



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

## Evaluating alternative stem cell hypotheses for adult corneal epithelial maintenance

**Citation for published version:**

West, JD, Dora, N & Collinson, JM 2015, 'Evaluating alternative stem cell hypotheses for adult corneal epithelial maintenance', *World Journal of Stem Cells*, vol. 7, no. 2, pp. 281-299.  
<https://doi.org/10.4252/wjsc.v7.i2.281>

**Digital Object Identifier (DOI):**

[10.4252/wjsc.v7.i2.281](https://doi.org/10.4252/wjsc.v7.i2.281)

**Link:**

[Link to publication record in Edinburgh Research Explorer](#)

**Document Version:**

Peer reviewed version

**Published In:**

World Journal of Stem Cells

**Publisher Rights Statement:**

Copyright ©1995-2014 Baishideng Publishing Group Inc. All rights reserved. Articles published by this open-access journal are distributed under the terms of the Creative Commons Attribution-Noncommercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

**General rights**

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy**

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact [openaccess@ed.ac.uk](mailto:openaccess@ed.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.



Name of journal: World Journal of Stem Cells

ESPS Manuscript NO: 12919

## **Evaluating alternative stem cell hypotheses for adult corneal epithelial maintenance**

John D West, Natalie J Dorà, J Martin Collinson

John D. West and Natalie J. Dorà, Genes and Development Group, Centre for Integrative Physiology, College of Medicine and Veterinary Medicine, University of Edinburgh, Hugh Robson Building, George Square, Edinburgh, EH8 9XD, United Kingdom

J Martin Collinson, Institute of Medical Sciences, School of Medical Sciences, College of Life Sciences and Medicine, Foresterhill, University of Aberdeen, Aberdeen, AB25 2ZD, United Kingdom

**Running title:** Corneal epithelial maintenance

**Key words:** eye; cornea; corneal epithelium; limbal epithelium; stem cell; lineage tracing

**Author contributions:** West JD drafted and designed the manuscript, Dorà NJ performed research, West JD and Dorà NJ prepared the figures, West JD, Dorà NJ and Collinson JM wrote, critically revised and approved the manuscript

**Correspondence to:** John D. West, University Reader, Genes and Development Group, Centre for Integrative Physiology, College of Medicine and Veterinary Medicine, University of Edinburgh, Hugh Robson Building, George Square, Edinburgh, EH8 9XD, United Kingdom. john.west@ed.ac.uk

Telephone: +44-131-6503112

Fax: +44-131-6511706

**Supported by** Grants from the Wellcome Trust (No. 088876/Z/09/Z) and the UK Biotechnology and Biological Sciences Research Council (No. BB/J015172/1 and No. BB/J015237/1).

## **ABSTRACT**

In this review we evaluate evidence for three different hypotheses that explain how the corneal epithelium is maintained. The limbal epithelial stem cell (LESC) hypothesis is most widely accepted. This proposes that stem cells in the basal layer of the limbal epithelium, at the periphery of the cornea, maintain themselves and also produce transient (or transit) amplifying cells (TACs). TACs then move centripetally to the centre of the cornea in the basal layer of the corneal epithelium and also replenish cells in the overlying suprabasal layers. The LESCs maintain the corneal epithelium during normal homeostasis and become more active to repair significant wounds. Second, the corneal epithelial stem cell (CESC) hypothesis postulates that, during normal homeostasis, stem cells distributed throughout the basal corneal epithelium, maintain the tissue. According to this hypothesis, LESCs are present in the limbus but are only active during wound healing. We also consider a third possibility, that the corneal epithelium is maintained during normal homeostasis by proliferation of basal corneal epithelial cells without any input from stem cells. After reviewing the published evidence, we conclude that the LESC and CESC hypotheses are consistent with more of the evidence than the third hypothesis, so we do not consider this further. The LESC and CESC hypotheses each have difficulty accounting for one main type of evidence so we evaluate the two key lines of evidence that discriminate between them. Finally, we discuss how lineage-tracing experiments have begun to resolve the debate in favour of the LESC hypothesis. Nevertheless, it also seems likely that some basal corneal epithelial cells can act as long-term progenitors if limbal stem cell function is compromised. Thus, this aspect of the CESC hypothesis may have a lasting impact on our understanding of corneal epithelial maintenance, even if it is eventually shown that stem cells are restricted to the limbus as proposed by the LESC hypothesis.

