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The Gut Microbiome versus COVID-19

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Abstract

The digestive system is an environmental frontline involving digestive secretions, intestinal cell metabolism, and gut microbiome that significantly modulate multiple functions in organisms. Understanding the 'gut-lung axis', where gut residential microbiota play important roles, may help in the development of better prophylactics and intervention strategies for diseases caused by respiratory viruses, including coronavirus disease of 2019 (COVID-19). Gastrointestinal symptoms are common in COVID-19 patients and are generally indicative of disease complications. As we have learned so far, diarrhea and gut dysbiosis during SARS-CoV-2 infection should not be ignored, as they can be used to distinguish pathways of dysregulation of the immune system and the regulatory pathways upstream and downstream of viral primary binding receptors such as ACE2. This review presents evidence of microbiome signatures in the gut and respiratory system that may predict the severity and long-term outcomes of COVID-19. Understanding the factors (such as pro-inflammatory trends, modulation of metabolite availability, and impact on cell signaling and pathogenic properties) translating the effect of microbiome composition on the severity of respiratory infections should help in the development of new approaches for health monitoring, disease prevention, and treatment.

Keywords: GM • Microbiome • COVID-19 • SARS-CoV-2 • Dys-biosis • ACE2 • Immune system • Gut-lung axis

Abbreviations: SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus 2 • SARS: Severe Acute Respiratory Syndrome • COVID-19: Coronavirus Disease of 2019 • MERS: Middle East Respiratory Syndrome • RSV: Respiratory Syncytial Virus • GM: Gut Microbiome • GI: Gastrointestinal • IBD: Irritable Bowel Disease • ACE2: Angiotensin Converting Enzyme 2 • IFN-I: Type I Interferon (IFN- α) • B(O)AT1: Sodium-dependent Neutral Amino Acid Transporter • mTOR: The Mammalian Target of Rapamycin.

Introduction

The digestive system is an environmental frontline involving digestive secretions, intestinal cell metabolism, and gut microbiome (GM) [1,2] that significantly modulate multiple organism's functions [3]. Nutrients and prebiotics provide substrates for a dynamic GM, which is estimated to consist of over 1000 different microbial species belonging to five predominant phyla: Firmicutes, Bacteroidetes, Actinobacteria, Verrucomicrobia, and Proteobacteria [4-7]. Approximately 400 identified species are strictly anaerobic and hence will generally be found in mucosal regions such as the oral cavity and the gastrointestinal (GI) tract [2,6].

Two thirds of the GM is considered an individual-specific 'fingerprint' [2,8]; however, there are correlations found between GM composition and diet [1,9-11], geography, ethnicity [11-15], age, and longevity of the host [10,14-17], and certain diseases [8,18-22]. GM characteristics of members of the same family cluster in functional and taxonomic space, stressing the role of environmental and genetic factors in the evolution of an individual's GM [23].

Gut virome composition also appears to be unique to and varies with people's age and geography, with few viral populations shared within a subset of people. There are 33,242 unique viral populations (97.7% of which are phages) that have been identified in the human gut [24]. As is the case with the microbiome, consistent variations in viral diversity could also be seen in healthy versus sick participants [25].

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The current global COVID-19 pandemic [26] is caused by a beta coronavirus, severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2), that in the most vulnerable individuals can cause an immune response resembling cytokine storm syndrome [27], a dysregulated secretion of cytokines that triggers systemic inflammation and hypercoagulation, and, in children, a multisystem inflammatory syndrome that resembles Kawasaki disease [28]. Approximately 20% of patients with COVID-19 develop serious complications, often accompanied by fatal acute respiratory distress syndrome.

Increasing evidence points to an intimate relationship between the gastrointestinal and respiratory tracts, which is known as the 'gut-lung axis' [29-31]. The intestinal flora is involved in host nutrient absorption and metabolism and has a profound impact on all organ systems, human health, and disease [7,32-35].

However, intestinal microflora disorders reduce host antiviral immune responses, thereby aggravating the lung damage caused by these infections [36]. Changes in the lung microbiota were identified in COVID-19 patients, with a shift towards bacteria found in the intestinal tract correlating with the onset of acute respiratory distress syndrome. Probiotics have already been recommended for anti-viral therapy and prophylactics [37,38].

Literature Review

In this review, we bring together data pointing to the potential significance of modulation of the gut microbial community and its functional influence on the organism's immunity and resistance to prophylactics and treatment of COVID-19 disease. These factors should be addressed as we prepare for future challenges.

COVID-19 gastrointestinal manifestations

GI symptoms in COVID-19 patients are generally indicative of complications. However, they are common in these patients, with a meta-analysis showing that the symptoms were present in 17.6% of infected patients

[39,40]. Similarly, approximately 25% of patients with SARS and MERS exhibited GI symptoms [41,42].

Studies have detected SARS-CoV-2 RNA in anal swabs and stool samples from almost 50% of patients with COVID-19 [43-46]. mRNA analysis demonstrated that the duration of viral shedding from stool was longer than that from respiratory samples [47], suggesting that the digestive tract might be a site for virus replication and activity. Prolonged digestive symptoms, especially diarrhea, are negatively correlated with gut microbiota richness and composition [48,49], and are accompanied by elevated levels of fecal calprotectin, an indicator of inflammatory responses in the gut [50]. It should, however, be noted that gut microbiome dysbiosis is also characteristic of diabetes, obesity, autoimmune, and aging-related diseases, all of which are associated with an increased risk of severe COVID-19 [13].

