *Sus scrofa*, *J Archaeol Sci.* 2014;47:179–193.
- 30 **Leus K.** Foraging behaviour, food selection and diet digestion of *Babyrousa babyrussa* (Suidae, Mammalia), Ph.D. thesis, The University of Edinburgh, Edinburgh. 1994.
- 31 **Imoto H.** A morphological study on the cheek tooth of Japanese wild boar (*Sus scrofa leucomystax*). *J Kyushu Dent Soc.* 1977;30:754–796 (in Japanese with English abstract).
- 32 **Ito M, Macdonald AA, Leus K, Balik IW, Arimbawa IWGB, Hasegawa Y, Atmaja IDGA.** Coconut feeding of the babirusa (*Babyrousa* spp.). *Jap J Zoo Wild Med.* 2020; 25(3):91-100.
- 33 **Herring SW, Scapino RP.** Physiology of feeding in miniature pigs, *J Morphol.* 1973;141:427–460.
- 34 **Herring SW.** The dynamics of mastication in pigs, *Arch Oral Biol.* 1976;21:473–480.
- 35 **Menegaz RA, Baier DB, Metzger KA, Herring SW, Brainerd EL.** XROMM analysis of tooth occlusion and temporomandibular joint kinematics during feeding in juvenile miniature pigs, *J Exp Biol.* 2015;218:2573–2584.

- 36 **Grossnickle DM.** The evolutionary origin of jaw yaw in mammals, *Sci Rep.* 2017:7, 45094, <http://dx.doi.org/10.1038/srep45094>.
- 37 **Kneepkens AFLM, Macdonald AA.** Cranial muscles of the Sulawesi Babirusa (*Babyrousa celebensis*). *Anat Histol Embryol.* (DOI: 10.1111/j.1439-0264.2009.00987.x). 2010:39:120-137.
- 38 **Li J, Parada C, Chai Y.** Cellular and molecular mechanisms of tooth root development. *Devel (Camb).* 2017:144:374-384.
- 39 **Seo H, Kim, J, Hwang Jae J, Jeong Ho-Gul, Han Sang-Sun, Park W, Ryu K, Seomun H, Kim J-Y, Cho E-S, Park J-C, Hu K-S, Kim H-J, Kim Dong-Hyun, Cho S-W.** Regulation of root patterns in mammalian teeth. *Sci reps.* 2017:7:12714-13 DOI:10.1038/s41598-017-12745-1
- 40 **Huang XF, Chai Y.** Molecular regulatory mechanism of tooth root development. *Int J Oral Sci.* 2012:4:177–181, <https://doi.org/10.1038/ijos.2012.61>.
- 41 **Emonet E-G, Kullmer O.** Variability in premolar and molar root number in a modern population of *Pan troglodytes verus*. *Anat Rec.* 2014:297:1927-1934.
- 42 **Self CJ.** Cricetid rodents: Is molar root morphology an indicator of diet? *Zoomorph.* 2015a:134(2):309–316.
- 43 **Kupczik K, Stynder DD.** Tooth root morphology as an indicator for dietary specialization in carnivores (Mammalia: Carnivora). *Biol. J Linn Soc.* 2012:105:456–471.
- 44 **Self CJ.** Dental root size in bats with diets of different hardness. *J Morphol.* 2015b:276:1065–1074.
- 45 **Nasution RE, Yamada I.** Pisang-pisang liar di Indonesia. Pusat Penelitian dan Pengem Bangan Biologi, Lipi, Bogor. 2001.
- 46 **Hastuti H, Purnomo P, Sumardi I, Daryono BS.** Diversity wild banana species (*Musa spp.*) in Sulawesi. *Biodiversitas.* 2019:20(3):824–832. doi:10.13057/biodiv/d200328.

Tables

Table 1	<i>Babyrousa babyrussa</i> and <i>Babyrousa celebensis</i> Skulls Studied.
Table 2	<i>Babyrousa babyrussa</i> and <i>Babyrousa celebensis</i> Tooth Roots.
Table 3	Adult Babirusa (<i>Babyrousa babyrussa</i> and <i>Babyrousa celebensis</i>) Maxillary Molar Teeth Measurements.
Table 4	Adult Babirusa (<i>Babyrousa babyrussa</i> and <i>Babyrousa celebensis</i>) Mandibular Molar Teeth Measurements.

Figures

- Figure 1 A. Right lateral view of Sulawesi babirusa (*Babyrousa celebensis*) OUM 05266 illustrating anatomical exposure of the roots of the teeth.
B. Right lateral 3D CT-scan view of roots of the molar and premolar teeth of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728.
- Figure 2 A. Occlusal view of the left maxillary molar teeth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z,2002.210.1. The list of symbols¹⁴ describes the cusps of the maxillary molar teeth, left to right; 209, 210 and 211: 1: paracone; 2: protocone; 3: metacone; 4: tetracone; and 5: pentacone. (bar = 10mm).
B. Occlusal view of the left mandibular molar teeth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z,2002.210.1. The list of symbols¹⁴ describes the cusps of the mandibular molar teeth, left to right: 309, 310 and 311. 1: protoconid; 2: metaconid; 3: hypoconid; 4: entoconid; 5: pentaconid; and 6: hexaconid. (bar = 10mm).
C. 3D CT-scan view of the underside of the enamel layer comprising the occlusal surface of the left maxillary molar teeth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z,2002.210.1.
D. 3D CT-scan view of the underside of the enamel layer comprising the occlusal surface of the left mandibular molar teeth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z,2002.210.1.
- Figure 3 A. Cross-section CT-scan of the 209 tooth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z, 1991.15.5 illustrating the four main roots (mb=mesiobuccal, mp=mesiopalatal, db=distobuccal, dp=distopalatal) and the positions of three additional roots (m=mesial, c=central, d=distal).

