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Feline non-erosive immune-mediated polyarthritis: a multicentre, retrospective study of 20 cases (2009-2020)

Citation for published version:

Wootton, F, Glanemann, B, Langley-Hobbs, S, Breheny, C, Fowlie, S, Whitworth, F, Silvestrini, P, Threlfall, A, Sorrell, S & Black, V 2022, 'Feline non-erosive immune-mediated polyarthritis: a multicentre, retrospective study of 20 cases (2009-2020)', *Journal of Feline Medicine and Surgery*, vol. 24, no. 10, 1098612X221107783, pp. E401-E410. <https://doi.org/10.1177/1098612X221107783>

Digital Object Identifier (DOI):

[10.1177/1098612X221107783](https://doi.org/10.1177/1098612X221107783)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Journal of Feline Medicine and Surgery

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1 **Title:**

2 Feline non-erosive immune-mediated polyarthritis: a multicentre, retrospective study of 20 cases (2009-
3 2020)

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17 **Abstract**

18 **Case Series Summary:** Multicentre retrospective case series. Cats with non-erosive IMPA were identified
19 from seven referral hospitals between 2009 - 2020. Data was obtained from hospital records and referring
20 veterinarians were contacted for follow-up. Twenty cases were identified; there were 12 neutered males
21 (60%), one entire male (5%) and seven neutered females (35%). Common clinical signs included lameness
22 (20/20) and pyrexia (10/18). Three cats presented with and two cats developed ligament laxity during
23 treatment.

24 Thirteen cats (65%) were diagnosed with non-associative IMPA and seven cats (35%) with associative IMPA.
25 Comorbidities identified included: chronic enteropathy (3/6), FIV (1/6), feline herpes virus (1/6),
26 bronchopneumonia (1/6) and discospondylitis (1/6). Sampling of the tarsal joints most frequently identified
27 an increased proportion of neutrophils consistent with IMPA.

28 Eighteen cats (90%) received immunosuppressants. Eleven cats were started on prednisolone, eight had a
29 poor response resulting in addition of second agent, euthanasia or acceptance of persisting signs. One cat
30 received ciclosporin and required an alternative second agent due to adverse effects. Five cats were started
31 on prednisolone and ciclosporin, three had a poor response and required an alternative second agent. One
32 cat received prednisolone and chlorambucil and had a good response. Two cats (10%) received meloxicam
33 and had a good response although clinical signs recurred when medication was tapered.

34 A good outcome was achieved in 14/20 cats (70%) with IMPA. In the cats with a poor outcome 4/6 were
35 euthanised and 2/6 had chronic lameness.

36 **Relevance and Novel Information:** Prognosis for feline IMPA can be good. Multimodal immunosuppression
37 was often required. IMPA should be considered in lame cats, with or without pyrexia when there is no
38 evidence of trauma or infection. The tarsal joints should be included in the multiple joints chosen for
39 sampling. Ligament laxity can occur in non-erosive feline IMPA.

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41 **Keywords:** immune-mediated polyarthritis, non-erosive arthritis, ligament laxity, arthrocentesis,
42 polyarthritis

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89 **Introduction**

90 Immune mediated polyarthritis (IMPA) is a rarely reported condition in cats compared to dogs.¹ It is
91 diagnosed based on the identification of sterile, neutrophilic inflammation in multiple joints and is defined
92 as erosive or non-erosive based on radiographic findings.² IMPA is caused by an aberrant immune response,
93 either targeted at the joints directly or occurring as a result of the deposition of circulating immune
94 complexes within the joints.¹ Non-erosive IMPA can be split into two main subcategories: associative and
95 non-associative. The terms associative and non-associative address the limitations of previous categorization
96 models as it reflects the spectrum of disease and our incomplete understanding of the pathogenesis of
97 immune mediated disorders.³ Cases in which significant comorbidities are identified during investigations
98 are categorized as associative. In associative IMPA, it is considered possible that the comorbidity identified is
99 the underlying cause. However, the comorbidity could be incidental and not the primary cause of IMPA.
100 Cases in which no comorbidities are identified, are categorized as non-associative IMPA. In non-associative
101 IMPA, although no comorbidity was identified in the investigations performed, it does not exclude the
102 possibility of an underlying cause being present that was either not understood or not detected in the
103 investigations performed. IMPA has previously been associated with pneumonia, pyelonephritis,
104 toxoplasmosis, feline leukemia virus (FeLV), feline infectious peritonitis (FIP) and myelodysplastic
105 disorders.^{4,5}

106 There is little published information about the clinical findings in cats with non-erosive IMPA. To the
107 authors' knowledge, at the time of submission, there has only been one previous case series including

108 thirteen cats with non-erosive IMPA. The cats in that study were identified at two referral centers in the UK
109 between 1977 and 1986.⁴

110 The aim of this case series was to describe the clinical features of associative and non-associative IMPA in
111 cats as well as the investigations performed, treatment and outcome.

