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Solar light and metal-doped TiO$_2$ to eliminate water-transmitted bacterial pathogens: Photocatalyst characterization and disinfection performance

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Abstract

The present study deals with the inactivation of *Escherichia coli* and *Klebsiella pneumoniae* in water by means of heterogeneous photocatalysis under simulated solar irradiation. For this purpose, novel Mn-, Co- and Mn/Co-doped TiO$_2$ catalysts were prepared. A straightforward, simple and inexpensive process has been developed based on a co-precipitation method for the synthesis of metal-doped catalysts, which were subsequently assessed in terms of their disinfection efficiency. The effect of various operating conditions, such as metal dopant (Mn-, Co- and Mn/Co), dopant concentration (0.02-1 wt%), catalyst concentration (25-250 mg/L), bacterial concentration ($10^2$-$10^8$ CFU/mL), treatment time (up to 60 min), toxic effects on bacteria and photon flux (4.93-5.8$\times$10$^{-7}$ einstein/(L.s)), was examined under simulated solar irradiation. Metal-doped TiO$_2$ samples were prepared reproducibly and doping shifted the optical absorption edge to the visible region. Their activity was superior to the respective of commercially available P25 titania. The reference strains of *E. coli* and *K. pneumoniae* proved to be readily inactivated during photocatalytic treatment of aqueous samples, since disinfection occurred rapidly (i.e. after only 10 min of irradiation) with the dopant concentration affecting the overall process to a certain extent. Disinfection follows a pseudo-first order kinetic rate in terms of both bacteria removal. Inactivation of the bacteria is attributed to the oxidative degradation of their cells and increase of their cell permeability and not to the potential toxicity of the metal-doped semiconductors, which did not exhibit any bactericidal properties. It has been shown that the improved activity of the Mn-, Co-, and binary Mn/Co doped TiO$_2$ is accredited to the fact that they can be activated in the visible part of the spectrum, in the absence of UV light (i.e. >420 nm).

Keywords:

Water disinfection; metal doping; solar photocatalysis; *E. coli*; *K. pneumoniae*
1. Introduction

Occurrence of bacterial pathogens and faecal contamination in surface water may pose high health risks, as they are considered major agents of waterborne diseases. Given that potable water is an essential requirement, efficiency of disinfection techniques is imperative for the adequate inactivation of microorganisms and the protection of public health [1]. The most popular disinfection techniques nowadays involve chemical compounds, filtration or radiation (e.g. chlorination, ozonation, UV irradiation etc.), which may act by different means like inhibition of enzymatic activity or destruction of cellular components [2]. However, considerable disadvantages including toxic by-products generated during chlorination, high cost of ozonation and action limitation depending on source-water turbidity when UV irradiation is applied, have led to the development of alternative methods [3, 4].

Semiconductor photocatalysis has emerged as a promising technique for microbial inactivation in various aqueous matrices, including diverse types of bacteria, fungi, viruses, and spores [5-7]. Titanium dioxide (TiO\textsubscript{2}) is widely used as a photocatalyst in these processes due to its high efficiency, low toxicity, physico-chemical stability and low cost [2, 6, 8, 9]. A drawback, regarding most commercially available TiO\textsubscript{2} catalysts, is that they are mainly active under UV spectral range because of the high required band gap energy (~3.2 eV) for excitation of the semiconductor. Therefore, the bactericidal potential of TiO\textsubscript{2} photocatalysis has been extensively studied with the use of UV light, which is a small fraction of the total solar-light spectrum, excluding solar source of energy, which is abundant and free of cost [5, 10-12].

For this purpose, over the last decade, research interest has been focused on the use of solar irradiation for photocatalysis and thus the exploitation of the visible light energy. The photocatalytic efficiency of TiO\textsubscript{2} has been improved by many different strategies, which have
been adopted for either morphological or chemical modifications of the catalyst [2, 8, 13]. The latter involve incorporation of additional components in the TiO$_2$ structure, like non-metal or/and noble and transition metal deposition. Doping the TiO$_2$ by several metals such as copper, cobalt, manganese, etc. broadens the absorption spectrum of these semiconductors towards the visible light region, as new energy levels are formed between the valence and conduction band [1, 12-19]. The nature and the amount of the doping agent usually play an important role concerning the photocatalytic activity. On the other hand, some possible limitations have been reported like photo-induced corrosion and promoted charge recombination at some metal sites [2, 8, 20].