GM dysbiosis is associated with COVID-19 disease

Comorbidities commonly accompanying severe COVID-19 are known to be associated with alterations in bacterial taxa from the phyla Bacteroidetes and Firmicutes [51-54], which were reported to regulate angiotensin-converting enzyme 2 (ACE2) expression in rodents [55]. Assessment of GMs of COVID-19 hospital patients conducted in China [56] showed that the abundance of *Coprobacillus*, *Clostridium ramosum*, and *Clostridium hathewayi* species positively correlate, while the abundance of *Faecalibacterium prausnitzii* (an anti-inflammatory bacterium) negatively correlates with COVID-19 severity. During hospitalization, abundances of *Bacteroides dorei*, *Bacteroides thetaiotaomicron*, *Bacteroides massiliensis*, and *Bacteroides ovatus* were inversely correlated with SARS-CoV-2 load in fecal samples from the patients. However, gut dysbiosis persisted even after clearance of SARS-CoV-2 from the throat and resolution of respiratory symptoms.

Changes in GM composition may be indicative of shifts in nutritional availability in the gut, which could be due to the diet, pharmacological interventions, or absorptive properties of the gut endothelium. Fecal samples with high SARS-CoV-2 mRNA load had an abundance of bacterial species with higher capacity for glycolysis, de novo nucleotide synthesis, and amino acid biosynthesis (e.g., *Collinsella aerofaciens*, *Collinsella tanakaei*, *Streptococcus infantis*, *Morganella morganii*), whereas fecal samples with low-to-none SARS-CoV-2 mRNA had a higher ratio of short-chain fatty acid-producing bacteria [57,58] of the *Parabacteroides*, *Bacteroides*, *Alistipes*, and *Lachnospiraceae* genera [59]. Another important finding was the significant depletion of butyrate-producing bacteria in H1N1 patients versus healthy controls [60,61], suggesting that butyrate may be involved in the modulation of inflammation during viral pneumonia. This is supported by the observation that a high-fiber diet, which increases the production of short-chain fatty acids, enhances the antiviral CD8+ T-cell immune response during influenza virus infection, attenuates neutrophil-mediated lung injury, and consequently improves survival [36,62]. MRx-4DP0004, a strain of the butyrate-producing bacterium *Bifidobacterium breve*, originally developed for asthma treatment [63], has shown the potential to downregulate specific pathological aspects of the hyper-inflammatory response while maintaining the appropriate antiviral response. However, significantly reduced levels of butyrate producers were linked to overgrowth of pathogenic bacteria and increased intestinal mucosal permeability and endotoxin intoxication and, consequently, inflammation with cytokine release [64-66]. Recent studies further suggest that gut microbiome composition may predict a predisposition to severe COVID-19 due to a hyper-inflammatory response [49].

Gut microbiome and viral infections shift in GM composition

An increase in the proinflammatory component in GM has also been observed for other respiratory viruses such as influenza [67]. In a lemur model, several commensal taxa, essential for a healthy gut microbiome, decreased, whereas the abundance of potential pathogens, such as *Neisseria*, increased upon adenovirus infection [68].

The clinical manifestations and transmission routes of seasonal influenza A (H1N1) are similar to those of COVID-19 [69], and a comparison of the microbiota in these two diseases was of interest from etiological and diagnostic

perspectives. The microbial signature associated with detectable levels of SARS-CoV-2 mRNA was similar to that of other respiratory viruses such as influenza and respiratory syncytial virus (RSV) (ignoring the small sample size of the study) [70]. However, patients with H1N1 displayed lower diversity and different overall microbial composition compared with COVID-19 patients, and biomarkers were proposed for distinguishing the two cohorts [71].

COVID-19 patients still had significantly reduced GM bacterial diversity, significantly higher relative abundance of opportunistic pathogens such as *Streptococcus*, *Rothia*, *Veillonella*, and *Actinomyces*, and lower relative abundance of beneficial symbionts, compared with healthy controls. Changes in the enteric environment and immune factors caused by *Actinomyces*, particularly, were shown to aggravate the damage caused by inflammatory bowel disease [60,72], and an association of *Actinomyces* with COVID-19 infection may also have a prognostic value.

Compared with healthy individuals, levels of *Agathobacter*, *Fusicatenibacter*, *Roseburia*, and *Ruminococcaceae* UCG-013 in COVID-19 patients were depleted and negatively correlated with levels of inflammatory markers (CRP, procalcitonin, or D-dimer). Moreover, CRP and D-dimer levels were positively correlated with COVID-19-enriched bacteria (*Streptococcus*, *Rothia*, *Veillonella*, and *Actinomyces*). In the H1N1 cohort, there was a positive correlation between inflammatory cytokines; IL-2, IL-4, and IL-6, and abundance of *Fingoldia*, *Anaerococcus*, *Peptoniphilus*, *Intestinibacter*, and *Prevotella* genera [71].

Understanding whether the composition of the GM can be modified to improve the outcome of viral respiratory infections, particularly in COVID-19 patients, is of great significance.

Immune system modulation by gut microbiome in COVID-19 and other viral diseases

One of the ways in which microbiota can be involved in disease progression is by affecting the inflammatory state of the gut and the systemic levels of proinflammatory cytokines.

Higher abundance of *Klebsiella*, *Streptococcus*, and *Ruminococcus* genera correlated with elevated levels of proinflammatory cytokines and increased disease severity, while increased levels of *Lactobacillus* species correlated with higher levels of anti-inflammatory IL-10 and improved disease prognosis [49]. Cases of COVID-19-induced Kawasaki disease-like complications in young children [28,73] were also associated with dysbiotic gut microbiome, with increased levels of *Streptococcus* and decreased levels of *Lactobacillus* species compared with healthy children [74]. Compared with healthy individuals, COVID-19 patients had significantly higher systemic levels of IL-6 and TNF- α , and levels of cytokines IL18 and IgA in the gut were also affected by COVID-19 infection [75]. High IL18 and IgA levels correlated with overrepresentation of the previously mentioned *Streptococcus* genera, as well as *Clostridium* and *Bifidobacterium*. Increased abundance of *Lactobacillus* was correlated with high IL18 expression and poor prognosis, as well as a lack of the genera *Bacteroidetes*, *Roseburia*, *Faecalibacterium*, *Coprococcus*, and *Parabacteroides* in COVID-19 patients relative to healthy controls [75].