112 **Case Series Summary**

113 Ethical approval was granted by the University of Bristol Animal Welfare and Ethics Review Body (veterinary
114 investigation number [VIN] 18/068), the Royal Veterinary College (RVC) Social Sciences Research Ethical
115 Review Board (reference URN SR2019-0482) and the University of Liverpool Veterinary Research Ethics
116 Committee (reference VREC736).

117 Cases were recruited from seven referral hospitals in the United Kingdom from January 2009 to April 2020.
118 Electronic medical records were searched to identify cats diagnosed with IMPA. Cats were included for
119 analysis if they met the following criteria: complete clinical records, no evidence of erosive lesions on CT
120 and/or radiographs of at least one affected joint, an increased proportion of non-degenerate neutrophils
121 (>10% of nucleated cells present) in the synovial fluid of two or more joints with no evidence of intracellular
122 or extracellular bacteria and a minimum of two months follow up.¹ Twenty cats met the inclusion criteria.

123 The following details were extracted from the medical records of all cats: signalment, presenting clinical
124 signs, onset and duration of clinical signs, treatment prior to referral, initial physical examination findings,
125 results of diagnostic investigations, final diagnosis, treatment and outcome.

126 Information regarding outcome was recorded from referral hospital follow up appointments and/or
127 communication records or via telephone updates with the primary care practice. If a patient had died the
128 reason for euthanasia or cause of death was recorded. Outcome was considered good if clinical signs of
129 IMPA were completely controlled whilst on medication without significant side effects, if medications could
130 be tapered without a relapse in clinical signs, if death occurred for reasons unrelated to IMPA or if a relapse
131 occurred but was successfully managed with treatment and clinical signs resolved. Outcome was considered
132 poor in cats whose clinical signs of IMPA were not able to be controlled with treatment or if death was
133 considered to be related to IMPA or treatment. Relapse was defined as a recurrence in clinical signs relating
134 to IMPA.

135 Statistical analysis was performed using SPSS v26 (IBM Corp). Normality was assessed for continuous
136 variables with the Shapiro Wilk test. Results were reported as mean with standard deviation if normally
137 distributed and as medians with range if they were not normally distributed.

138 *Signalment:*

139 The mean age of included cats was 8.8 years (Standard deviation +/- 4.78 years). The median body weight
140 was 4kg (range 2.11 – 8.8kg). The most common breed was the Domestic, 13 cats [65%] (11 [55%] Domestic
141 Shorthair and two [10%] Domestic Longhair). The remaining cats included three Maine Coons (15%), and
142 one each (5%) of the following: Ragdoll, Siamese, Persian cross and Bengal cross. There was a male
143 predominance, with 12 neutered males (60%), one entire male (5%) and seven neutered females (35%). As
144 the breed and sex distribution for each hospital included in this study is unknown, it is not possible to
145 confirm that males or Maine Coon cats were significantly overrepresented.

146 *History and treatment before referral:*

147 A summary of presenting signs is shown in Table 1. Twelve cats (60%) had acute onset of clinical signs
148 (<24hrs) and the remaining eight cats (40%) had a more insidious onset of signs (range two weeks to one
149 year). The most common presenting sign was lameness, reported in all cats. In seven cats (35%) the
150 lameness was present in a single limb and three cats (15%) were non-ambulatory. Rectal temperature at the
151 onset of clinical signs was recorded in 18 cases. Ten cats (55%) were pyrexia (Temperature >39.2C⁶), seven
152 cats (39%) were normothermic and one cat (6%) was hypothermic.

Clinical sign	Number affected (/20)
Lameness	20
Pyrexia	10
Inappetence	8
Reluctance to move	6
Lethargy	4

153 Table 1 – Summary of presenting clinical signs in twenty cats with immune mediated polyarthritis

154 Treatment prior to referral was highly variable. Eighteen cats received treatment prior to referral; treatment
155 data was unavailable for one cat and one cat was referred without any treatment other than fluid
156 resuscitation. The most common treatments were NSAIDs, administered to 13 cats (68%) and antimicrobials
157 administered to 11 cats (58%). Opioid analgesia was administered to nine cats (47%). Corticosteroids were
158 administered to three (16%), gastro-protectants to one (5%) and codeine to one (5%).

159 *Diagnostic tests*

160 Complete blood count (CBC), serum biochemistry, joint fluid cytology, abdominal imaging and joint imaging
161 were performed in every cat. Thoracic imaging was performed in 18/20 cats. Other specific diagnostic tests
162 are documented in Table 2 and the imaging modalities used are summarised in Table 3.

163 The most common abnormal findings on CBC were neutrophilia in 10 cats (6 non-associative and 4
164 associative) and lymphopenia in six cats (4 non-associative and 2 associative), three cats (2 non-associative
165 and 1 associative) had a mild anaemia (packed cell volume > 20 and $\leq 24\%$) with a reference interval (RI) 24 -
166 45 %. The most common serum biochemistry abnormalities were hyperglobulinemia in six (5 non-associative
167 and 1 associative) and hypoalbuminemia in five (3 non-associative and two associative). Of the six cats with
168 hyperglobulinemia, the elevation was mild [46 to ≤ 50 g/L] in four, moderate [50 to < 55 g/L] in one and
169 severe [>55 g/L] in one with a RI 25 - 45 g/l. Of the five cats with hypoalbuminemia this was mild [20 to \leq
170 26 g/L] in four and moderate [15 to < 20 g/L] in one with a RI 25 - 45 g/l.