Up until now, various studies have been conducted in terms of the evaluation of disinfection efficiency of doped catalysts during water treatment under visible light, using mostly *Escherichia coli* as a model microorganism [3-5, 10, 14, 16, 18, 21-24]. In most cases, inactivation of bacteria has been attributed to the decomposition of bacterial outer membrane due to phospholipid peroxidation of the membrane, caused primarily by hydroxyl radicals, generated during treatment [25-27]. Apart from *E. coli*, the information regarding the behaviour of other bacteria is very limited. The bacterial content of water consists of many groups and species, which exhibit variable tolerance in disinfection as a result of differences in cellular structure. *Klebsiella pneumoniae* is considered as an emerging human pathogen and can be transmitted through water consumption, but it has been merely studied as far as its resistance against disinfection is concerned [28, 29].

In this perspective, the objectives of the present work were (i) to prepare novel cobalt- and manganese-doped titania materials and investigate their structural properties, and (ii) to study their potential to purify aqueous samples in terms of *E. coli* and *K. pneumoniae* reference strains removal under solar radiation. For this purpose, several operating parameters were investigated, namely catalyst type and loading, dopant concentration, initial bacterial
concentration, as well as photon flux, which typically influence disinfection effectiveness. Furthermore, scanning electron microscopy (SEM) was employed to detect destruction of cellular structure induced by photocatalysis.

2. Experimental

2.1 Materials

Titanium (IV) oxysulfate hydrate (TiOSO₄·xH₂O), manganese (II) acetate tetrahydrate (Mn(CH₃COO)_2), cobalt (II) acetate tetrahydrate (Co(CH₃COO)_2) and ammonium hydroxide (25% NH₄OH) purchased from Aldrich were applied. Commercially available titanium dioxide (TiO₂ P25) was purchased from Degussa - Evonik Corp. (physicochemical characteristics are anatase:rutile 75:25, particle size of 21 nm and BET area of 50 m²/g) and was used as benchmark.

2.2 Preparation of metal-doped TiO₂

A co-precipitation method was used to prepare metal-doped TiO₂ nanoparticles with molar ratio in different concentrations in the range of 0.02 - 1 wt%. Doped titanium dioxide was precipitated at pH ~7 from aqueous solution of TiOSO₄ titanium (IV) oxysulfate hydrate and dopant (Mn or Co or Mn/Co) by the addition of ammonia. After aging the suspension overnight, the precipitate was filtered and dried under air at 373K. The residue was crushed to a fine powder and calcined in a furnace at 973 K for 3 h. More details can be found in previous work [13].

2.3 Catalyst characterization techniques
Powder X-ray diffraction patterns were collected on a Rigaku D/MAX-2000H rotating anode diffractometer (CuKα radiation) equipped with the secondary pyrolytic graphite monochromator operated at 40 kV and 80 mA over the 2θ collection range of 10–80°. The scan rate was 0.05° s⁻¹. The average particle size (D in nm) of nanoparticles was calculated from the line broadening of the X-ray diffraction peak according to the Scherrer formula, as follows:

\[ D = \frac{k\lambda}{\beta \cos \theta} \]  

where \( k \) is the shape factor (~0.9), \( \lambda \) is the wavelength of the X-ray radiation (1.54 Å for CuKα), \( \beta \) is the full width at half maximum (FWHM) of the diffraction peak measured at 2θ, and \( \theta \) is the Bragg angle.

The phase content of TiO₂ samples was calculated using the formula:

\[ \% f_A = \left[ \frac{1}{1 + 1.265 \times \frac{I_R}{I_A}} \right] \times 100 \]  

where \( f_A \) is the content of anatase, and \( I_A \) and \( I_R \) are the integrated intensities of the anatase (100) and rutile (110) peaks respectively.

The UV – Visible diffuse reflectance spectra of the final powders were measured on a Perkin Elmer LAMBDA 950 with BaSO₄, as reference standard. The diffuse reflectance spectra were plotted as the Kubelka – Munk function, F(R), versus wavelength based on the Kubelka – Munk equation:

\[ F(R) = \frac{(1-R)^2}{(2R)} \]  

where the reflectance \( R = \frac{R_{\text{sample}}}{R_{\text{reference}}} \). The band gaps were then determined from the Kubelka – Munk function and the Tauc plots.

Surface morphology and elemental analysis of the samples were carried out using scanning electron microscopy (SEM) and an energy dispersive spectrometer (EDS) on a JSM-6390LV instrument. The microscopic nanostructures were studied by transmission electron microscopy (TEM) working at 200kV (JEM-2100 instrument equipped with LaB6 filament).
2.4 Disinfection experiments

The bacterial strains used in the present study were *E. coli* ATCC 23716 (American Type Culture Collection, Rockville, Md. USA) and *K. pneumoniae* NCTC 5056 (Public Health England Culture Collections). Both reference strains were inoculated separately in 10 mL of nutrient broth (HiMedia Laboratories) and grown overnight at 37°C. The concentration of bacterial cells in the suspension was estimated measuring its optical density at 600 nm (Shimadzu UV1240 spectrophotometer) where, according to McFarland scale, an absorbance of 0.132 corresponds approximately to a cell density of $1.5 \times 10^8$ CFU/mL. Plate counts were also performed for accurate bacterial count. In each case, suspensions were properly diluted to achieve the desired initial bacterial concentration, which was used for the subsequent experiments.