The proinflammatory gut environment is characteristic of patients suffering from a range of conditions, including diabetes, obesity, irritable bowel disease (IBD), and high blood pressure, which are typically correlated with the severity of COVID-19. One can expect some degree of causation between GM composition and the disease-promoting environment; however, the vector in the link still needs to be validated in larger patient cohorts or in an experimental setting (although this option is more problematic).

Both innate and adaptive immune system responses are triggered by SARS-CoV-2 infection. However, in severe COVID-19 patients, the numbers of B cells, CD4+ and CD8+ T cells, and monocytes are lower [76,77]. GM is a known modulator of the immune system [78,79], and the immune system plays a leading role in gut colonization by microbiota, as the development of regulatory T cells and innate lymphoid cells help maintain gut and lung homeostasis. Microbial metabolites regulate the host immune system [80-83] and may also participate in the immune response triggered by viral replication in the gut.

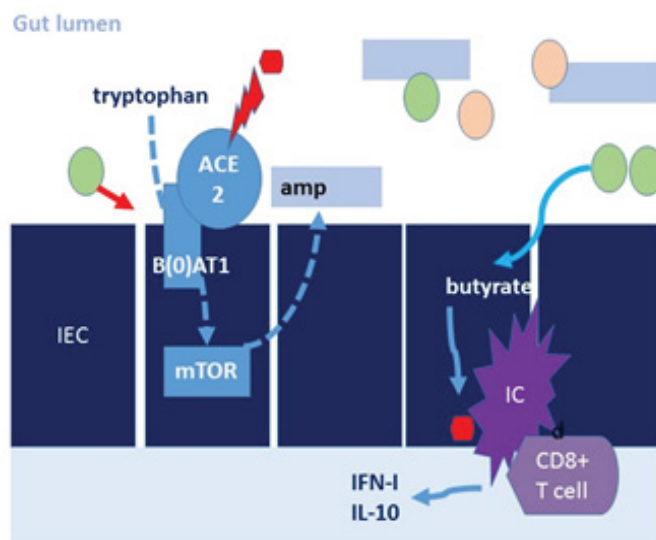


Figure 2. Role of the gut-lung axis in COVID-19. Red arrows represent suppression and blue arrows represent activation of downstream processes.

Table 1. Statements and evidences.

Main Statements, Evidence	References
Compositions of GM and lung commensal microbiomes correlate with systemic changes in a state of immune system	
GM influences immune response to respiratory viral infections (gut-lung axis).	[37,38]
A balance of the microbiota is necessary for homeostasis: a comprehensive study of immune system modulation by GM in mice.	[55]
Lung commensal microbiota critically regulates the generation of virus-specific CD4 and CD8 T-cells and antibody responses following respiratory influenza virus infection.	[36]
Gut-associated <i>Bacteroides</i> presence in lung positively correlates with elevated inflammatory markers in plasma.	[107]
GM composition changes in adenovirus and H1N1 infections.	[68-71]
Lung-infection derived CCR9+CD4+ T-cells can be recruited to the small intestine.	[88,89]
Influenza virus infection induces microbiota-mediated Th17 cell-dependent inflammation.	[88]
IFN-I molecules produced in the lung supported a depletion of obligate anaerobic bacteria and enrichment of Enterobacteriaceae in the gut.	[61]
GM composition and systemic immune markers correlate.	[49,50,71,36,62,73,75-83]
Microbiota signatures in the gut and respiratory system may predict severity and long-term outcomes in COVID-19 patients	
COVID-19 severity negatively correlates with <i>Bacteroides</i> , Firmicutes species abundancies.	[49,51-54,73,74]
COVID-19 severity positively correlates with <i>Coprobacillus</i> , <i>Clostridium ramosum</i> , and <i>Clostridium hathewayi</i> .	[56]
Negative correlations with <i>Faecalibacterium prausnitzii</i> , <i>Bacteroides</i> species.	[56]
<i>Bacteroides</i> genus, <i>Streptococcus</i> genus and <i>Clostridiales</i> order were negatively correlated with the inflammatory cytokines, <i>Ruminococcus</i> genus, <i>Blautiagenus</i> and <i>Lactobacillus</i> genus showed positive associations.	[49]
IL-2, IL-4, and IL-6 levels correlate with abundancies of <i>Fingoldia</i> , <i>Anaerococcus</i> , <i>Peptoniphilus</i> , <i>Intestinibacter</i> , and <i>Prevotella</i> genera.	[71]
GM may play a causative role in gut-lung axis	
GM modulates absorption of nutrients that may play a role in aetiology of systemic or organ pathologies.	[29-31,32-35,57]
Microbial metabolites (short-chain FA, desaminotyrosine) regulate the immune system.	[49,58,64,65,78,80-87]
GM modulates host's immune system.	[49,60-66]
GM effects ACE2 expression in the gut.	[90,97,98]
Intestinal microflora disorders reduce host antiviral immune response.	[36,32-35]
Changes in GM composition may effect viral infection outcome	
Immune system modulation is an intermediate step in the gut-lung axis	[60-66]
Nutrient absorption and intestinal microflora disorders reduce host antiviral immune response.	[29-31,32-35,36]
Immune system modulations in viral pneumonia correlate with the GM composition.	[36,49,62]
<i>Bifidobacterium breve</i> positively modulates an outcome of viral pneumonia.	[65]
dysbiosis causes changes in intestinal permeability and endotoxin intoxication.	[64,65]
GM composition shifts are linked to hiperinflammatory response.	[49]
Influenza-specific CD8+ T-cell function and Type I interferon (IFN-I) signalling in macrophages are associated with GM signalling.	[49,62,65,78,85-88]
IL18, IGA levels can be affected by GM Firmicutes abundancies.	[49,75]
Microbial metabolites regulate immune system.	[49,64,80-83]
IL-2, IL-4, and IL-6 systemic levels correlate to <i>Fingoldia</i> , <i>Anaerococcus</i> , <i>Peptoniphilus</i> , <i>Intestinibacter</i> , and <i>Prevotella</i> abundancies	[71]
ACE receptors expression is influenced by GM and, in their turn, influence GM composition	[90,97,98]
Pre- and probiotics therapies may be complementary to vaccines, treatment, and during patients' rehabilitation	
Richness of GM composition is decreased in pathologies associated with a viral infection severity.	[3,37,38,48,49]