171 *Classification of cases*

172 Cats were split into two subcategories of IMPA; associative or non-associative based on the final diagnosis
173 made by the clinician. Thirteen cats (65%) were presumed to have non-associative IMPA. Comorbidities
174 were identified in the remaining seven cats (35%) and these were categorized as associative IMPA. The
175 comorbidities identified in the seven cats with associative IMPA included; gastrointestinal disease (3), FIV
176 (1), chronic feline herpes virus (1) bacterial bronchopneumonia (1) and discospondylitis (1).

Diagnostic test	Number of cats in which the test was performed (/20)
Complete Blood Count	20 (100%)
Biochemistry	20 (100%)
Urinalysis	14 (70%)

Urine culture	11 (55%)
FIV/FeLV testing	14 (70%)
Specific Feline Pancreatic Lipase	6 (30%)
Antinuclear antibody (ANA)	1 (5%)
Rheumatoid factor	1 (5%)
Blood culture	1 (5%)
Cobalamin (B12) and folate	5 (25%)
Joint fluid cytology	20 (100%)
Joint fluid culture	9 (45%)
Faecal tests (culture, parasitology)	5 (25%)
PCR (blood, tissue, joint fluid, saliva)	Feline calicivirus: 6 (30%) <i>Mycoplasma felis</i> : 11 (55%) Feline herpesvirus: 3 (15%) Feline Chlamydia: 1 (5%) <i>Borrelia</i> : 6 (30%) <i>Bartonella henselae</i> : 4 (20%) Anaplasma: 2 (10%) <i>Mycobacterium</i> species: 1 (5%) <i>Toxoplasma gondii</i> : 2 (10%) <i>Tritrichomonas foetus</i> : 1 (5%)
Serology	<i>Toxoplasma gondii</i> : 9 (45%) Feline Coronavirus: 2 (10%)

177 Table 2 – Diagnostic tests performed in twenty cats diagnosed with immune mediated polyarthritis

178 Footnote: FIV= feline immunodeficiency virus, FeLV = feline leukemia virus

179

Imaging performed	Number of cats in which the test was performed (/20)	Imaging modality used
Thoracic Imaging	18 (90%)	Radiographs: 12 (60%) CT: 6 (30%)
Abdominal Imaging	20 (100%)	Ultrasound alone: 12 (60%) CT: 5 (25%) Ultrasound and radiographs: 3 (15%)
Joint Imaging	20 (100%)	Radiographs: 17 (85%) CT: 3 (15%)

Echocardiography	12 (60%)	N/A
MRI	1 (5%)	N/A
Endoscopy	1 (5%)	N/A
Bronchoscopy	1 (5%)	N/A

180 Table 3 – Diagnostic imaging performed in twenty cats diagnosed with immune mediated polyarthritis

181 *Joints affected*

182 All cats had an increased proportion of non-degenerate neutrophils identified on cytology of \geq two joints.

183 Neutrophils made up > 10% of the total nucleated cell count on cytology in all affected joints.¹ No infectious
184 organisms were identified on cytology or culture of any joint aspirates. However, only nine cats had joint
185 fluid culture performed. Total nucleated cell counts were not available for all samples due to the small
186 volume of synovial fluid obtained in most cases.

187 The median number of joints sampled was four joints with a range of 2 - 10 joints. The joints sampled most
188 commonly were the carpus (15 cats [75%]), stifle (15 cats [75%]) and tarsus (14 cats [70%]). The elbow was
189 sampled in eight cats (40%) and the shoulder was sampled in one cat (5%). All tarsal joints that were
190 aspirated had an increased proportion of non-degenerate neutrophils identified. The proportion of joint
191 aspirates with findings suggestive of IMPA (neutrophils > 10% of the nucleated cell count and no intra or
192 extracellular bacteria) obtained in this cohort is outlined in Figure 1.

193 Three cats (15%) presented with ligament laxity and two cats (10%) developed ligament laxity five months
194 and 12 months into treatment with corticosteroids. Three of the five cats had tarsal joint laxity resulting in a
195 plantigrade stance, and two cats had bilateral carpal laxity and a palmigrade stance (Figure 2). Radiographs
196 were obtained in 4/5 cats after the development of joint laxity and there were no erosive lesions in any of