## **CORE TIP**

This review article evaluates the evidence for different hypotheses that have been proposed to explain how the corneal epithelium is maintained. It identifies core observations in favour of the conventional limbal epithelial stem cell (LESC) hypothesis and an alternative corneal epithelial stem cell (CESC) hypothesis and describes how lineage-tracing experiments are helping to reconcile the two sets of conflicting evidence in

favour of the LESC hypothesis.

## **INTRODUCTION**

It is widely accepted that adult corneal epithelium is maintained by stem cells located in a region called the limbus, at the corneal periphery. However, this limbal epithelial stem cell (LESC) hypothesis has been challenged by an alternative corneal epithelial stem cell (CESC) hypothesis, based on experimental studies with mice [1]. This accepts that LESCs exist but proposes that they only contribute to corneal epithelial repair in response to wounding and that, during normal homeostasis, the corneal epithelium is maintained solely by stem cells scattered throughout the corneal epithelium itself. It has also been proposed that, in the absence of a wound, the corneal epithelium is maintained entirely by proliferation of its own basal cells without any involvement of stem cells [2]. The main purpose of this review is to compare the evidence for the alternative LESC and CESC hypotheses in order to identify where there is common ground and where differences need further experimental investigation. However, we also consider whether the experimental evidence is consistent with the possibility that the corneal epithelium is maintained without stem cells.

## **THE CORNEAL AND LIMBAL EPITHELIA**

The cornea is the specialised, avascular, transparent, dome-shaped region of the anterior ocular surface, which refracts light through the pupil to the lens and provides a protective, impermeable barrier. It consists of three cellular layers: (i) the inner corneal endothelium, which, despite its name, is a type of epithelium; (ii) the middle corneal stroma, comprising specialised fibroblasts, called keratocytes, embedded in a collagen and proteoglycan matrix and (iii) the outer, non-keratinised, stratified squamous epithelium, comprising 5-6 layers of keratinocytes, which is kept moist by the tear film. The corneal epithelium is a very dynamic tissue. Differentiated cells are continuously shed from the outer layer and replaced by cells produced in the proliferative basal layer. According to the conventional stem cell paradigm these proliferative basal cells are considered to be transient (or transit) amplifying cells (TACs) and are replenished by stem cells. The early generation TACs are sometimes referred to as progenitor cells.

The limbus is a narrow transition zone, which encircles the cornea (Figs. 1 & 2). The stroma and epithelial layers of the cornea extend into the limbus where they become the limbal stroma and limbal epithelium. However, the corneal endothelium does not extend

into the limbus and is replaced by the drainage channels of the trabecular meshwork. On the other side of the limbus, the stroma merges with the sclera, which forms most of the ocular surface (the white part of the human eye) and the limbal epithelium becomes the conjunctiva. The conjunctiva is an epithelium, which covers the anterior sclera, folds back to form the conjunctival sac and lines the inner surface of the eyelids (Fig. 1). Thus, the conjunctiva attaches the eyeball to the eyelids and orbit and permits some rotation of the eyeball in the orbit. Unlike the transparent cornea, both the limbus and the conjunctiva are vascularised.

The human limbus contains radial fibrovascular ridges, called the palisades of Vogt, which project upwards from the stroma deep into the epithelium (Fig. 2) but many species, including mice, do not have limbal palisade structures. Another anatomical difference between these species is that, in the mouse, the corneal epithelium is thickest in the centre of the cornea and has fewer cell layers in the peripheral cornea and limbus whereas, in humans, the limbal epithelium is thicker (about 8-10 cell layers) than the corneal epithelium (5-6 layers) [3].