Immune system can be modulated via balancing a GM composition.	[60-66,85]
High fibre diet supports positive outcome in viral pneumonia.	[36,62]
GM-produced metabolites modulate anti-viral immune response.	[58,62-65,78,80-87]
<i>Bifidobacterium breve</i> supplementation positively modulates an outcome of viral pneumonia.	[66]

plasma [107]. The relative abundances of opportunistic pathogens such as *Streptococcus*, *Rothia*, *Veillonella*, *Erysipelatoclostridium*, and *Actinomyces*, were also increased in the lungs of COVID-19 patients. Among these, *Rothia* is already known to contribute to the pathogenesis of pneumonia, especially in immune compromised individuals [108]. *Streptococcus* and *Rothia* were also associated with susceptibility to secondary bacterial lung infection in patients with avian H7N9 virus infection [109], and the opportunistic pathogens *Prevotella*, *Fingoldia*, and *Peptoniphilus*, were enriched in lung microbiomes of H1N1 patients [60,71].

Discussion

The severity of COVID-19 is associated with gastrointestinal symptoms [39,40,45], and we can suggest with high confidence that the digestive tract may be a site for virus replication and activity [43-47].

Prolonged diarrhea in COVID-19 patients is negatively correlated with gut microbiota richness and composition [48,49] and positively correlated with markers of ongoing inflammation [50]. Such associations are typical of dysbiosis, and dysbiosis is typical in diabetes, obesity, and autoimmune and aging-related diseases, which are comorbidities associated with severe COVID-19 outcomes [13,51-54]. Modulation of some of the listed diseases by pre- and probiotics has already been considered and shown to be successful [7,9,16,19-21,31,51-54]. We suggest that a similar approach may be effective with respect to prophylactic, and treatment of COVID-19 and other viral diseases [36-38,62, 66,90,96] as well as along the vaccination [110].

Alterations in Bacteroidetes and Firmicutes [28,49,51-54,71,74] taxa were the most common in severe COVID-19 cases, with an increase in the abundance of species relying on their own basic nucleotide and amino acid biosynthesis and glycolysis, and a decrease in the abundance of short-fatty acid producers [56-59]. The latter are also known to possess anti-inflammatory properties [60-66,71,78, 81-87], and modulate ACE2 receptor expression and amino acids metabolism (Figure 1) in the gut lining [49,55,97,98]. Thus, fiber-based diets, butyrate-rich fermented products, enrichment of a diet for particular cofactors involved in butyrate-generating pathways, and decreased sugar intake would play beneficial roles in GM-led protection from inflammation. ACE2 and its inhibitors have been explored as potential targets [90,97] and targeting of amino acid transport and biosynthesis in bacteria or amino acid supplementation in the host [49,97] can be also considered.

Changes in the composition of GM may result in loss of antiviral resistance and respiratory system complications [55,83,85,87]. However, respiratory viral infections may also lead to changes in GM composition [89,90] (Figure 2). Interesting correlation data [36,51-59,68-71,75] suggest that a link exists between the niches (gut-lung and gut-immune system); however, the causative vectors of most observed correlations have not yet been defined. We propose that the complex system of cross-regulation in the case of GM-led pathology (Figure 2) may be modulated by careful targeting of immune components (such as proinflammatory interleukins, for instance). The main evidence supporting our statement has been collated and presented in Table 1 for evaluation.

Conclusion

We believe that further understanding of the physiological impact of the 'gut-lung axis' may help in the development of better prophylactic and intervention strategies for COVID-19 and other diseases caused by respiratory viruses. The dysbiosis manifestation and microbiota signatures in the gut and respiratory system, associated with higher mortality risk and long-term outcomes in COVID-19 patients, may be used in health monitoring during and after epidemics and be addressed by new pre- and probiotics therapies.

Conflict of Interest

There are no conflicts of interest associated with this publication.