197 the affected joints. The timing of joint imaging in relation to the onset of joint laxity was variable (2 weeks, 5
198 weeks, 1 month and 3 months). As it can take several weeks for erosive lesions to become visible on
199 radiographs, it is not possible to definitively prove erosive lesions did not develop in the cats imaged. Two
200 cats developed joint laxity whilst on prednisolone after 5 months and 12 months of treatment. It is not
201 possible to identify if the IMPA or prednisolone therapy was the cause of the joint laxity in these cases.
202 However, the cat which developed laxity 5 months into treatment had a chronic, one year history of
203 lameness without ligament laxity, prior to diagnosis, which suggests that prednisolone is more likely to have
204 been the cause of the ligament laxity in this cat. The other cat had a 3 week history of lameness prior to
205 diagnosis and developed ligament laxity after 12 months of prednisolone. The three cats which presented
206 with joint laxity had an acute onset of signs and none had received corticosteroids prior to presentation. No
207 cats underwent surgical arthrodesis. A differential diagnosis for a plantigrade stance in a cat is tibial nerve
208 neuropathy and diabetic neuropathy. None of the three cats were diabetic. Tibial nerve neuropathy was
209 excluded in one cat with evidence of subluxation of the calcaneoquartal joint on CT, the second cat had an
210 unremarkable examination performed by a neurologist and the final cat had excessive range of motion
211 detected on orthopedic examination.

212 *Infectious disease testing*

213 Infectious disease screening was variable between cases and was dependent on the clinician's decision.
214 Fourteen of the twenty cats (70%) either tested negative for *Mycoplasma species* on PCR of joint fluid (10)
215 or received a two week course of oral doxycycline without being tested (4). FIV and FeLV testing was
216 performed in fourteen (70%) of the cats; one cat in this case series was positive for FIV infection. The

217 diagnosis of FIV was confirmed with proviral DNA PCR testing in addition to a positive SNAP FIV antibody
218 test.

219 *Treatment and outcome*

220 A summary of the treatment and outcomes in the non-associative IMPA cases is found in Table 4 and
221 associative IMPA cases in Table 5. Specific treatments that were administered for the comorbidities
222 identified in cases of associative IMPA are included in Table 5.

223 All cats with presumed non-associative IMPA were treated with immunosuppressants. Eight of the 13 cats
224 were started on prednisolone monotherapy. Only three cats (38%) had a good response to treatment, of the
225 other five cats, four were administered second line immunosuppressants (chlorambucil [3] and ciclosporin
226 [1]), in addition to the prednisolone and one was euthanased. Two cats responded well to the addition of
227 chlorambucil and one failed to respond to both chlorambucil and also to an alternative second agent,
228 mycophenolate. The cat started on ciclosporin as a second agent failed to improve but responded well to an
229 alternative second agent, chlorambucil, alongside prednisolone. Five cats were started on treatment at
230 diagnosis with prednisolone and ciclosporin as a second agent. Two of these cats (40%) responded well, the
231 remaining three cats were started on an alternative second agent, chlorambucil alongside prednisolone. Of
232 the three cats started on chlorambucil, one responded well, one was euthanised when failing to respond
233 and one developed adverse effects with chlorambucil but responded well to an alternative
234 immunosuppressive agent, leflunomide together with the prednisolone.

235 Five of the seven cats with associative IMPA were treated with immunosuppressants. Two cats received
236 prednisolone monotherapy and showed a poor response to treatment, one cat failed to respond to the

237 addition of chlorambucil despite successful treatment of the concurrent pneumonia with antibiotic therapy,
238 and was euthanised. The second cat suffered adverse effects and was not started on alternative treatment
239 due to poor compliance; this cat had evidence of ongoing IMPA despite resolution of the clinical signs of the
240 concurrent chronic enteropathy. One cat was started on prednisolone monotherapy at an anti-inflammatory
241 dose. This patient did not respond to treatment and was euthanised within two days; this cat tested positive
242 for FIV. One cat received ciclosporin monotherapy, developed adverse effects within 5 days of treatment
243 and responded well to chlorambucil monotherapy alongside resolution of the clinical signs of the concurrent
244 chronic enteropathy. One cat was treated with prednisolone and chlorambucil alongside resolution of the
245 clinical signs of the concurrent enteropathy and responded well to treatment.

246 Overall, a good outcome was achieved in 14/20 (70%) cats with IMPA: 10/13 cats (77%) with non-associative
247 IMPA and 4/7 cats (57%) with associative IMPA. Two of the fourteen cats with a good outcome were not
248 treated with immunosuppressive agents and received meloxicam alone, although both cats displayed
249 recurrence of clinical signs if the meloxicam was withdrawn. In one cat immunosuppression was avoided
250 due to chronic feline herpesvirus and concerns for recrudescence. The second cat was not given
251 immunosuppressants due to concerns this would exacerbate the suspected concurrent discospondylitis. The
252 remaining 12 cats with a good outcome received immunosuppressants. Seven cats (35%) were tapered off
253 all medications and had no reported relapses during a follow up period between four months and five years
254 (median follow up two years). One cat was tapered off all medications and had a single relapse five months
255 after, which was treated successfully with prednisolone monotherapy. Two cats were responding well to
256 treatment and were being successfully tapered off medication at the time of writing, six months and 10

257 months post diagnosis. Three cats were euthanised for reasons unrelated to IMPA and at the time of
258 euthanasia all cats had no clinical signs of IMPA.

