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Citation for published version:

Skelly, M & Spoel, SH 2021, 'Characterising plant deubiquitinases with in vitro activity-based labelling and ubiquitin chain disassembly assays', *Bio-protocol*, vol. 11, no. 9, e4015.
<https://doi.org/10.21769/BioProtoc.4015>

Digital Object Identifier (DOI):

[10.21769/BioProtoc.4015](https://doi.org/10.21769/BioProtoc.4015)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Bio-protocol

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Characterising Plant Deubiquitinases with *in vitro* Activity-based Labelling and Ubiquitin Chain Disassembly Assays

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[Abstract] Post-translational modification of proteins by ubiquitin is an essential cellular signaling mechanism in all eukaryotes. Ubiquitin is removed from target proteins by a wide range of deubiquitinase (DUB) enzymes with different activities and substrate specificities. Understanding how DUBs function *in vitro* is a vital first step to uncovering their cellular roles. Here, we provide protocols for the rapid analysis of DUB activity *in vitro* by activity-based labelling with the suicide probe, HA-ubiquitin vinyl sulfone (HA-UbVS), and ubiquitin chain disassembly assays. We have previously used these methods to analyse the activity of the *Arabidopsis thaliana* DUB, UBP6, but in principle, these protocols are applicable to any DUB of interest.

Keywords: Ubiquitin, Deubiquitinase, Ubiquitin vinyl sulfone, Proteasome, Cell signaling, Proteostasis

[Background] Regulation of cellular protein homeostasis (proteostasis), the intricate balance between protein synthesis, modification, and degradation, is essential for the survival of all organisms (Balch *et al.*, 2008). Proteostasis is maintained by a diverse range of post-translational modifications (PTMs) that increase the size and functionality of proteomes (Skelly *et al.*, 2016). Ubiquitination, the covalent attachment of the small regulatory protein ubiquitin to target proteins, is a prevalent PTM involved in numerous cellular processes in eukaryotes. Ubiquitin is conjugated predominantly to lysine residues of target substrates via its C-terminus, which requires the sequential actions of three types of enzyme; E1 ubiquitin-activating enzymes, E2 ubiquitin-conjugating enzymes, and E3 ubiquitin ligases. Ubiquitin itself contains seven lysine residues that can also be modified, facilitating formation of polyubiquitin chains of various linkage types. Different ubiquitin chain topologies have specific signalling roles and are associated with different cellular processes. For example, K48-linked chains generally target proteins for degradation by the proteasome, a large multi-protein complex that is responsible for the majority of proteolytic activity in eukaryotic cells. However, ubiquitin modifications can also be reversed by the action of a wide range of deubiquitinase (DUB) enzymes with diverse specificities for different ubiquitin linkage types (Mevisen and Komander, 2017). Since deubiquitination determines the stability, activity, or function of proteins, entire proteomes can be shaped by the activities of specific DUBs. Many human diseases, including cancers, are associated with dysregulated DUB activity, demonstrating the physiological and pathological importance of these enzymes (Harrigan *et al.*, 2018; Clague *et al.*, 2019).

To understand DUB function, their activity can be analysed by various *in vitro* methods to determine substrate specificity and regulation of activity (Cho *et al.*, 2020). These methods include activity-based

protein profiling using synthetic ubiquitin suicide probes that irreversibly label DUB active sites (Borodovsky *et al.*, 2002; Gong *et al.*, 2018; Schauer *et al.*, 2020), as well as assays in which purified ubiquitin chains are incubated with DUBs and chain disassembly is monitored by western blotting or gel staining. Recently, we used these methods to uncover the signalling roles of the *Arabidopsis thaliana* proteasome-associated deubiquitinases, UBP6 and UBP7, in plant immunity (Skelly *et al.*, 2019). Here, we describe protocols for analysing recombinant UBP6 activity by HA-UbVS labelling and *in vitro* deubiquitination assays. Importantly, these methods can be applied to any DUB of interest and in the case of HA-UbVS, labelling can also be applied to protein extracts from any organism to uncover proteome-wide DUB activity. Characterising the activity and regulation of DUBs is vital to understand how these enzymes act *in vivo* and will advance the fields of ubiquitin signalling and proteostasis across eukaryotes.