B. Photograph of the 209 tooth alveolus of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.934 with openings into the bony matrix corresponding to positions of the roots described in 3A.

C. Cross-section CT-scan of the 309 tooth of Buru babirusa (*Babyrousa babyrussa*) NMS,Z,2001.142 illustrating the four roots (mb=mesiobuccal, ml=mesiolingual, db=distobuccal, dl=distolingual).

D. Lateral 3D CT-scan of the 309 and the protoconid and underlying mesiobuccal root of 310 of Buru or Sula babirusa (*Babyrousa babyrussa*) SMF 19704 DB.

- Figure 4
- A. Cross-sectional CT-scans of the 210 tooth of A. Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728 illustrating the four roots (mb=mesiobuccal, mp=mesiopalatal, db=distobuccal, dp=distopalatal).
- B. Buru or Sula babirusa (*Babyrousa babyrussa*) NMSZ.1991.15.1.
- C. Buru or Sula babirusa (*Babyrousa babyrussa*) NMS,Z,1991.15.5.
- D. Buru babirusa (*Babyrousa babyrussa*) NMSZ.1992.10.28, to illustrate relative positions, sizes and shapes of the four main roots, and the variety of 'C' shapes of the distal palatal roots. Note the presence of additional roots in C and D.

- Figure 5
- A. Cross-sectional CT-scan of the 310 tooth in Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.966 to illustrate the four main roots and the central ridge of bone between these roots.
- B. Photograph of the 310 tooth alveolus of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.730 with openings into the bony matrix corresponding to positions of the roots and the bony ridge described in 5A.
- C. Rostral CT-scan of the 310 tooth in Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.966 to illustrate the bony ridge between the roots.
- D. Lateral CT-scan of the 310 tooth in Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.966 to illustrate the bony ridge between the roots.

- Figure 6
- A. 3D CT-scan view of the 211 tooth roots of Buru babirusa (*Babyrousa babyrussa*) SNSD 040320183 DB to illustrate the distal orientation of the distal and distobuccal roots.
- B. 3D CT-scan view of the 211 tooth roots of Buru babirusa (*Babyrousa babyrussa*) SMF 19706 DB to illustrate the swollen shape and distal orientation of the distal root.
- C. 3D CT-scan view of the 211 tooth roots of Sulawesi babirusa (*Babyrousa celebensis*) NMS,Z,1878.3.1 to illustrate the distal orientation and rod-like appearance of the distal and distobuccal roots.

D. 3D CT-scan view of the 211 tooth roots of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960 to illustrate the distal orientation and splintered appearance of the distal and distobuccal roots.

- Figure 7
- A. Lateral-sectional CT scan of the 211 tooth root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960 to illustrate the complexity of the distal root structure.
 - B. 3D CT-scan view of the 211 tooth roots of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960 seen from a distopalatal viewpoint.
 - C. Cross-sectional CT scan of the 211 tooth root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960.
 - D. Distal view of the longitudinal-sectional CT scan of the 211 distal roots of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960 to illustrate the splayed arrangement of these roots.
 - E. Distal view of the longitudinal-sectional CT scan of the 211 mesiopalatal root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 17.960.

- Figure 8
- A. Cross-sectional CT scan of the left mandibular 311 tooth root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728.
 - B. Mesial-sectional CT scan of the 311 distal tooth roots of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728 to illustrate the mesial orientation of the roots.
 - C. Mesial-sectional CT scan of the 311 distal tooth roots of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728 to illustrate the more distal and splayed orientation of these roots.
 - D. Lateral 3D CT scan of the 311 tooth root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728 to illustrate the shapes and orientation of the (l to r) mesiobuccal, buccal and distobuccal roots.
 - E. Lateral-sectional CT scan of the 311 tooth root of Sulawesi babirusa (*Babyrousa celebensis*) GNM 4.728 to illustrate the shape and distal orientation of the distobuccal root.

Contributors statement (derived from <https://casrai.org/credit/>).

- | | |
|-----|--|
| AAM | Conceptualization, Data curation, Investigation, Formal Analysis, Writing; |
| TS | Conceptualization, Methodology, Data curation, Writing – review & editing |
| BZ | Conceptualization, Investigation, Writing – review & editing |
| AK | Resources, Writing – review & editing |
| MG | Resources, Writing – review & editing |
| BA | Methodology, Data curation, Investigation, Writing – review & editing |

RL Data curation
K von P Methodology, Investigation, Data curation, Writing – review & editing
SS Methodology, Investigation, Data curation, Writing – review & editing
IK Methodology, Data curation, Investigation, Writing – review & editing