259 **Discussion**

260 This is the largest case series to date, describing the clinical features and outcome of cats with non-erosive
261 IMPA. In this study non-associative IMPA was found to be more common than associative IMPA. In the
262 previous case series 7/13 cats [54%] had associative IMPA with comorbidities identified at the time of IMPA
263 diagnosis.⁴ Overall, the prognosis for cats with non-erosive IMPA appeared to be favourable, with most
264 responding well to treatment and 70% having a good outcome.

265 Only 5/13 (38%) cats with non-associative IMPA and 1/5 (20%) cats with associative IMPA, treated with
266 immunosuppressants responded well to first line treatment. The remaining 12 cats received additional
267 immunosuppressive agents or were euthanised. The need for multimodal immunosuppression did not
268 always result in a poor outcome and overall, of the nine cats who received additional agents, six cats (67%)
269 had a good outcome.

270 In the previous case series by Bennett and Nash⁴ all thirteen cats with IMPA were treated with prednisolone
271 monotherapy (1mg/kg q12hours, tapered over six weeks when possible). In this case series 6/13 cats (46%)
272 showed a good response to treatment and were successfully tapered off prednisolone. The remaining seven
273 cats were euthanised as a result of the IMPA, due to either a poor response to treatment (3) or an inability
274 to taper the dose of prednisolone (4). The results of our study could suggest that cats with IMPA may not
275 respond well to first line treatment, but this does not necessarily predict the final outcome and second line
276 treatment should be considered.

277 The most commonly identified comorbidity in our case series was gastrointestinal disease. None of the cats
278 in this case series were diagnosed with neoplasia. This contrasts with the case series in which Bennett and
279 Nash⁴ reported 4/13 cats with IMPA had associated neoplasia. However, one cat in the current study was
280 euthanised within four months of diagnosis as a result of suspected lymphoma, based on the development
281 of a marked circulating lymphocytosis. Unfortunately, flow cytometry and histopathology were not
282 performed, and this was not confirmed.

283 FIV was identified as a comorbidity in one cat in this population. Immune mediated disease in cats with FIV
284 most commonly occurs secondary to excessive antibody production in response to chronic infection. This
285 results in hypergammaglobulinemia and immune complex deposition.⁷⁻⁹ This cat did not receive
286 immunosuppressive therapy; the poor outcome may reflect the presence of the FIV infection, or the
287 absence of immunosuppressive therapy. To the authors' knowledge FIV has not been previously reported in
288 association with non-erosive, IMPA. FIV has been associated with feline chronic progressive polyarthritis,
289 which is an erosive form of IMPA.¹⁰ Six cats in this study were not tested for FIV and considering it is often
290 an asymptomatic infection, it is possible the true prevalence in this population was underestimated. In the
291 previous case series Bennett and Nash⁴ reported three cats with IMPA were FeLV positive. However, the
292 FeLV vaccination only became widely available after the previous case series was reported and this may
293 have impacted the prevalence of FeLV in the population at that time.

294 In the present study 10/18 (55%) cats, in which rectal temperature was recorded, were pyrexia on
295 presentation. This is similar to the previous case series where 8/13 cats (61%) were found to be pyrexia.⁴ As
296 a result of this, the absence of pyrexia cannot be used to exclude IMPA in a cat presenting with lameness.

297 This also correlates with the current literature on canine IMPA. Pyrexia is reported in approximately 50% of
298 dogs with IMPA which is a common cause of “pyrexia of unknown origin” (PUO).^{1,11,12} In a study of 101 dogs
299 with PUO, IMPA was found to be the diagnosis in 20% and some of these dogs presented without any
300 obvious evidence of joint pain or inflammation on physical examination.¹³ In a recent study of 106 cats with
301 PUO, IMPA was the diagnosis in only 3% of these cases.⁶ The difference in the prevalence is likely to be
302 reflective of the variability in the incidence of canine and feline IMPA and the higher prevalence of infectious
303 diseases in cats.¹⁴

304 Septic bacterial arthritis occurs most commonly as a result of cat bites and usually affects one joint in a
305 single limb.¹⁴ Interestingly, seven cats in this population presented to their primary care practice with single
306 limb lameness despite an increased proportion of non-degenerate neutrophils being identified within
307 synovial fluid samples taken from multiple joints. IMPA in cats should therefore not be excluded based on
308 involvement of a single limb alone and further investigations should be performed if there is no evidence of
309 a wound or bacterial infection. Another interesting observation is the high yield of an increased proportion
310 of non-degenerate neutrophils within the tarsal joints of cats in this study; this has also been found to be the
311 most commonly affected joint in dogs.¹²

312 Two cats in this study were treated with NSAIDs and clinical signs of IMPA such as lameness and reluctance
313 to walk, resolved whilst on treatment in both cats. Attempts to withdraw NSAIDs were unsuccessful due to
314 recurrence of clinical signs. These findings could reflect that in these cases there was an ongoing
315 inflammatory arthropathy and the NSAIDs were treating the joint pain successfully, rather than the
316 underlying pathology. Unfortunately, neither cat had repeat joint aspirates performed so it is not possible to

317 confirm this. One of these cats had concurrent discospondylitis with IMPA; it is therefore possible that the
318 relapse in clinical signs of IMPA seen in this cat was as a result of failure to treat this comorbidity. However,
319 this was considered less likely as the clinical signs of the discospondylitis, completely resolved with antibiotic
320 treatment. A positive response to treatment with NSAIDs in cats presenting with lameness of unknown
321 origin should also not be used to exclude IMPA and further investigations should be considered if relapse of
322 clinical signs is seen when NSAIDs are withdrawn.