Materials and Reagents

1. HA-ubiquitin-vinyl sulfone (Boston Biochem, catalog number: U-212)
2. 26S proteasome (Ub-VS treated) (Ubiquigent, catalog number: 65-1020-010)
3. Recombinant purified His₆-T7-UBP6 (Skelly *et al.*, 2019) or other DUB of interest
4. Ubiquitin substrates for deubiquitination assays:
 - Poly-ubiquitin (Ub₃-7) K48-linked (Boston Biochem, catalog number: UC-220)
 - Poly-ubiquitin (Ub₃-7) K63-linked (Boston Biochem, catalog number: UC-320)
 - Di-ubiquitin K48-linked (Boston Biochem, catalog number: UC-200B)
 - Di-ubiquitin K63-linked (Boston Biochem, catalog number: UC-300B)
5. UltraPure Tris (Invitrogen, catalog number: 15504-020)
6. MgCl₂ hexahydrate (Sigma-Aldrich, catalog number: M2670)
7. Adenosine triphosphate (ATP) (Sigma-Aldrich, catalog number: A26209)
8. Dithiothreitol (DTT) (Sigma-Aldrich, catalog number: 1114740005)
9. Glycerol (Sigma-Aldrich, catalog number: G5516)
10. Sodium dodecyl sulfate (SDS) (Sigma-Aldrich, catalog number: L3771)
11. Bromophenol blue (Sigma-Aldrich, catalog number: B8026)
12. Anti-HA antibody (ThermoFisher, catalog number: 26183)
13. Anti-ubiquitin antibody (Santa Cruz Biotechnology, catalog number: sc-8017)
14. Antibody against the DUB being analysed, e.g., anti-T7 for His₆-T7-UBP6 (Millipore, catalog number: 69522)
15. Double-distilled H₂O (ddH₂O)
16. 15% SDS-PAGE gel (e.g., compatible with Bio-Rad Mini-PROTEAN system)
17. SuperSignal West Pico PLUS Chemiluminescent Substrate (ThermoFisher, catalog number: 34577)
18. 10× DUB labelling buffer (store at -20°C) (see Recipes)
19. 10× deubiquitination buffer (store at -20°C) (see Recipes)

20. 2× SDS-PAGE loading buffer (see Recipes)
21. DUBs, proteasomes, HA-UbVS, and ubiquitin substrates (see Recipes)

Equipment

1. 1.5 ml microcentrifuge tubes (e.g., Starlab, catalog number: S1615-5500)
2. Pipettes and tips (e.g., Gilson Pipettman, various volumes)
3. Heat block (e.g., ThermoFisher, catalog number: 88870004)
4. Protein gel electrophoresis and western blotting apparatus (e.g., Bio-Rad Mini-PROTEAN system)

Procedure

A. HA-UbVS labelling

1. Set up 10- μ l reactions including the following: 1 μ l 10× DUB labelling buffer, 5 μ l 700 nM DUB, 1.25 μ l 80 nM 26S proteasome (if required), 1 μ l 7 μ M HA-UbVS, and 1.75 μ l ddH₂O. Control samples should be included in which components are omitted and replaced with ddH₂O.
2. Incubate for 30 min at room temperature or in a heat block at 22°C.
3. Add 10 μ l 2× SDS-PAGE loading buffer to each sample.
4. Incubate for 10 min in a heat block at 70°C .
5. Perform SDS-PAGE with a 15% gel and western blotting using standard methods with:
 - a. Anti-HA antibodies (at 1:5000 dilution) to detect free HA-UbVS (~9.8 kDa) and HA-UbVS-labelled DUB (Figure 1).
 - b. Antibodies (at antibody-specific dilutions) against the appropriate DUB or epitope tag to detect both unlabelled and labelled DUB. Any standard imaging system can be used depending on the secondary antibodies (e.g., film or CCD imaging system for chemiluminescence).

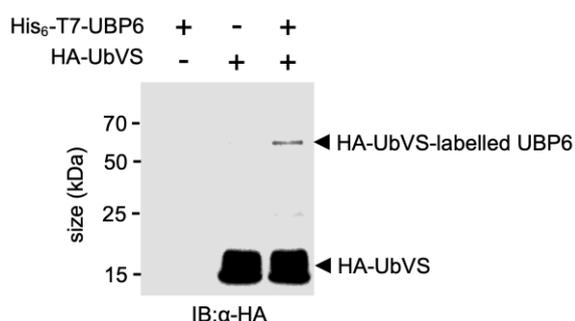


Figure 1. HA-UbVS labelling of recombinant purified His₆-T7-UBP6. A typical western blot showing free and HA-UbVS-labelled recombinant His₆-T7-UBP6. Adapted from Figure 6B of Skelly *et al.* (2019).