Photocatalytic experiments were conducted in batch type, laboratory scale photoreactor. Solar irradiation experiments were carried out in a solar radiation simulator system (Newport, model 96000) equipped with a 150 W xenon ozone-free lamp and an Air Mass 1.5 Global Filter (Newport, model 81094), simulating solar radiation reaching the surface of the earth at a zenith angle of 48.2°. According to the spectral irradiance data given by the manufacturer, simulated solar radiation contains about 5% UV-A radiation, and 0.1% UV-B radiation, while the filter cuts radiations with wavelengths lower than 280 nm. The incident radiation intensity on the photochemical reactor in the UV region of the electromagnetic spectrum was measured actinometrically using 2-nitrobenzaldehyde (Sigma-Aldrich) as the chemical actinometer [30, 31] and it was found to be $5.8 \times 10^{-7}$ einstein/(L.s), which corresponds to an irradiance of $1.31 \times 10^{-2}$ W/m². Additional runs were performed with (i) a 420 nm cutoff filter to remove all the UV light (FSQ-GG420, 50.8 mm x 50.8 mm), (ii) filter (FSQ-ND02, 50.8 mm×50.8 mm, 80% transmittance at 632.8 nm) to reduce irradiance to $5.3 \times 10^{-7}$ einstein/(L.s), and (iii) filter
(FSQ-ND04, 50.8 mm×50.8 mm, 40% transmittance at 632.8 nm) to reduce irradiance to 4.93 $10^7$ einstein/(L·s). Reactions took place in an open, double-walled, cylindrical glass vessel under continuous stirring.

In a typical run, the bacteria suspension was spiked in sterile water of 200 mL which were then loaded in the reaction vessel with the appropriate amount of catalyst. The solution was left in the dark under stirring for 20 min in order to equilibrate and then exposed to solar irradiation; this moment was taken as the starting point (time zero) of the disinfection experiment. Temperature was maintained at 25±2°C with a temperature control unit. The external reaction vessel was covered with aluminum foil to reflect irradiation exerting the outer wall of the reaction vessel. At specific time intervals samples of about 1.5 mL of the reaction solution were withdrawn and analyzed with respect to viable bacterial cells applying conventional culture method. All disinfection experiments were performed in triplicate.

Disinfection rate was measured in terms of *E. coli* and *K. pneumoniae* inactivation. The detection and enumeration of both bacteria in the solution were performed using the serial dilution streak plate procedure. The media used in the study were HiCrome Coliform Agar (HiMedia Laboratories) and M-FC agar M1124 (HiMedia Laboratories) for *E. coli* and *K. pneumonia*, respectively. Incubation was performed at 37°C for 20-24 h before viable counts were determined.

SEM observation of bacterial cells was carried out before and after treatment in each case, with the view to detect any destruction of cellular structure of reference strains induced by photocatalysis.

### 3. Results and discussion

#### 3.1. Structural and optical properties of metal-doped TiO$_2$ nanoparticles
The XRD patterns of Mn-, Co-doped TiO$_2$, and binary Mn/Co co-doped TiO$_2$ with dopant concentrations in the solution ranging from 0.02 to 1% for Mn- and Co-doped and 0.04 to 0.1% for the binary Mn/Co co-doped, respectfully, calcined at 700$^\circ$C for 3 h, are shown in Figure 1. Peak at 25.3$^\circ$ corresponds to the crystal plane 101 of the anatase phase. When the dopant concentration was in the range of 0.02 - 0.3 wt% the samples were monophasic with only the anatase polymorph TiO$_2$ being detected. Catalysts with 1 wt% of Mn- and Co-dopants, respectively exhibited a mixture of phases with both anatase and rutile. The peaks at 2$\theta$ values of 25.3, 37.6, 48.2, 53.9, 54.8, 62.7 and 75.2 corresponding to the (101), (004), (200), (105), (211), (204) and (215) planes, respectively are all anatase signature peaks. No obvious diffraction peaks attributed to MnO$_2$ or CoO$_2$ were observed at low dopant concentrations. Results of the particle size and phase content are shown in Table 1.