References

- Ma, Ning, Yanan Tian, Yi Wu, and Xi Ma. "Contributions of the interaction between dietary protein and gut microbiota to intestinal health." *Curr Protein Peptide Sci* 18 (2017): 795-808.
- Integrative, HMP, Lita M Proctor, Heather H Creasy, and Jennifer M Fettweis, et al. "The integrative human microbiome project." *Nat* 569 (2019): 641-648.
- Qin, Yufeng, and Paul A Wade. "Crosstalk between the microbiome and epigenome: messages from bugs." *J Biochem* 163 (2018): 105-112.
- Thursby, Elizabeth, and Nathalie Juge. "Introduction to the human gut microbiota." *Biochem J* 474 (2017): 1823-1836.
- Rajilić-Stojanović, Mirjana, and Willem M De Vos. "The first 1000 cultured species of the human gastrointestinal microbiota." *FEMS Microbiol Rev* 38 (2014): 996-1047.
- Hugon, Perrine, Jean-Charles Dufour, Philippe Colson, and Pierre-Edouard Fournier, et al. "A comprehensive repertoire of prokaryotic species identified in human beings." *The Lancet Infect Dis* 15 (2015): 1211-1219.
- Huttenhower, Curtis, Dirk Gevers, Rob Knight, and Sahar Abubucker, et al. "Structure, function and diversity of the healthy human microbiome." *Nat* 486 (2012): 207.
- Walker, Alan W, Jennifer Ince, Sylvia H Duncan, and Lucy M Webster, et al. "Dominant and diet-responsive groups of bacteria within the human colonic microbiota." *The ISME J* 5 (2011): 220-230.
- Claesson, Marcus J, Ian B Jeffery, Susana Conde, and Susan E Power, et al. "Gut microbiota composition correlates with diet and health in the elderly." *Nat* 488 (2012): 178-184.
- Senghor, Bruno, Cheikh Sokhna, Raymond Ruimy, and Jean-Christophe Lagier. "Gut microbiota diversity according to dietary habits and geographical provenance." *Human Microbiome J* 7 (2018): 1-9.
- Zhernakova, Alexandra, Alexander Kurilshikov, Marc Jan Bonder, and Ettje F Tigchelaar, et al. "Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity." *Sci* 352 (2016): 565-569.
- Fortenberry, J Dennis. "The uses of race and ethnicity in human microbiome research." *Trends Microbiol* 21 (2013): 165-166.
- Gupta, Vinod K, Sandip Paul, and Chitra Dutta. "Geography, ethnicity or subsistence-specific variations in human microbiome composition and diversity." *Front Microbiol* 8 (2017): 1162.
- Falony, Gwen, Marie Joossens, Sara Vieira-Silva, and Jun Wang, et al. "Population-level analysis of gut microbiome variation." *Sci* 352 (2016): 560-564.
- Brooks, Andrew W, Sambhawa Priya, Ran Blekhan, and Seth R Bordenstein. "Gut microbiota diversity across ethnicities in the United States." *PLoS Biol* 16 (2018): e2006842.
- Arbolea, Silvia, Claire Watkins, Catherine Stanton, and R Paul Ross. "Gut bifidobacteria populations in human health and aging." *Front Microbiol* 7 (2016): 1204.
- Yatsunenkov, Tanya, Federico E Rey, Mark J Manary, and Indi Trehan, et al. "Human gut microbiome viewed across age and geography." *Nat* 486 (2012): 222-227.
- Goodrich, Julia K, Jillian L Waters, Angela C Poole, and Jessica L Sutter, et al. "Human genetics shape the gut microbiome." *Cell* 159 (2014): 789-799.
- Schroeder, Bjoern O, and Fredrik Bäckhed. "Signals from the gut microbiota to distant organs in physiology and disease." *Nat Med* 22 (2016): 1079.
- Peterson, Daniel A, Daniel N Frank, Norman R Pace, and Jeffrey I Gordon.