It is widely accepted that some basal limbal epithelial cells are stem cells [4, 5] and that the limbal stroma, vasculature and other associated cell types provides a suitable stem cell niche microenvironment, which is required to maintain the limbal epithelial stem cells (LESCs) in a relatively undifferentiated state [6, 7]. The limbal palisades (Fig. 2) increase the area of interface between the limbal epithelium and stroma, so increasing the size of the region that is likely to harbour the LESC niches. Furthermore, it has been suggested that LESC niches may be particularly enriched in two types of epithelial crypts associated with the palisades. One type of crypt is formed by the regions of limbal epithelium between the upward-projecting stroma of the limbal palisades and these have been named 'limbal crypts' (LCs) [8, 9]. The other type of crypt (named 'limbal epithelial crypts'; LECs) are more sparsely distributed (only 6-7 per eye) and are formed from epithelial projections from the periphery of the limbal palisades, which extend either radially from the limbus into the conjunctival stroma or circumferentially within the limbus (perpendicular to the palisades) [10, 11]. However, many species do not have limbal palisades and associated crypts, which could, therefore, be considered to be species-specific adaptations, possibly related to eye size. Thus, if the limbus provides a niche microenvironment, presumably it is either not dependent on these structures or differs among species. Maintenance of the

niche microenvironment is more likely to depend on the presence of the vasculature and other cell types that are present in the limbus in all species.

## **ALTERNATIVE HYPOTHESES OF CORNEAL EPITHELIAL MAINTENANCE**

We consider two hypotheses, which propose alternative ways that stem cells may maintain the corneal epithelium, and a third hypothesis, which does not include stem cells. For the present purposes, we define a stem cell as an undifferentiated cell with high proliferative potential that is capable of renewing itself and also producing one or more differentiated cell types with lower proliferative potential. While most adult stem cells are multipotent, generating multiple cell types, stem cells that maintain the corneal epithelium are generally presumed to be unipotent, only producing the corneal epithelial cells. Although one report shows that they may also produce the goblet cells that enter the corneal epithelium in response to large wounds <sup>[12]</sup>, we have not considered this possible additional role of the stem cells in this review.

### **Limbal epithelial stem cell (LESC) hypothesis**

According to the conventional LESc hypothesis (Figs. 3A,D), LESCs act as a source of new basal corneal epithelial cells (the TACs) during normal corneal epithelial homeostasis and become more active during episodes of significant wound healing <sup>[13]</sup>, although small wounds may be healed without upregulating stem cells. In this scheme LESCs remain in the limbus where they maintain themselves and also generate the first generation of TACs. Some of these early TACs move to the overlying, non-mitotic suprabasal epithelial layers, and become terminally differentiated. Other early generation TACs continue to divide and move centripetally in the basal layer to maintain the corneal epithelium. Once cells leave the basal layer they differentiate and move rapidly through the suprabasal layers to the superficial layer from where they are shed. It seems that both daughter cells of a dividing basal cell usually share the same fate <sup>[14]</sup> so that they either both remain in the proliferative basal layer or both move suprabasally. It is not known what determines whether cells leave the basal layer. For example, it could be a combination of declining suprabasal cell numbers, caused by cell loss, and overcrowding in the basal layer, caused by cell proliferation, as described for the *Drosophila notum* <sup>[15]</sup>.

### **Corneal epithelial stem cell (CESC) hypothesis**

The CESC hypothesis accepts that there are stem cells in the limbus but proposes that these are only activated for repairing wounds and that during normal homeostasis the corneal epithelium is maintained by stem cells scattered throughout the corneal epithelium itself [1]. This hypothesis is based largely on surgical transplantation experiments in mice. These experiments showed that labelled limbal epithelial tissue, transplanted to the limbus of immunologically compromised mice, only produced labelled clones in the cornea if the host corneal epithelium was subsequently removed. The authors reasoned that if LESC were active during normal homeostasis, as the LESC proposes, the donor limbal tissue should colonise the cornea without being stimulated to do so by wounding. However, others have pointed out that the CESC hypothesis is not consistent with some of the earlier experimental evidence [16-18].