Figure 2 shows the UV-VIS absorption as a function of wavelength for metal-doped catalysts. Dopants at different concentrations caused considerable absorption shifts towards the visible range (400-800 nm) in comparison to the absorption threshold of P25 at 400 nm. Moreover, increasing the dopant concentration led to a significant decrease in the band gap energy below the value of 3.2 eV, which is required for excitation of the commercial P25 (Table 2). The band gap energy ranged between 2.4 and 2.87 eV and 2.4 and 2.97 eV for Mn and Co-doped catalysts, respectively. In the case of Co and Mn/Co-doped TiO$_2$, an extra sub-band gap energy was recorded in the range 1.4-1.6 eV, depending on the dopant concentration (Table 2). The new absorption shoulder of these catalysts at 400-800 nm, related to the presence of dopant (Figures 2b and 2c), may play key role in enhancing the overall photocatalytic activity within the visible range. The textural features of the catalysts were investigated with SEM (images are not shown for the sake of brevity); no specific morphology changes were detected, while the spherical shape particles of all the samples demonstrated some degree of agglomeration and the diameter ranged from 0.1 to 40 $\mu$m.
TEM images of the 1 wt% Mn- and 1 wt% Co-doped TiO$_2$ nanoparticles (Figure 3) confirmed the agglomeration of nanoparticles with size ranging between 35 and 45 nm, consistent with XRD measurements.

3.2 Photocatalytic disinfection

3.2.1 Effect of catalysts concentration

Given that the concentration of catalyst in slurry photocatalytic treatment strongly affects the overall process, preliminary runs were performed with both bacterial reference strains testing different commercial and metal-doped TiO$_2$ loadings. Tested concentrations were in the range 25-250 mg/L for *E. coli* and 100-250 mg/L for *K. pneumoniae*. Higher loadings were selected for *K. pneumoniae*, since it is a bacillus with a prominent capsule, which prevents cell destruction by bactericidal factors [28]. According to the results, increasing the catalyst loading improved inactivation rates for both microorganisms. In the case of *E. coli*, optimal inactivation rates (approximately 4-Log reduction within 5 min of treatment) were achieved when catalysts loading was 100 mg/L, beyond which disinfection reached a plateau.

Generally, increasing the photocatalyst concentration leads in lower penetration of light into the slurry, deteriorating disinfection efficiency [3]. Conversely, higher catalysts concentration was required for satisfactory decrease of *K. pneumoniae* population. In the presence of 250 mg/L of various types of TiO$_2$ that were used in the present study, total bacterial killing was recorded in almost 10 min with an initial bacterial density of $10^5$ CFU/mL. Therefore, all subsequent experiments were conducted using 100 mg/L and 250 mg/L of catalysts for *E. coli* and *K. pneumoniae* treatment, respectively. Optimal catalyst loadings may vary among studies as they depend on many parameters such as photocatalytic reactor geometry, light intensity, type of photocatalytic reactor etc. Apart from the trivial reference of *K. pneumoniae*
in photocatalytic studies, inactivation of *E. coli* is usually achieved in the range of loadings used in this work [2, 4, 22].

3.2.2. Effect of catalyst type

Bacterial inactivation was recorded only under solar irradiation, when destruction of the cells occurred. Efficiency of the metal-doped TiO$_2$ catalysts was assessed during a series of photocatalytic experiments, whose results are shown in Figures 4, 5 and 6. It is observed that Mn- and Co-doped catalysts showed better photocatalytic effectiveness than the commercially available P25, in terms of both bacteria inactivation (Figures 4 and 5). Although P25 TiO$_2$ is well known for its high photoreactivity due to the slow recombination of the electron-hole pair and large surface area [3], metal dopants improved the activity of catalysts considerably, as bacteria killing took place in almost 10 min of treatment. Moreover, it was observed that increasing the dopant concentration, disinfection efficiency was improved, while an increase beyond 0.3 wt% did not show any significant enhancement of the process.

Comparing Mn- and Co-, their effect was dependent of the experimental conditions in question and the type of the specific bacterial reference strains. In the case of *E. coli*, a 6-Log reduction was recorded in 10 min of solar irradiation using 0.1 wt% Mn-doped catalyst. Similar population reduction was achieved in longer treatment period (15 min) and at higher dopant concentration (1 wt%) when TiO$_2$ was doped with Co. Lower metal quantities than the aforementioned ones proved to be insufficient for total *E. coli* inactivation even after 30 min of treatment. As far as *K. pneumoniae* inactivation is concerned (Figure 6) it is observed that the 0.3 wt% Mn- and 1% Co-doped TiO$_2$ catalysts showed optimum photocatalytic performance when compared to the others and disinfection took place after only 10 min of treatment. Despite that this specific emerging pathogen is considered persistent during various treatments and disinfection techniques, photocatalysis with metal-doped catalysts seems quite
promising, since it may demonstrate complete inactivation in short periods with initial cell
densities as high as $10^5$ CFU/mL.