- "Metagenomic approaches for defining the pathogenesis of inflammatory bowel diseases." *Cell Host Microbe* 3 (2008): 417-427.
21. Finegold, Sydney M. "Desulfovibrio species are potentially important in regressive autism." *Med Hypoth* 77 (2011): 270-274.
 22. Midtvedt, Tore. "The gut: a triggering place for autism—possibilities and challenges." *Microb Ecol Health Dis* 23 (2012): 18982.
 23. Vasieva O, Sorokin A, Murzabaev M, and Babiak P, et al. Study on the analysis of personal gut microbiomes. *J Comput Sci Syst Biol* 12 (2019): 71–79.
 24. <https://phys.org/news/2020-08-human-gut-viral-fingerprint.html>
 25. Gregory, Ann C, Olivier Zabolocki, Ahmed A Zayed, and Allison Howell, et al. "The gut virome database reveals age-dependent patterns of virome diversity in the human gut." *Cell Host Microbe* 28 (2020): 724-740.
 26. Chan, Jasper Fuk-Woo, Shuofeng Yuan, Kin-Hang Kok, and Kelvin Kai-Wang To, et al. "A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster." *The Lancet* 395 (2020): 514-523.
 27. Moore, John B, and Carl H June. "Cytokine release syndrome in severe COVID-19." *Sci* 368 (2020): 473-474.
 28. Jones, Veena G, Marcos Mills, Dominique Suarez, and Catherine A Hogan, et al. "COVID-19 and Kawasaki disease: novel virus and novel case." *Hosp Pediat* 10 (2020): 537-540.
 29. Enaud, Raphaël, Renaud Prevel, Eleonora Ciarlo, and Fabien Beauflis, et al. "The gut-lung axis in health and respiratory diseases: a place for inter-organ and inter-kingdom crosstalks." *Front Cellul Infect Microbiol* 10 (2020): 9.
 30. Marsland, Benjamin J, Aurélien Trompette, and Eva S Gollwitzer. "The gut–lung axis in respiratory disease." *Ann Amer Thora Soci* 12 (2015): S150-S156.
 31. Van der Lelie, Daniel, and Safiyh Taghavi. "COVID-19 and the gut microbiome: more than a gut feeling." *Msystems* 5 (2020).
 32. Gurung, Manoj, Zhipeng Li, Hannah You, and Richard Rodrigues, et al. "Role of gut microbiota in type 2 diabetes pathophysiology." *EBioMed* 51 (2020): 102590.
 33. O'Hara, Ann M, and Fergus Shanahan. "The gut flora as a forgotten organ." *EMBO Reports* 7 (2006): 688-693.
 34. Belkaid, Yasmine, and Oliver J Harrison. "Homeostatic immunity and the microbiota." *Immun* 46 (2017): 562-576.
 35. Srinath, BS, Rajesh P Shastry, and Sukesh B Kumar. "Role of gut-lung microbiome crosstalk in COVID-19." *Res Biomed Eng* (2020): 1-11.
 36. Ichinohe, Takeshi, Iris K Pang, Yosuke Kumamoto, and David R Peaper, et al. "Microbiota regulates immune defense against respiratory tract influenza A virus infection." *Proc Nat Acad Sci* 108 (2011): 5354-5359.
 37. Maeda, Naoyoshi, Risa Nakamura, Yoshitaka Hirose, and Shinji Murosaki, et al. "Oral administration of heat-killed *Lactobacillus plantarum* L-137 enhances protection against influenza virus infection by stimulation of type I interferon production in mice." *Internat Immunopharmacol* 9 (2009): 1122-1125.
 38. Wang, Dawei, Bo Hu, Chang Hu, and Fangfang Zhu, et al. "Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus–infected pneumonia in Wuhan, China." *JAMA* 323 (2020): 1061-1069.
 39. Han, Chaoqun, Caihan Duan, Shengyan Zhang, and Brennan Spiegel, et al. "Digestive symptoms in COVID-19 patients with mild disease severity: clinical presentation, stool viral RNA testing, and outcomes." *Amer J Gastroenterol* (2020).
 40. Cheung, Ka Shing, Ivan FN Hung, Pierre PY Chan, and KC Lung, et al. "Gastrointestinal manifestations of SARS-CoV-2 infection and virus load in fecal samples from a Hong Kong cohort: systematic review and meta-analysis." *Gastroenterol* 159 (2020): 81-95.
 41. Donnelly, Christl A, Azra C Ghani, Gabriel M Leung, and Anthony J Hedley, et al. "Epidemiological determinants of spread of causal agent of severe acute respiratory syndrome in Hong Kong." *The Lancet* 361 (2003): 1761-1766.
 42. Assiri, Abdullah, Jaffar A Al-Tawfiq, Abdullah A Al-Rabeeah, and Fahad A Al-Rabiah, et al. "Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study." *The Lancet Infect Dis* 13 (2013): 752-761.
 43. Wölfel, Roman, Victor M Corman, Wolfgang Guggemos, and Michael Seilmaier, et al. "Virological assessment of hospitalized patients with COVID-2019." *Nat* 581 (2020): 465-469.
 44. Xu, Yi, Xufang Li, Bing Zhu, and Huiying Liang, et al. "Characteristics of pediatric SARS-CoV-2 infection and potential evidence for persistent fecal viral shedding." *Nat Med* 26 (2020): 502-505.
 45. Jin, Xi, Jiang-Shan Lian, Jian-Hua Hu, and Jianguo Gao, et al. "Epidemiological, clinical and virological characteristics of 74 cases of coronavirus-infected disease 2019 (COVID-19) with gastrointestinal symptoms." *Gut* 69 (2020): 1002-1009.
 46. Du, Mulong, Guoshuai Cai, Feng Chen, and David C Christiani, et al. "Multiomics evaluation of gastrointestinal and other clinical characteristics of COVID-19." *Gastroenterol* 158 (2020): 2298-2301.
 47. Zheng, Shufa, Jian Fan, Fei Yu, and Baihuan Feng, et al. "Viral load dynamics and disease severity in patients infected with SARS-CoV-2 in Zhejiang province, China, January–March 2020: retrospective cohort study." *BMJ* 369 (2020).
 48. Vandeputte, Doris, Gwen Falony, Sara Vieira-Silva, and Raul Y Tito, et al. "Stool consistency is strongly associated with gut microbiota richness and composition, enterotypes and bacterial growth rates." *Gut* 65 (2016): 57-62.
 49. Gou, Wanglong, Yuanqing Fu, Liang Yue, and Geng-dong Chen, et al. "Gut microbiota may underlie the predisposition of healthy individuals to COVID-19." *MedRxiv* (2020).
 50. Effenberger, Maria, Felix Grabherr, Lisa Mayr, and Julian Schwaerzler, et al. "Faecal calprotectin indicates intestinal inflammation in COVID-19." *Gut* 69 (2020): 1543-1544.
 51. Turnbaugh, Peter J, Ruth E Ley, Michael A Mahowald, and Vincent Magrini, et al. Gordon. "An obesity-associated gut microbiome with increased capacity for energy harvest." *Nat* 444 (2006): 1027-1031.
 52. Emoto, Takuo, Tomoya Yamashita, Naoto Sasaki, and Yushi Hirota, et al. "Analysis of gut microbiota in coronary artery disease patients: a possible link between gut microbiota and coronary artery disease." *J Atheroscler Thromb* (2016).
 53. Yang, Tao, Monica M Santisteban, Verma Rodriguez, and Eric Li, et al. "Gut dysbiosis is linked to hypertension." *Hyperten* 65 (2015): 1331-1340.
 54. Ley, Ruth E, Peter J Turnbaugh, Samuel Klein, and Jeffrey I Gordon. "Human gut microbes associated with obesity." *Nat* 444 (2006): 1022-1023.
 55. Geva-Zatorsky, Naama, Esen Sefik, Lindsay Kua, and Lesley Pasman, et al. "Mining the human gut microbiota for immunomodulatory organisms." *Cell* 168 (2017): 928-943.
 56. Zuo, Tao, Fen Zhang, Grace CY Lui, and Yun Kit Yeoh, et al. "Alterations in gut microbiota of patients with COVID-19 during time of hospitalization." *Gastroenterol* 159 (2020): 944-955.
 57. Hwang, Nakwon, Taekil Eom, Sachin K Gupta, and Seong-Yeop Jeong, et al. "Genes and gut bacteria involved in luminal butyrate reduction caused by diet and loperamide." *Genes* 8 (2017): 350.
 58. Ratajczak, Weronika, Aleksandra Ryl, Arnold Mizerski, and Kinga Walczakiewicz, et al. "Immunomodulatory potential of gut microbiome-derived short-chain fatty acids (SCFAs)." *Acta Biochim Polon* 66 (2019): 1-12.
 59. Zuo, Tao, Qin Liu, Fen Zhang, and Grace Chung-Yan Lui, et al. "Depicting SARS-CoV-2 faecal viral activity in association with gut microbiota composition in patients with COVID-19." *Gut* 70 (2021): 276-284.
 60. Qin, Nan, Beiwen Zheng, Jian Yao, and Lihua Guo, et al. "Influence of H7N9 virus infection and associated treatment on human gut microbiota." *Scient Reports* 5 (2015): 1-12.
 61. Deriu, Elisa, Gayle M Boxx, Xuesong He, and Calvin Pan, et al. "Influenza virus affects intestinal microbiota and secondary salmonella infection in the gut through type I interferons." *PLoS Pathog* 12 (2016): e1005572.
 62. Trompette, Aurélien, Eva S Gollwitzer, Céline Pattaroni, and Isabel C Lopez-Mejia, et al. "Dietary fiber confers protection against flu by shaping Ly6c– patrolling monocyte hematopoiesis and CD8+ T cell metabolism." *Immunity* 48 (2018): 992-1005.
 63. Raftis, Emma J, Margaret I Delday, Philip Cowie, and Seánín M McCluskey, et al. "*Bifidobacterium breve* MRx0004 protects against airway inflammation in a severe asthma model by suppressing both neutrophil and eosinophil lung infiltration." *Scient Reports* 8 (2018): 1-13.
 64. Haase, Stefanie, Aiden Haghikia, Nicola Wilck, and Dominik N Müller, et al. "Impacts