### **Germinative basal layer (GBL) hypothesis**

A third possibility harks back to earlier explanations before the importance of tissue stem cells was recognised. Like the CESC hypothesis, this proposes that the corneal epithelium is normally maintained entirely from cells in the basal layer of the corneal epithelium but unlike the other two hypotheses it proposes there are no stem cells in either the limbal or corneal epithelia. Haddad and Faria-e-Sousa [2] referred to the basal layer as the 'germinative basal layer' and proposed this alternative mechanism for corneal epithelial maintenance to explain the results of their label-retaining cell experiment with rabbits. These results are inconsistent with other label-retaining cell experiments, as discussed below. However, we have considered this hypothesis because there is evidence that some other adult tissues are maintained during normal homeostasis by proliferation of more differentiated cell types. In such cases, stem cells are either absent or only active during wound repair. For example, this type of tissue maintenance has been proposed for pancreas  $\beta$ -cells [19], epidermis [20], lung [21] and liver hepatocytes [22, 23].

## **EXPERIMENTAL EVIDENCE AND EVALUATION OF ALTERNATIVE HYPOTHESES**

The three alternative hypotheses are discussed below with respect to the available experimental evidence and evaluations are summarised in Table 1.



## Cells with high proliferative potential

One of the hallmarks of stem cells is that they have a greater proliferative potential than TACs and this can be identified using *in vitro* colony-forming assays with cultured cells. The proliferation characteristics of cultures of explanted epithelial cells can be investigated in culture and clones derived from single cells can be classified as holoclones, meroclones and paraclones. These are thought to represent *in vitro* descendants of stem cells, TACs and differentiated cells, respectively [24]. (On indicator dishes in culture, holoclones form large, smooth-edged, fast-growing colonies with large numbers of small tightly packed cells. Meroclones form smaller colonies that are irregular in outline and include a mixture of small tightly packed cells and larger more loosely packed cells, which are predominantly at the edge). Paraclones form small, diffuse colonies and most cells are large, flattened and loosely packed.) Clonal analysis of cells from the human ocular surface epithelia identified holoclone-producing cells in the limbal epithelium but not the corneal epithelium from a 54 year old individual [25], suggesting that stem cells are present in the limbal epithelium.

However, there is also evidence that some cells of the central cornea are self-sustaining and have high proliferative potential. Majo et al showed that cultured corneal epithelial cells of many species produced colonies of cells *in vitro* although there were significant species differences [1]. Pig corneal epithelial cells grew particularly well and clonal analysis of cultured pig cells identified holoclone-producing cells in the central corneal epithelium as well as the limbus. This suggests that stem cells are present in the central corneal epithelium of pigs as well as the limbus but this result cannot be evaluated fully as the age of pigs was not given and fetal cells with stem cell characteristics may persist in younger individuals [26-28].

The production of clonogenic spheres of cells in culture has also been associated with the presence of stem cells and these have been isolated from the human limbus and central cornea, although isolation is most efficient from the limbus and from younger individuals [29]. These culture experiments imply that both the limbal epithelium and the central corneal epithelium have cells that are able to behave like stem cells in clonogenic assays *in vitro* which argues against the LESC and GBL hypotheses. However, if culture conditions unmasked proliferative potential of corneal epithelial cells, which is not expressed during normal homeostasis *in vivo*, this result would be compatible with all

three hypotheses.

Despite their slow proliferation *in vivo* (see next section and reference [30]), human limbal epithelial cells grow well in culture and have a higher mitotic rate than corneal epithelial cells [31]. Furthermore, clinical observations indicate that human limbal tissue is superior to central corneal tissue for treating patients with severely wounded corneal epithelia, which is likely to reflect a greater proliferative potential. The corneal epithelium can be restored using grafts of human limbal epithelial tissue [32, 33] or cells cultured from explanted limbal tissue [34-38]. Although the limbus is the preferred source of cells for clinical therapeutic use, this does not help determine whether LESC are active during normal tissue homeostasis (LESC hypothesis) or only during wound healing (CESC hypothesis).

There is also evidence that the central cornea of several species contains highly proliferative cells. For example, rabbit central cornea is able to survive for months after the limbus is removed or separated from the cornea [39, 40], although corneal integrity slowly degenerates and it does not heal properly after corneal wounding. Similarly, the mouse corneal epithelium was able to sustain itself for four months after the limbus was cauterised to destroy the limbal epithelium [1]. It has also been reported that some patients with symptoms of total LESC deficiency retain central islands of normal corneal epithelium for several years [41] and in one case this appeared to be sufficient to restore the corneal epithelium [42]. These studies show that the central cornea can maintain itself to some extent when the limbus is eliminated or disconnected. This implies that the central corneal epithelium has cells that are able to act as progenitors, if LESC are unable to maintain the corneal epithelium. However, again this does not show whether these cells act as progenitors during normal homeostasis so it does not provide conclusive evidence against the LESC hypothesis.