Findings of the current study highlight the acceleration of disinfection process when metal-
doped catalysts are employed. The higher concentrations of Co required is probably attributed
to the fact that as a transition metal, it may act as a recombination site for the photo-induced
charge carriers thus, lowering the quantum efficiency [8]. Fisher et al., who worked with *E. coli* and *Enterococcus faecalis* reported more rapid inactivation of both species in the
presence of TiO$_2$ doped with 1 wt% copper under solar irradiation compared to the respective
performed with undoped catalyst [1]. The bacterium-killing efficiency of doped TiO$_2$ is
referred to a quite extended variety of microorganisms, including *Listeria monocytogenes*,
*Shigella flexneri*, *Vibrio parahaemolyticus*, *Pseudomonas aeruginosa* and many others,
highlighting the advantage of dopants application in photocatalytic treatments [6, 23, 25, 28].

In further experiments, Mn/Co co-doped TiO$_2$ nanoparticles were successfully prepared
and their disinfection potential was tested under solar irradiation with *E. coli* and *K. pneumoniae* reference strains. Inactivation rates are shown in Figure 6. Co-doped catalysts
induced a more rapid total bacterial killing in comparison with those with single metal dopant
at respective concentration. Surprisingly, a 5-Log reduction of *K. pneumoniae* population was
recorded in 15 min with both co-doped TiO$_2$, while *E. coli* required 30 min for complete
elimination with the highest dopants concentration (0.1 wt%). Similar inactivation rates for *E. coli* were observed in a relevant study, where TiO$_2$ nanoparticles co-doped with N and Ag
were used for disinfection under visible light irradiation [32]. In another case, the application
of N and S co-doped P25 resulted in a 4-Log *E. coli* inactivation after 90 min of exposure to
visible light [33]. According to the general observation, composite dopants can compensate
the disadvantages of the individual components, inducing a synergistic effect [2, 8].
In order to further investigate the destruction of cellular structure of reference strains induced by photocatalysis, SEM was employed and selective images are shown in Figure 7. During photocatalysis the first oxidative stress is caused to bacteria when the catalyst nanoparticles interact with intact cells. The detrimental effect is expanded towards the cytoplasmic membrane, increasing cell permeability and allowing the outlet of intracellular components, which finally cause cell death [14, 25]. In many cases the resultant change in cell permeability is confirmed by potassium ion (K\(^+\)) leakage [34]. In the case of *K. pneumoniae* the remnants of polysaccharide capsules combined with material released from the cell are visible (Figures 7h to 7j). The progressive massive generation of hydroxyl radicals during the process overcomes any protection mechanism of bacterial cells, whose density in reaction mixture decreases with increasing time (Figures 4-6). In the course of treatment inactivation becomes slow, which, according to Vijay et al., is attributed to the protection provided to remaining active cells by metabolites excreted from the destructed ones [4].

### 3.2.3. Effect of initial bacterial concentration

A series of experiments was carried out to assess bacterial inactivation as a function of initial concentration of the cells in the reaction mixture. In general, an increase in bacterial density led to a decrease in the inactivation. Retardation of disinfection was more pronounced in the case of *E. coli*. For instance, when initial concentrations were 10\(^4\) and 10\(^6\) CFU/mL the period for total bacterial killing was 15 and 30 min, respectively, while complete inactivation was not achieved in the presence of higher concentration (10\(^8\) CFU/mL), where residual *E. coli* cells reached a plateau showing an overall decrease of 6 orders of magnitude. The almost same trend was recorded for *K. pneumoniae*, which was inactivated in 10 and 15 min when starting concentrations were 10\(^2\) and 10\(^3\) CFU/mL, respectively.
Disinfection rates can be fitted satisfactorily to a pseudo-first order kinetic expression, as can be seen in Figure 8; from the slopes of the resulting straight lines kinetic rate constants were calculated at 0.53 \( (r^2=0.96) \), 0.83 \( (r^2=0.92) \) and 0.77 min\(^{-1} \) \( (r^2=0.80) \) for initial \( E. \ coli \) concentration of \( 10^4 \), \( 10^6 \) and \( 10^8 \) CFU/mL, respectively. The corresponding values for \( K. \ pneumoniae \) were 0.43 \( (r^2=0.92) \), 0.57 \( (r^2=0.95) \) and 1.08 min\(^{-1} \) \( (r^2=0.99) \) for initial concentration of \( 10^2 \), \( 10^3 \) and \( 10^5 \) CFU/mL.

Usually, the disinfection ability of common techniques is inversely proportional to initial bacterial concentration. However, the required time for total inactivation depends on the tested bacterial species in each case. Residual cells of the reference strains after long treatment periods (> 30min) when high initial densities were employed, may be explained by the survival of a resistant subpopulation as a result of protection by clumping of microorganisms or even by genetically conferred resistance [35].