323 *Mycoplasma* species, have been implicated in rare cases of feline polyarthritis.¹⁴ *Mycoplasma gateae* has
324 been identified in the synovial fluid of a small number of cats with naturally occurring erosive arthritis and
325 has also been experimentally reproduced with intravenous inoculation into healthy cats.^{15,16} *Mycoplasma*
326 *felis* has been documented to cause non-erosive monoarthritis in two immunocompetent cats and has been
327 cultured from a cat with polyarthritis and suspected severe immunocompromise.^{17,18} Infectious disease
328 screening for *Mycoplasma* species was variable in this population. It is not possible to definitively conclude
329 that the six cats which did not receive testing or treatment for *Mycoplasma* were not infected. However, 5/6
330 cats had a good response to immunosuppressive treatment, making *Mycoplasma* infection unlikely in any of
331 the cases.

332 Five cats in this study presented with, or developed, joint ligament laxity whilst on prednisolone therapy.
333 Ligament laxity has been identified in multiple cases of feline erosive polyarthritis such as feline rheumatoid
334 arthritis and feline periosteal proliferative polyarthritis.^{2,10,19} To the authors knowledge ligament laxity has
335 not been previously documented in cases of feline non-erosive IMPA but has been reported in five dogs with
336 non-erosive IMPA.²⁰ The prognosis for dogs with ligament laxity and non-erosive IMPA is poor²⁰. The

337 outcome in the five cats in this study with ligament laxity was variable. One cat was euthanised due to the
338 severity of clinical signs within two days of diagnosis. Two cats required multimodal immunosuppression due
339 to frequent relapses and poor response to treatment. Unfortunately, it is not possible to comment on the
340 ability to withdraw medication in either case, as both were still receiving treatment at the time of writing.
341 One of the cats with ligament laxity had associative IMPA and responded well to meloxicam. The final cat
342 responded poorly to prednisolone monotherapy and had severe adverse effects; further
343 immunosuppression and NSAIDs were recommended but not provided. This cat had ongoing chronic
344 lameness on follow up but received no further treatment.

345 There are several limitations of the current study. As this study was retrospective and spanned several
346 referral centers the investigations and treatment provided were not standardised. It is therefore difficult to
347 directly compare the efficacy of specific treatments and treatment recommendations cannot be established.
348 Future prospective studies would be required to investigate treatment protocols. In addition, cases were
349 recruited from a referral population only. It is considered possible that some cases of IMPA identified in
350 primary care practices might have responded more favourably to prednisolone monotherapy and would not
351 have been presented to a referral centre, thus leading to a selection bias in the cases included in this study.
352 However, the number of cases in this cohort which had received prior immunosuppressive therapy was low
353 (3 cats [15%]). Most cats presented to the referral centres soon after the onset of clinical signs making the
354 findings of this study relevant to both referral and primary care practices. Some cats showed significant
355 adverse effects with ciclosporin therapy and were changed to an alternative second agent. Blood ciclosporin
356 levels were not monitored in these cats so it is not possible to confirm if high levels were present in these
357 cases.

358 The definition of IMPA is poorly defined in the veterinary literature with no consensus on the diagnostic
359 criteria. A cut-off of two affected joints to make a diagnosis of IMPA has commonly been used and accepted
360 as an inclusion criteria in dogs and was extrapolated to this study.^{11,21,22} In the human literature the
361 involvement of five joints is required to make a diagnosis of polyarthritis.²³ It is therefore possible that the
362 cases in this study with less than five affected joints (9 cats) were misdiagnosed as IMPA. In addition, due to
363 the small volume inherently available from joint aspirates in cats, total nucleated cell counts were not
364 available for all cases.

365 Cases were divided into associative and non-associative based on the diagnosis made by the primary
366 clinician. In associative IMPA the comorbidities identified were not proven to be responsible for the non-
367 erosive IMPA. In addition to this, some cases of associative IMPA did not resolve despite resolution of the
368 comorbidities; this could suggest that the disease identified was incidental and not related to the IMPA. In
369 two cases, thoracic imaging was not performed so it is not possible to exclude a comorbidity present in the
370 thorax for these cases. One case without thoracic imaging was considered to have non-associative IMPA and
371 it is therefore possible this case was categorised incorrectly as a result of this. Infectious disease screening
372 was highly variable and so the prevalence of diseases such as FIV, cannot be established.