B. *In vitro* deubiquitination assays

1. Set up 10 μ l reactions including the following: 1 μ l 10 \times deubiquitination buffer, 1 μ l 200 nM DUB, 1 μ l 12.5 nM 26S proteasome (if required), 1 μ l 4 μ M ubiquitin substrate, and 6 μ l ddH₂O. Control samples should be included in which components are omitted and replaced with ddH₂O.
2. Incubate in a heat block at 30°C for the desired time [2-16 h for His₆-T7-UBP6 (Skelly *et al.*, 2019), but dependent on the DUB and substrate].
3. Add 10 μ l 2 \times SDS-PAGE loading buffer to each sample.
4. Incubate for 10 min in a heat block at 70°C .
5. Perform SDS-PAGE with a 15% gel and western blotting using standard methods with anti-ubiquitin antibodies to detect the original ubiquitin chain substrate and observe any DUB-mediated chain disassembly as lower-order ubiquitin species. Any standard imaging system can be used depending on the secondary antibodies (*e.g.*, film or CCD imaging system for chemiluminescence).

Recipes

1. 10 \times DUB labelling buffer (store at -20°C)
500 mM Tris pH 7.4
50 mM MgCl₂
10 mM DTT
10 mM ATP
2. 10 \times deubiquitination buffer (store at -20°C)
500 mM Tris pH 7.4
50 mM MgCl₂
10 mM DTT
50 mM ATP
3. 2 \times SDS-PAGE loading buffer (store at room temperature, add DTT fresh before use)
20% glycerol
120 mM Tris pH 6.8
4% SDS
0.02% Bromophenol Blue
100 mM DTT
4. DUBs, proteasomes, HA-UbVS, and ubiquitin substrates
Diluted in ddH₂O from stocks to the required concentrations described above

Acknowledgments

This work was supported by a Royal Society University Research Fellowship (UF090321), a BBSRC grant (BB/L006219/1), and the European Research Council (ERC) under the European Union's

Horizon 2020 research and innovation programme (grant agreement No. 678511). This protocol was derived from Skelly *et al.* (2019).

Competing interests

The authors declare that no competing interests exist.

References

1. Balch, W. E., Morimoto, R. I., Dillin, A. and Kelly, J. W. (2008). [Adapting proteostasis for disease intervention](#). *Science* 319(5865): 916-919.
2. Borodovsky, A., Ovaa, H., Kolli, N., Gan-Erdene, T., Wilkinson, K. D., Ploegh, H. L. and Kessler, B. M. (2002). [Chemistry-based functional proteomics reveals novel members of the deubiquitinating enzyme family](#). *Chem Biol* 9(10): 1149-1159.
3. Cho, J., Park, J., Kim, E. E. and Song, E. J. (2020). [Assay Systems for Profiling Deubiquitinating Activity](#). *Int J Mol Sci* 21(16): 5638.
4. Clague, M. J., Urbe, S. and Komander, D. (2019). [Breaking the chains: deubiquitylating enzyme specificity begets function](#). *Nat Rev Mol Cell Biol* 20(6): 338-352.
5. Gong, P., Davidson, G. A., Gui, W., Yang, K., Bozza, W. P. and Zhuang, Z. (2018). [Activity-based ubiquitin-protein probes reveal target protein specificity of deubiquitinating enzymes](#). *Chem Sci* 9(40): 7859-7865.
6. Harrigan, J. A., Jacq, X., Martin, N. M. and Jackson, S. P. (2018). [Deubiquitylating enzymes and drug discovery: emerging opportunities](#). *Nat Rev Drug Discov* 17(1): 57-78.
7. Mevissen, T. E. T. and Komander, D. (2017). [Mechanisms of Deubiquitinase Specificity and Regulation](#). *Annu Rev Biochem* 86: 159-192.
8. Schauer, N. J., Magin, R. S., Liu, X., Doherty, L. M. and Buhrlage, S. J. (2020). [Advances in Discovering Deubiquitinating Enzyme \(DUB\) Inhibitors](#). *J Med Chem* 63(6): 2731-2750.
9. Skelly, M. J., Frungillo, L. and Spoel, S. H. (2016). [Transcriptional regulation by complex interplay between post-translational modifications](#). *Curr Opin Plant Biol* 33: 126-132.
10. Skelly, M. J., Furniss, J. J., Grey, H., Wong, K. W. and Spoel, S. H. (2019). [Dynamic ubiquitination determines transcriptional activity of the plant immune coactivator NPR1](#). *Elife* 8: e47005.