3.3 Why does doping enhance photocatalytic disinfection?

Results of the present study highlight the superiority of metal-doped catalysts compared to P25, in terms of bacterial removal in aqueous samples under simulated solar irradiation. In this respect, an attempt was made to identify the likely reasons for the improved activity of doped catalysts.

Firstly, a point of consideration should be the biocidal nature of the metals used for the preparation of catalysts. In some cases, metal-doped catalysts induce cell destruction of bacteria, due to the toxicity of metal ions released into the reaction solution [18]. This may occur even at minute metal concentrations according to the oligodynamic effect [36]. Accordingly, in order to assess the instantaneous toxicity of the novel Mn- and Co-doped catalysts to microorganisms during their treatment, a set of experiments was performed in the dark with the catalysts which had the lowest (0.02%) and highest (1%) concentration of metal
dopants. The catalyst concentration in the aqueous solution was 250 mg/L and initial bacterial concentration of both reference strains was $10^4$ CFU/mL. Bacterial density of *E. coli* and *K. pneumoniae* remained stable within the period of the treatment, which lasted almost 1 h. It is clearly shown that the metal-doped catalysts were not toxic to the selected reference strains regarding their short-term toxic effects. Many metals are toxic to microorganisms at micro- or millimolar concentrations, resulting in the development of resistance mechanisms in microbial cells. In the case of cobalt, it is required as a trace element in procaryotes and eucaryotes to fulfill a variety of metabolic functions. At high intracellular concentration the redox active metal ion Co$^{2+}$ is highly toxic [37]. According to our results, no bacterial inactivation was recorded in the presence of both catalysts, indicating that no stress was induced in the cells during their exposure to both metals.

Secondly, doping shifted catalyst absorption to the visible region up to 600 nm, as well as decreased the band gap energy as clearly shown in Figure 2 and Table 2. Therefore, it was possible to use the main part of the solar spectrum and surpass one of P25 drawbacks, which is its low activity beyond the range of UV light. To investigate this effect further, experiments were performed using a filter to cut-off UV light below 420 nm. The exclusion of UV light led to a considerable inactivation of bacterial populations, which was more pronounced in the case of *E. coli*. Specifically, in the presence of 0.1 and 1 wt% of Mn- and Co-doped TiO$_2$ the overall decrease of *E. coli* cells after 30 min of treatment was 74.3% and 94%, respectively (data for *E. coli* and Mn are shown in Figure 9a; for the sake of brevity data with Co are not shown). For the same period of time and with the use of 1 wt% Mn- and Co-doped TiO$_2$, the respective inactivation of *K. pneumoniae* reached 28% and 44% (data for *K. pneumoniae* and Co are shown in Figure 9b; for the sake of brevity data with Mn are not shown). Noticeably, the commercial P25 catalyst exhibited no photocatalytic activity above 420 nm and bacterial populations of both strains remained intact throughout the process (data not shown). The
metal-doped catalysts (0.1 wt% Mn and 1 wt% Co) showed satisfactory photocatalytic activity within the visible spectral range of the provided irradiation, causing detrimental effects on the bacteria, which partly inactivated the cells in the reaction mixture. Their efficiency was improved with the addition of UV light, as bacterial populations reached zero levels within 10 min of treatment. The contribution of UV light to the overall photocatalytic activity is clearly shown in Figure 9 as some experiments were also performed at reduced photon fluxes. Total removal of *E. coli* and *K. pneumoniae* occurred within 10 min only at $5.8 \times 10^{-7}$ einstein/(L.s), while three-fold or even longer periods were required to achieve the same inactivation at $5.3 \times 10^{-7}$ and $4.93 \times 10^{-7}$ einstein/(L.s). The detrimental effect of decreasing flux to disinfection performance was more pronounced in the case of *K. pneumoniae*, highlighting the resistant nature of the strain.

These findings verify the fact that chemical doping may act as a satisfactory means for sensitization of TiO$_2$ in the visible light region, which is responsible for the enhanced photocatalytic activity recorded in this work. Moreover, dopants on the surface of TiO$_2$ may act as an electron trap, thus promoting interfacial charge transfer and delaying the recombination of the light-induced electron-hole pair [38], which also leads to enhanced activity.

4. Conclusions

The present study focused on solar photocatalytic disinfection with the use of novel cobalt- and manganese-doped titania materials and reference strains of *E. coli* and *K. pneumoniae*.

- Metal-doped TiO$_2$ was prepared successfully and doping shifted the optical absorption edge to the visible region.