### **Cell division characteristics and identification of slow-cycling label-retaining cells**

Stem cell populations maintain themselves and produce more differentiated cells throughout the lifetime of the organism. This is sometimes interpreted as requiring stem cells to divide asymmetrically (producing one stem cell and one TAC) but this does not mean that each division of every stem cell has to be asymmetric as long as the population average achieves this. There have been few attempts to identify asymmetrically dividing

cells in the ocular surface [43] and results are insufficient to discriminate among the three hypotheses.

A more widely studied characteristic of many stem cells is that they are relatively quiescent so divide infrequently. A slow cell division rate is not an obligatory phenotype of stem cells but it has been used to try to locate the stem cells that maintain the corneal epithelium. Slow-cycling cells (including putative stem cells) are usually identified as 'label-retaining cells'. These are cells that retain a DNA-label such as BrdU or <sup>3</sup>H-TdR, or a chromatin label such as GFP-tagged histone-2B, after prolonged labelling and a chase period to dilute the label from more rapidly dividing cells. The prolonged period of labelling is to label as many cells as possible including relatively quiescent stem cells that divide infrequently. The chase period is calibrated to dilute label from most cells in the tissue but not any slow-cycling cells (which include putative quiescent stem cells). This method is useful for identifying the location of putative stem cells but is not specific and will also identify other slow-cycling cell types and cells that divide during the labelling period and then stop dividing when they terminally differentiate.

The ocular surface is a suitable tissue for label-retaining cell experiments because the corneal epithelial TACs divide quite frequently so will readily dilute the label. (The average mitotic rate has been estimated for rats as 14.5% per day for the whole corneal epithelium [44] and this equates to approximately 37% for just the mitotic basal layer, based on the relative basal and suprabasal cell numbers in mouse corneas [45].) Similarly, BrdU experiments indicate that almost 50% of basal corneal epithelial cells are in S-phase of the cell cycle, during a 24-hour labelling period [46]. The effectiveness of the chase period is also helped by the constant loss of cells from the superficial layer, as stem cells will not be lost in this way. It has been estimated that once cells leave the basal corneal epithelial layer the time to cell loss (turnover time) is only about 7 days (range 3<sup>1</sup>/<sub>2</sub> to 14 days) for mice, rats and humans [45, 47-50] but a longer turnover time of between 14 and 21 days has been estimated for rabbits [51].

Most investigations have identified label-retaining cells in the basal limbal epithelium but not in the corneal epithelium either after wounding [5, 13] or during normal homeostasis in mice [13, 45, 52-54], rats [55, 56] and rabbits [57]. Two caveats about the exclusive location of label-retaining cells to the limbus in these experiments should be mentioned. (i) Species differences in cell cycle kinetics and technical differences between studies may

affect the number of cells that remain labelled so the chase period needs to be optimised for each species. For example, in these studies, chase periods for treatments without wounding varied from 4 to 11 weeks. (ii) Using a relatively short chase period of 4 weeks, Chen et al [56] showed that approximately 20% of the label-retaining cells were slow-cycling Langerhans cells rather than putative slow-cycling stem cells. (These Langerhans cells also shared two other characteristics of putative stem cells, discussed below, as they were positive for the marker ABCG2 and had a high nucleus to cytoplasm ratio.) Nevertheless, the results of all these studies consistently identified label-retaining cells in the basal limbal epithelium but not in the basal corneal epithelium and it is likely that most of these will be stem cells. Thus, these studies favour the LESC hypothesis, unless there is an additional stem cell population in the corneal epithelium, which is not slow cycling. They also argue against the GBL hypothesis unless none of the limbal label-retaining cells are stem cells.