- of microbiome metabolites on immune regulation and autoimmunity." *Immunol* 154 (2018): 230-238.
65. Antunes, Krist Helen, José Luís Fachi, Rosemeire de Paula, and Emanuelle Fraga da Silva, et al. "Microbiota-derived acetate protects against respiratory syncytial virus infection through a GPR43-type 1 interferon response." *Nat Commun* 10 (2019): 1-17.
 66. <https://www.4dpharmapl.com/en/newsroom/press-releases/clinical-update-phase-ii-COVID-19-study>
 67. Bradley, Konrad C, Katja Finsterbusch, Daniel Schnepf, and Stefania Crotta, et al. "Microbiota-driven tonic interferon signals in lung stromal cells protect from influenza virus infection." *Cell Reports* 28 (2019): 245-256.
 68. Corman, Victor M, Jörg U Ganzhorn, Jacques Rakotonandriana, and Yedidya R Ratovonamana, et al. "Adenovirus infection is associated with altered gut microbial communities in a non-human primate." *Scient Reports* 9 (2019): 1-12.
 69. Iuliano, A Danielle, Katherine M Roguski, Howard H Chang, and David J Muscatello, et al. "Estimates of global seasonal influenza-associated respiratory mortality: a modelling study." *The Lancet* 391 (2018): 1285-1300.
 70. Shen, Zijie, Yan Xiao, Lu Kang, and Wentai Ma, et al. "Genomic diversity of severe acute respiratory syndrome–coronavirus 2 in patients with coronavirus disease 2019." *Clin Infect Dis* 71 (2020): 713-720.
 71. Gu, Silan, Yanfei Chen, Zhengjie Wu, and Yunbo Chen, et al. "Alterations of the gut microbiota in patients with coronavirus disease 2019 or H1N1 influenza." *Clin Infect Dis* 71 (2020): 2669-2678.
 72. Nahum, Ari, Gregory Filice, and Ashish Malhotra. "A complicated thread: abdominal actinomycosis in a young woman with Crohn disease." *Case Reports Gastroenter* 11 (2017): 377-381.
 73. Toubiana, Julie, Clément Poirault, Alice Corsia, and Fanny Bajolle, et al. "Kawasaki-like multisystem inflammatory syndrome in children during the COVID-19 pandemic in Paris, France: prospective observational study." *BMJ* 369 (2020).
 74. Esposito, Susanna, Ilaria Polinori, and Donato Rigante. "The gut microbiota-host partnership as a potential driver of Kawasaki syndrome." *Front Pediatr* 7 (2019): 124.
 75. Tao, Wanyin, Guorong Zhang, Xiaofang Wang, and Meng Guo, et al. "Analysis of the intestinal microbiota in COVID-19 patients and its correlation with the inflammatory factor IL-18." *Med Microecol* 5 (2020): 100023.
 76. Cao, Xuetao. "COVID-19: immunopathology and its implications for therapy." *Nat Rev Immunol* 20 (2020): 269-270.
 77. Diao, Bo, Chenhui Wang, Yingjun Tan, and Xiewan Chen, et al. "Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19)." *Front Immunol* 11 (2020): 827.
 78. Round, June L, S Melanie Lee, Jennifer Li, and Gloria Tran, et al. "The Toll-like receptor 2 pathway establishes colonization by a commensal of the human microbiota." *Sci* 332 (2011): 974-977.
 79. Cebula, Anna, Michal Seweryn, Grzegorz A Rempala, and Simarjot Singh Pabla, et al. McIndoe, Timothy L. Denning, Lynn Bry, Piotr Kraj, Pawel Kisielow, and Leszek Ignatowicz. "Thymus-derived regulatory T cells contribute to tolerance to commensal microbiota." *Nat* 497 (2013): 258-262.
 80. Furusawa, Yukihiro, Yuuki Obata, Shinji Fukuda, and Takaho A Endo, et al. "Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells." *Nat* 504 (2013): 446-450.
 81. Smith, Patrick M, Michael R Howitt, Nicolai Panikov, and Monia Michaud, et al. "The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis." *Sci* 341 (2013): 569-573.
 82. Hepworth, Matthew R, Thomas C Fung, Samuel H Masur, and Judith R Kelsen, et al. "Group 3 innate lymphoid cells mediate intestinal selection of commensal bacteria-specific CD4+ T cells." *Sci* 348 (2015): 1031-1035.
 83. Hooper, Lora V, Dan R Littman, and Andrew J Macpherson. "Interactions between the microbiota and the immune system." *Sci* 336 (2012): 1268-1273.
 84. Sencio, Valentin, Adeline Barthélémy, Luciana P Tavares, and Marina G Machado, et al. "Gut dysbiosis during influenza contributes to pulmonary pneumococcal superinfection through altered short-chain fatty acid production." *Cell Reports* 30 (2020): 2934-2947.
 85. Atarashi, Koji, Takeshi Tanoue, Kenshiro Oshima, and Wataru Suda, et al. "T reg induction by a rationally selected mixture of Clostridia strains from the human microbiota." *Nat* 500 (2013): 232-236.
 86. Tanoue, Takeshi, Koji Atarashi, and Kenya Honda. "Development and maintenance of intestinal regulatory T cells." *Nat Rev Immunol* 16 (2016): 295-309.
 87. Steed, Ashley L, George P Christophi, Gerard E Kaiko, and Lulu Sun, et al. "The microbial metabolite desaminotyrosine protects from influenza through type I interferon." *Sci* 357 (2017): 498-502.