- Dopants significantly enhanced (by a factor of 2-3) the photocatalytic activity of TiO$_2$ under solar irradiation, in terms of both bacteria inactivation.
Dopants concentrations affected the overall process up to a certain level, while an increase beyond 0.3 wt% did not show significant enhancement of disinfection rates. It is implied that doping levels >0.3 wt% introduce heavy modification of the TiO$_2$ semiconductor, thus hindering the production/diffusion of charges under light and, consequently, precluding the effective interaction of these charges to inactivate bacteria.

Comparing Mn and Co dopants, their performance varied depending on operating parameters, such as the reference strain and the initial bacterial concentration.

All catalysts were effective for the removal of *K. pneumoniae*, which is considered as an opportunistic pathogen highly resistant in various water treatments.

The improved activity of metal-doped titania is accredited to the optical absorption shifts towards the visible region and to the recombination delay of the electron-hole pair, since metals did not exhibit any bactericidal properties and catalysts were considerably sensitized in the absence of UV light.

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References


Table 1.

Average particle size and phase composition for Mn-, Co- and Mn/Co-doped TiO₂ catalyst.

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<th>Dopant concentration (molar ratio)</th>
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Table 2. Effect of Mn and Co dopant level on band gap.

<table>
<thead>
<tr>
<th>Dopant concentration (molar ratio)</th>
<th>Mn-doped TiO$_2$</th>
<th>Co-doped TiO$_2$</th>
<th>Mn/Co-doped TiO$_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Indirect band gap (eV)</td>
<td>Indirect band gap (eV)</td>
<td>Indirect band gap (eV)</td>
</tr>
<tr>
<td>0.02</td>
<td>2.7</td>
<td>2.97</td>
<td></td>
</tr>
<tr>
<td>0.04</td>
<td>2.85</td>
<td>2.85 [1.6$^*$]</td>
<td>3 [1.6$^*$]</td>
</tr>
<tr>
<td>0.1</td>
<td>2.75</td>
<td>2.83 [1.55$^*$]</td>
<td>2.7 [1.5$^*$]</td>
</tr>
<tr>
<td>0.3</td>
<td>2.6</td>
<td>2.7 [1.5$^*$]</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>2.4</td>
<td>2.4 [1.41$^*$]</td>
<td></td>
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</tbody>
</table>

$^*$ Sub-band gap
Figure captions

Figure 1
XRD patterns of a) Mn-doped TiO$_2$ (0.02-1 wt%), b) Co-doped TiO$_2$ (0.02-1 wt%) and c) Mn & Co co-doped TiO$_2$ (binary dopant concentration range: 0.04-0.1 wt%).

Figure 2
UV-Vis absorption of P25 and a) Mn-doped TiO$_2$ (0.02-1 wt%), b) Co-doped TiO$_2$ (0.02-1 wt%) and c) Mn & Co co-doped TiO$_2$ (binary dopant concentration range: 0.04-1 wt%).

Figure 3
TEM images of a) Mn-doped TiO$_2$ (1 wt%) and b) Co-doped TiO$_2$ (1 wt%).

Figure 4
E. coli inactivation in the presence of different Mn- and Co-doped TiO$_2$ catalysts and the commercially available TiO$_2$ (P25, Evonik). Catalyst concentration is 100 mg/L.

Figure 5
K. pneumoniae inactivation in the presence of different a) Mn- and b) Co-doped TiO$_2$ catalysts and the commercially available TiO$_2$ (P25, Evonik). Catalyst concentration is 250 mg/L.

Figure 6
E. coli and K. pneumoniae inactivation in the presence of Mn & Co co-doped TiO$_2$. Catalyst concentration is 100 and 250 mg/L for E. coli and K. pneumoniae inactivation, respectively.

Figure 7
SEM images of E. coli (a-e) and K. pneumoniae (f-j) without treatment (negative controls) and after photocatalytic treatment in the presence of metal-doped TiO$_2$.

Figure 8
Effect of initial bacterial concentration on disinfection efficiency for a) E. coli (conditions: 0.02 wt% Co, 100 mg/L) and b) K. pneumoniae (conditions: 0.3 wt% Mn, 250 mg/L).
Figure 9

Effect of photon flux and wavelength on the inactivation of a) *E. coli* (conditions: 0.1 wt% Mn, 100 mg/L) and b) *K. pneumoniae* (conditions: 1 wt% Co, 250 mg/L).
Figure 1

(a) $x_a = 86\%$

Intesity (a.u.)

(b) $x_a = 50.4\%$

Intesity (a.u.)

(c) $x_a = 76\%$

Intesity (a.u.)