A completely different result was found for one study with rabbits [2], which prompted the authors to conclude that the corneal epithelium is not maintained by LSCs but by virtually all the cells of the basal corneal epithelium (referred to here as the GBL hypothesis). Rabbits were given 3 intravitreal injections of  $^3\text{H-TdR}$  at intervals of 4 days. After a 41-day chase (49 days after the first injection) the labelling index was higher in the corneal epithelium (17.8%) than the limbal epithelium (3.8%) [2]. However, the high labelling index suggests that many of the labelled cells were TACs, rather than slow-cycling stem cells, and the chase period was insufficient to detect label-retaining cells. Paradoxically, the chase period was comparable to that used in an earlier experiment, which identified label-retaining cells in the limbus but not corneal epithelium of rabbits [57]. In this experiment, BrdU was infused from an osmotic mini-pump for 14 days, the pump was removed at 17 days, and the rabbits were left for a further 38-days chase period (i.e. until 55 days after the beginning of labelling). In the  $^3\text{H-TdR}$  study [2], autoradiography was used to detect label in high quality semi-thin sections and it is possible that this is more sensitive than the BrdU immunofluorescence used earlier [57]. If so, more cell divisions and a longer chase period would be required to dilute the  $^3\text{H-TdR}$  below detectable levels in the majority of cells in order to identify any label-retaining cells. It would, therefore, be worth repeating the  $^3\text{H-TdR}$  experiment with a longer chase period before drawing conclusions that contradict the other label-retaining cell studies.

## Movement of corneal epithelial cells

Early experimental evidence showed that cells moved from the limbus to the cornea to repair a corneal wound in guinea pigs [58]. The observation that donor corneal epithelial cells, transplanted to the centre of rabbit corneas, were replaced by host cells more quickly at the periphery of the transplant also suggested that new host cells were moving centripetally from the periphery of the cornea to replace the older donor cells [59]. Other indirect evidence that cells move centripetally from the limbus during homeostasis of unwounded corneas is reviewed elsewhere [60]. More importantly, direct observations of radial epithelial movement during normal corneal homeostasis have consistently demonstrated that cells move centripetally from the periphery to the centre of the cornea. This supports the LESC hypothesis, which proposes that LSCs remain in their limbal niche but TACs move centripetally to maintain the corneal epithelium.

These experiments also provided estimates of the rate of centripetal movement of corneal epithelial cells for the unwounded cornea. This was estimated to be 28  $\mu\text{m}/\text{day}$  from observations of one human subject over 24 hours using *in vivo* confocal microscopy [61]. For mouse corneas, the rate of centripetal movement has been estimated as 11-26  $\mu\text{m}/\text{day}$  using three different approaches involving direct observations of labelled cells. Corneal epithelial cells labelled with India ink moved 17  $\mu\text{m}/\text{day}$  over 7 days [62], patches of brightly fluorescent cells moved 26  $\mu\text{m}/\text{day}$  over 7 weeks in mosaic GFP transgenic mice [63] and fluorescent clones of cells extended 11  $\mu\text{m}/\text{day}$  over 12 weeks in K14-CreER<sup>T2</sup>;R26R-confetti transgenic mice (from 9 to 21 weeks after tamoxifen-activation of the reporter transgene) [64]. Furthermore, the evidence from the mosaic GFP transgenic mice [63] and tamoxifen-activated reporter transgenic mice [64] demonstrated that the same clonal lineage of cells moved across the full radius from the periphery to the centre. This is in contrast to cells in the conjunctiva, which do not move significantly at all [62, 65].

When Majo et al [1] proposed the CESC cell hypothesis they also proposed that the corneal and conjunctival epithelia continuously expand towards the limbus, which they described as a zone of equilibrium, so any movement in the corneal epithelium was predicted to be centrifugal. This is inconsistent both with the absence of movement in the conjunctiva [62, 65] and the convincing, direct evidence that movement of corneal epithelial cells is centripetal not centrifugal [62-64]. The evidence for centripetal cell movement in the corneal epithelium is inconsistent with the CESC hypothesis as originally proposed [1] but

there is no need to link the stem cell location and movement aspects of the original CESC hypothesis. In principle, it would be possible for the corneal epithelium to be maintained by stem cells, scattered throughout the tissue, without invoking centrifugal movement. It is likely that TACs produced by CESC would only move radially because evidence from various mosaics and chimaeras implies that lateral movement is constrained (discussed in the next section). In theory, radial movement of TACs could be either centripetal or centrifugal but, as noted above, evidence for centripetal movement is compelling.