373 A further limitation in this study is the lack of follow up arthrocentesis samples to definitively prove
374 remission and cases of relapse. In this study the presence of relapse was based on the recurrence of clinical
375 signs. Despite these limitations this study provides useful information on the presenting signs, treatment
376 options and outcomes in cases of feline IMPA.

377 **Conclusions**

378 The prognosis for feline IMPA can be good. However, multimodal immunosuppression is often required in
379 order to control clinical signs associated with disease. Sampling of the tarsal joints most frequently identified
380 changes consistent with IMPA in this cohort. Ligament laxity can occur in non-erosive IMPA but does not
381 appear to significantly impact overall prognosis. IMPA should be considered in lame cats, with or without
382 pyrexia when there is no clear evidence of trauma or infection.

383 **Author Note**

384 This paper was presented in part as an oral presentation at the 2019 BSAVA conference.

385 **Conflict of Interest**

386 The authors declared no potential conflicts of interest with respect to the research, authorship, and/or
387 publication of this article.

388 **Funding**

389 The authors received no financial support for the research, authorship, and/or publication of this article.

390 **Ethical Approval**

391 The work described in this manuscript involved the use of non-experimental (owned or unowned) animals.
392 Established internationally recognised high standards ('best practice') of veterinary clinical care for the
393 individual patient were always followed. Ethical approval from a committee was therefore not specifically
394 required for publication in JFMS. Although not required, where ethical approval was still obtained, it is
395 stated in the manuscript.

396 **Informed Consent statement**

397 Informed consent (verbal or written) was obtained from the owner or legal custodian of all animal(s)
398 described in this work (experimental or non-experimental animals, including cadavers) for all procedure(s)
399 undertaken (prospective or retrospective studies).

400 For any animals or people individually identifiable within this publication, informed consent (verbal or
401 written) for their use in the publication was obtained from the people involved.