$0.02\text{ Mn/TiO}_2$

$0.04\text{ Mn/TiO}_2$

$0.1\text{ Mn/TiO}_2$

$0.3\text{ Mn/TiO}_2$

$1\text{ Mn/TiO}_2$

$P25\text{ TiO}_2$

$0.02\text{ Co/TiO}_2$

$0.04\text{ Co/TiO}_2$

$0.1\text{ Co/TiO}_2$

$0.3\text{ Co/TiO}_2$

$1\text{ Co/TiO}_2$

$P25\text{ TiO}_2$

$0.02\text{ Co-Mn/TiO}_2$

$0.04\text{ Co-Mn/TiO}_2$

$0.1\text{ Co-Mn/TiO}_2$

$0.3\text{ Mn/TiO}_2$

$1\text{ Co-Mn/TiO}_2$
Figure 2

(a) Absorbance spectrum of P25 and different Mn-doped TiO$_2$ samples.

(b) Absorbance spectrum of P25 and different Co-doped TiO$_2$ samples.

(c) Absorbance spectrum of P25 and different Mn/Co-doped TiO$_2$ samples.
Figure 3

(a) [Image of TEM micrograph] 50 nm

(b) [Image of TEM micrograph] 50 nm
Figure 4

(a) E. coli inactivation, CFU/mL

(b) E. coli inactivation, CFU/mL

Time, min

- 0.02% Mn:TiO\textsubscript{2}
- 0.04% Mn:TiO\textsubscript{2}
- 0.1% Mn:TiO\textsubscript{2}
- 0.3% Mn:TiO\textsubscript{2}
- 1% Mn:TiO\textsubscript{2}
- P25

Time, min

- 0.02% Co:TiO\textsubscript{2}
- 0.04% Co:TiO\textsubscript{2}
- 0.1% Co:TiO\textsubscript{2}
- 0.3% Co:TiO\textsubscript{2}
- 1% Co:TiO\textsubscript{2}
- P25
Figure 5

(a) K. pneumoniae inactivation, CFU/mL

(b) K. pneumoniae inactivation, CFU/mL

- 0.02% Mn:TiO2
- 0.04% Mn:TiO2
- 0.1% Mn:TiO2
- 0.3% Mn:TiO2
- 1% Mn:TiO2
- P25

- 0.02% Co:TiO2
- 0.04% Co:TiO2
- 0.1% Co:TiO2
- 0.3% Co:TiO2
- 1% Co:TiO2
- P25
Figure 6

[Graph showing bacterial inactivation of K. pneumoniae and E. coli with different Mn/Co:TiO2 concentrations over time.]

- K. pneumoniae 0.04% Mn/Co:TiO2
- K. pneumoniae 0.1% Mn/Co:TiO2
- E. coli 0.04% Mn/Co:TiO2
- E. coli 0.1% Mn/Co:TiO2
Figure 7

a) E. coli - intact cells
b) 0.1 % Mn TiO₂ – 30min

c) 0.02 % Co TiO₂ – 30min
d) 0.04 Co/Mn co-doped TiO₂ – 30min

e) 0.1 % Mn/Co co-doped TiO₂ – 30min
f) K. pneumoniae – intact cells
Figure 8

a

\[ \ln\left(\frac{C_0}{C}\right) \]

\begin{align*}
\text{Time, min} & \quad 0 & \quad 5 & \quad 10 & \quad 15 \\
1.0E+08 \text{ CFU/mL} & \quad \Delta & & \quad \Delta \\
1.0E+06 \text{ CFU/mL} & \quad \Box & & \quad \Box \\
1.0E+04 \text{ CFU/mL} & \quad \bullet & & \quad \bullet \\
0 & & & & \end{align*}

b

\[ \ln\left(\frac{C_0}{C}\right) \]

\begin{align*}
\text{Time, min} & \quad 0 & \quad 2 & \quad 4 & \quad 6 & \quad 8 & \quad 10 \\
1.0E+05 \text{ CFU/mL} & \quad \Delta & \quad \Delta & \quad \Delta & \quad \Delta \\
1.0E+03 \text{ CFU/mL} & \quad \blacksquare & \quad \blacksquare & \quad \blacksquare & \quad \blacksquare \\
1.0E+02 \text{ CFU/mL} & \quad \blacklozenge & \quad \blacklozenge & \quad \blacklozenge & \quad \blacklozenge \\
0 & & & & \end{align*}
Figure 9

a

![Graph showing E. coli inactivation, CFU/mL vs. Time, min.](image)

- cut on 420nm
- 4.93 1E-7 einstein/(L.s)
- 5.3 1E-7 einstein/(L.s)
- solar/5.8 1E-7 einstein/(L.s)

b

![Graph showing K. pneumoniae inactivation, CFU/mL vs. Time, min.](image)

- cut on 420 nm
- 4.93 1E-7 einstein/(L.s)
- 5.3 1E-7 einstein/(L.s)
- solar/5.8 1E-7 einstein/(L.s)