The causes of centripetal movement are not known and suggestions include (i) population pressure from the periphery due to production of new TACs by LESC [66-68]; (ii) preferential loss of epithelial cells from the central cornea [30, 60]; (iii) differential stiffness of cornea and limbus [69] (iv) chemotaxis [62]; (v) stimulation by corneal nerves [70] and (vi) response to endogenous electric currents [71].

If the LESC hypothesis was incorrect, centripetal movement could still be explained by a mechanism other than population pressure from the limbus. However, if LESC were absent or only active during wound healing, a peripheral source of cells would be required to replace peripheral cells that move centripetally, during normal homeostasis. This might be provided by limbal TACs for the GBL hypothesis or CESC in or near the limbus for the CESC hypothesis. Thus, both the CESC and GBL hypotheses could account for centripetal movement of cells in separately maintained regions on the same radius. However, evidence from transgenic mice shows that a single clone of cells moves across the full radius [63, 64], implying that there is a single source of cells in the limbus or peripheral cornea, rather than multiple sources throughout the cornea. This is more difficult for the CESC and GBL hypotheses to explain unless it is argued that not all radial regions are maintained by a single CESC or progenitor TAC. Another problem for the CESC hypothesis is that the CESC would tend to move centripetally with the TACs, and so accumulate in the centre, unless the CESC were somehow stabilised in unidentified niches and the TACs could move past them. Overall, centripetal movement strongly favours the LESC hypothesis and it is difficult to reconcile this with the other two hypotheses without *ad hoc* assumptions.

### **Change in mosaic patterns after birth**

In addition to direct studies of cell movement in real time, changes in patterns in several

types of mosaic mice have provided additional evidence that cells emerge from the limbus at the periphery of the corneal epithelium and continue to move centripetally across the cornea. Mosaic patterns in adult mouse and rat chimaeras and mouse X-inactivation mosaics (*XLacZ* mosaics) are arranged as radial stripes in the corneal epithelium [27, 72, 73], which is consistent with either centripetal or centrifugal movement, without significant lateral dispersion. Similar radial stripes, have been observed with various endogenous markers in human corneas, including traces of pigment [58] and various opacities, cell inclusions or drug-induced lipidosis associated with vortex keratopathy (cornea verticillata) or hurricane keratopathy [60, 66, 74, 75]. In many cases the stripes form a spiral-pattern in the centre, which fits well with the more direct evidence for centripetal movement, discussed above, because centripetal movement of labelled cells transplanted to the rabbit limbus sometimes formed a similar spiral [76].

Before about 5 weeks of age, the pattern in X-inactivation mosaics is completely different from the adult radial stripes and the  $\beta$ -gal-positive and  $\beta$ -gal-negative cell populations initially form randomly orientated patches [27, 72]. Groups of  $\beta$ -gal-positive and  $\beta$ -gal-negative cells emerge from the periphery by about 5 weeks and extend as radial stripes across the cornea. The simplest interpretation is that the formation of stripes coincides with the onset of activation of stem cells in the limbus that generate new cells, which replace those produced during development [27]. This is supported by similar observations with mosaic transgenic mice [77, 78] and lineage tracing with a GFP-tagged lentiviral marker [79], as illustrated in Fig. 4, and is consistent with the LESC hypothesis but not with the CESC or GBL hypotheses.

One problem with these mosaic systems is that similar proportions of labelled and unlabelled cells were present so many of the radial stripes may comprise more than one adjacent clone that are similarly marked. Observations on *KRT5-LacZ*<sup>+/+</sup> transgenic mice showed that they had rare  $\beta$ -gal-positive stripes in a predominantly  $\beta$ -gal-negative corneal epithelium, so largely avoiding the problem of multiple adjacent clones [17]. The distribution of  $\beta$ -gal-positive stripes was not consistent with predictions of centrifugal extension of clones of labelled cells from  $\beta$ -gal-positive CESC distributed randomly in the corneal epithelium and the simplest interpretation is that the stripes represent clonal lineages derived from LESC located in the limbus. However, analysis of striped patterns

in *KRT5-LacZ*<sup>+/-</sup> corneas is not unequivocal and similar analyses with inducible lineage markers are required, as discussed below.