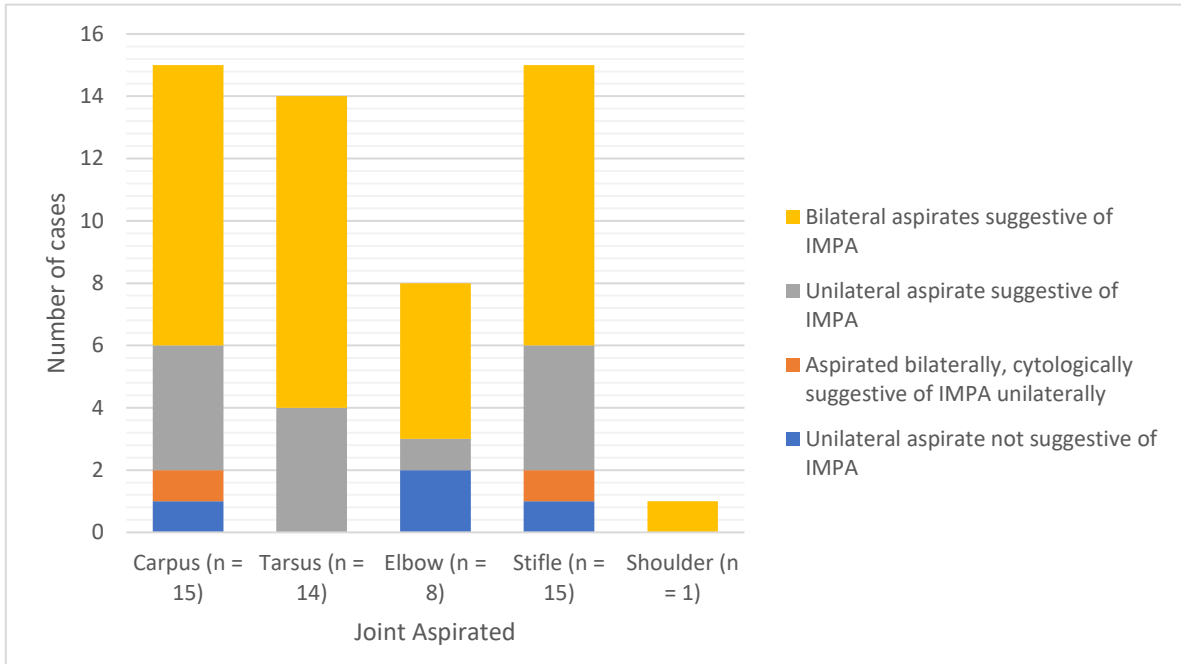
402 **References**

- 403 1 Stone M. Immune-Mediated Polyarthritides and Other Polyarthritides. In: Ettinger SJ, Feldman EC,
404 Cote E. (eds) Textbook of Veterinary Internal Medicine. 8th ed. Elsevier health sciences: 2017; Pp
405 861 – 866.
- 406 2 MacWilliams PS, Friedrichs KR. Laboratory evaluation and interpretation of synovial fluid.
407 Veterinary Clinics: Small Animal Practice. 2003; 33(1):153-78.
- 408 3 Garden OA et al. ACVIM consensus statement on the diagnosis of immune-mediated hemolytic
409 anemia in dogs and cats. Journal of veterinary internal medicine. 2019; 33(2):313-34.
410
- 411 4 Bennett D, Nash AS. Feline immune-based polyarthritides: a study of thirty-one cases. Journal of Small
412 Animal Practice. 1988; 29(8):501-23.
- 413 5 Pedersen NC. A review of feline infectious peritonitis virus infection [review]: 1963–2008. Journal of
414 feline medicine and surgery. 2009; 11(4):225-58.
- 415 6 Spencer SE, Knowles T, Ramsey IK, et al. Pyrexia in cats: retrospective analysis of signalment, clinical
416 investigations, diagnosis and influence of prior treatment in 106 referred cases. Journal of feline
417 medicine and surgery. 2017; 19(11):1123-30.

- 418 7 Ackley CD, Yamamoto JK, Levy N, et al. Immunologic abnormalities in pathogen-free cats
419 experimentally infected with feline immunodeficiency virus. *Journal of Virology* 1990; 64(11):
420 pp.5652-5655.
- 421 8 Gleich S, and Hartmann K. Hematology and serum biochemistry of feline immunodeficiency virus-
422 infected and feline leukemia virus-infected cats. *Journal of Veterinary Internal Medicine* 2009;
423 23(3): pp.552-558.
- 424 9 Hartmann K. Clinical aspects of feline retroviruses: a review. *Viruses* 2012; 4(11): pp.2684-2710.
- 425 10 Inkpen H. Chronic progressive polyarthritis in a domestic shorthair cat. *The Canadian Veterinary*
426 *Journal* 2015; 56(6): p.621
- 427 11 Clements DN, Gear RN, Tattersall J, et al. Type I immune-mediated polyarthritis in dogs: 39 cases
428 (1997–2002). *Journal of the American Veterinary Medical Association*. 2004; 224(8):1323-7.
- 429 12 Stull JW, Evason M, Carr AP, et al. Canine immune-mediated polyarthritis: clinical and laboratory
430 findings in 83 cases in western Canada (1991–2001). *The Canadian Veterinary Journal*. 2008
431 49(12):1195.
- 432 13 Dunn KJ, Dunn J. Diagnostic investigations in 101 dogs with pyrexia of unknown origin. *Journal of*
433 *small animal practice*. 1998; 39(12):574-80.
- 434 14 Lemetayer J, and Taylor S. Inflammatory joint disease in cats: Diagnostic approach and treatment.
435 *Journal of feline medicine and surgery* 2014; 16(7): pp.547-562.
- 436 15 Moise NS, Crissman JW, Fairbrother JF, et al. *Mycoplasma gateae* arthritis and tenosynovitis in cats:
437 case report and experimental reproduction of the disease. *American journal of veterinary research*
438 1983; 44(1): pp.16-21.

- 439 16 Zeugswetter F, Hittmair KM, de Arespacochaga AG, et al Erosive polyarthritis associated with
440 *Mycoplasma gateae* in a cat. *Journal of feline medicine and surgery* 2007; 9(3): pp.226-231.
- 441 17 Liehmann L, Degasperi B, Spergser J, et al. *Mycoplasma felis* arthritis in two cats. *Journal of small*
442 *animal practice* 2006; 47(8): pp.476-479.
- 443 18 Hooper PT, Ireland LA and Carter A. *Mycoplasma* polyarthritis in a cat with probable severe immune
444 deficiency. *Aust Vet J* 1985; 62: 352.
- 445 19 Pedersen NC, Pool RR, O'Brien T. Feline chronic progressive polyarthritis. *American Journal of*
446 *Veterinary Research* 1980; 41(4):522-35.
- 447 20 Whitworth F, Adamantos S, Frowde P, et al. Ligament Laxity in Nonerosive Immune-Mediated
448 Polyarthritis in Dogs: Five Cases (2009–2017). *Journal of the American Animal Hospital Association*
449 2019; 55(4):210-4.
- 450 21 Stull JW et al. Canine immune-mediated polyarthritis: clinical and laboratory findings in 83 cases in
451 western Canada (1991–2001). *The Canadian Veterinary Journal*. 2008; 49(12):1195.
- 452 22 Colopy SA et al. Efficacy of leflunomide for treatment of immune-mediated polyarthritis in dogs: 14
453 cases (2006–2008). *Journal of the American Veterinary Medical Association* 2010; 236(3):312-8
- 454 23 Alpay-Kanitez N, Çelik S, Bes C. Polyarthritis and its differential diagnosis. *European Journal of*
455 *Rheumatology* 2019; 6(4):167.

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Figure 1: Proportion of joint aspirates yielding synovial fluid samples cytologically suggestive (neutrophils > 10% of the nucleated cell count and no intra or extracellular bacteria) of immune mediated polyarthritis (IMPA) in 20 cats subsequently diagnosed with IMPA based on the involvement of ≥ 2 joints.