 88. Wang, Jian, Fengqi Li, Haiming Wei, and Zhe-Xiong Lian, et al. "Respiratory influenza virus infection induces intestinal immune injury via microbiota-mediated Th17 cell-dependent inflammation." *J Exper Med* 211 (2014): 2397-2410.
 89. Pan, Lei, Mi Mu, Pengcheng Yang, and Yu Sun, et al. "Clinical characteristics of COVID-19 patients with digestive symptoms in Hubei, China: a descriptive, cross-sectional, multicenter study." *The Amer J Gastroenterol* 115 (2020).
 90. Cole-Jeffrey, Colleen T, Meng Liu, and Michael J Katovich, et al. "ACE2 and microbiota: emerging targets for cardiopulmonary disease therapy." *J Cardiovascul Pharmacol* 66 (2015): 540.
 91. Gheblawi, Mahmoud, Kaiming Wang, Anissa Viveiros, and Quynh Nguyen, et al. "Angiotensin-converting enzyme 2: SARS-CoV-2 receptor and regulator of the renin-angiotensin system: celebrating the 20th anniversary of the discovery of ACE2." *Circulat Res* 126 (2020): 1456-1474.
 92. Zhang, Hao, Zijian Kang, Haiyi Gong, and Da Xu, et al. "The digestive system is a potential route of 2019-nCov infection: a bioinformatics analysis based on single-cell transcriptomes." *BioRxiv* (2020).
 93. Kuba, Keiji, Yumiko Imai, Shuan Rao, and Hong Gao, et al. "A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus–induced lung injury." *Nat Med* 11 (2005): 875-879.
 94. Chan, Jasper Fuk-Woo, Kin-Hang Kok, Zheng Zhu, and Hin Chu, et al. "Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan." *Emerg Micro Infect* 9 (2020): 221-236.
 95. Zhou, Peng, Xing-Lou Yang, Xian-Guang Wang, and Ben Hu, et al. "A pneumonia outbreak associated with a new coronavirus of probable bat origin." *Nat* 579 (2020): 270-273.
 96. He, Yu, Jianhui Wang, Fang Li, and Yuan Shi. "Main clinical features of COVID-19 and potential prognostic and therapeutic value of the microbiota in SARS-CoV-2 infections." *Front Microbiol* (2020): 1302.
 97. Hashimoto, Tatsuo, Thomas Perlot, Ateequr Rehman, and Jean Trichereau, et al. "ACE2 links amino acid malnutrition to microbial ecology and intestinal inflammation." *Nat* 487 (2012): 477-481.
 98. Lu, Chen Chen, Kun Ling Ma, and Xiong Zhong Ruan, et al. "Intestinal dysbiosis activates renal renin-angiotensin system contributing to incipient diabetic nephropathy." *Internat J Med Sci* 15 (2018): 816.
 99. Wang, Jun, Shanmeizi Zhao, Ming Liu, and Zhiyao Zhao, et al. "ACE2 expression by colonic epithelial cells is associated with viral infection, immunity and energy metabolism." *MedRxiv* (2020).
 100. Zhou, Jie, Cun Li, Guangyu Zhao, and Hin Chu, et al. "Human intestinal tract serves as an alternative infection route for Middle East respiratory syndrome coronavirus." *Sci Adv* 3 (2017): 4966.
 101. Xiao, Fei, Meiwang Tang, Xiaobin Zheng, and Ye Liu, et al. "Evidence for gastrointestinal infection of SARS-CoV-2." *Gastroenterol* 158 (2020): 1831-1833.
 102. Gao, Jing, Kang Xu, Hongnan Liu, and Gang Liu, et al. "Impact of the gut microbiota on intestinal immunity mediated by tryptophan metabolism." *Front Cellul Infect Microbiol* 8 (2018): 13.
 103. Zhao, Ye, Feidi Chen, Wei Wu, and Mingming Sun, et al. "GPR43 mediates microbiota metabolite SCFA regulation of antimicrobial peptide expression in intestinal epithelial cells via activation of mTOR and STAT3." *Mucosal Immunol* 11 (2018): 752-762.
 104. Liévin-Le Moal, Vanessa, and Alain L Servin. "The front line of enteric host defense against unwelcome intrusion of harmful microorganisms: mucins, antimicrobial peptides, and microbiota." *Clin Microbiol Rev* 19 (2006): 315-337.
 105. Kim, Seungbum, Katya Rigatto, Marcelo B Gazzana, and Marli M Knorst, et al. "Altered gut microbiome profile in patients with pulmonary arterial hypertension." *Hyperten* 75 (2020): 1063-1071.

106. Hemnes, Anna R, Anandharajan Rathinasabapathy, Eric A Austin, and Evan L Brittain, et al. "A potential therapeutic role for angiotensin-converting enzyme 2 in human pulmonary arterial hypertension." *Europ Respirat J* 51 (2018).
107. Dickson, Robert P, Benjamin H Singer, Michael W Newstead, and Nicole R Falkowski, et al. "Enrichment of the lung microbiome with gut bacteria in sepsis and the acute respiratory distress syndrome." *Nat Microbiol* 1 (2016): 1-9.
108. Cho, Eun-Jung, Heungsup Sung, Sook-Ja Park, and Mi-Na Kim, et al. "Rothia mucilaginosa pneumonia diagnosed by quantitative cultures and intracellular organisms of bronchoalveolar lavage in a lymphoma patient." *Ann Laborat Med* 33 (2013): 145.
109. Lu, Hai-feng, Ang Li, Ting Zhang, and Zhi-gang Ren, et al. "Disordered oropharyngeal microbial communities in H7N9 patients with or without secondary bacterial lung infection." *Emerg Microb Infect* 6 (2017): 1-11.
110. Khan M. "Rapid response: gut microbiome and COVID-19 mRNA vaccine." *BMJ* 372 (2021): n149.

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