### **Transplantation experiments**

Bradshaw et al [76] labelled rabbit limbal tissue *ex-vivo* and transplanted it back to the limbus of the donor rabbits after first debriding the corneal epithelium across the full diameter. The labelled cells quickly colonised the corneal epithelium but, as the corneal epithelium was completely removed, this is equivalent to wound healing rather than normal corneal homeostasis. Majo et al [1] transplanted either  $\beta$ -gal-positive limbal or central corneal tissue from transgenic mice into the limbus of  $\beta$ -gal-negative, immunocompromised mice and both sources of tissue produced similar results. Consistent with the earlier experiment with rabbits [76], labelled clones of donor cells moved centripetally into the corneal epithelium if the host corneal epithelium was removed but it failed to contribute to the corneal epithelium if the host cornea was left intact. Thus, although the transplanted limbal tissue contributed to corneal repair, it did not contribute to steady state corneal maintenance during normal tissue homeostasis, as predicted by the LESC hypothesis. This was a key result, which prompted Majo et al to propose the CESC hypothesis [1].

### **Circumstantial evidence**

In addition to the specific investigations discussed so far, there are two circumstantial observations that favour the limbus as a site for stem cells. First, tumours of the ocular surface commonly involve the limbus [80] and for other systems it has been suggested that tumour cells may preferentially arise from stem cells [81, 82]. This provides only weak, circumstantial evidence in favour of the LESC hypothesis.

Second, it is generally agreed that stem cells need a specialised niche environment to maintain the stem cell phenotype and this is likely to involve interactions with several cell types [6, 83]. For example, signalling from the microvasculature plays an important role in the mouse neural stem cell niche [84]. Undeniably, the limbus provides a more diverse population of cell types than cornea and this is enriched further by its blood supply and for this reason it seems arguably a more likely location for a stem cell niche than the cornea. As already mentioned, an additional issue is that a stem cell niche in the basal



corneal epithelium might be unstable because of the continuous centripetal movement of TACs. These considerations also make it more likely that stem cell niches would be located preferentially in the limbus rather than the cornea.

Although some tissues are maintained during normal homeostasis by stem cells in the main body of the tissue, the limbus is not the only putative stem cell niche with a more peripheral location. For example, there are two types of stem cells that maintain the epithelium that lines the intestinal crypts and villi: crypt base columnar cells and position +4 reserve stem cells. These are both located near the base of the intestinal crypts, from where they produce TACs, which move up the crypt and generate the different functional cell types of the villus epithelium [85]. Maintenance of the corneal epithelium by stem cells located in the limbal epithelium, as proposed by the LESC hypothesis, is essentially analogous to the way the intestinal epithelium is maintained. The circumstantial evidence that the limbus is a likely location for a stem cell niche supports the LESC hypothesis. However, it does not provide strong evidence against the CESC hypothesis, which accepts that LSCs exist, or the GBL hypothesis, which predicts there are no stem cells and so no niches.

### **Stem cell markers and phenotype**

The Holy Grail of stem cell research is to find a phenotype or cell marker that allows the stem cells to be unequivocally distinguished from all neighbouring cells, including early generation TACs, and isolated for further study. This has not yet proved possible for the putative stem cell population(s) that maintain the corneal epithelium. Early evidence that the basal limbal epithelium contained stem cells was produced by an immunohistochemical study of keratin 3 (K3), which is considered to be a corneal differentiation marker [4]. K3 is expressed in the basal and suprabasal layers of the rabbit corneal epithelium but only the suprabasal layers of the limbal epithelium, leading to the conclusion the basal limbal epithelium was less differentiated than the other epithelial layers. The mouse has no K3 [86] but K12, which normally pairs with K3, is present and expression is restricted to the cornea [87], as shown in Fig. 3B. Several authors have also noted that cell morphology of cells in the basal limbal epithelium was more characteristic of stem cells (smaller, euchromatin-rich, high nucleus to cytoplasm ratio) than the corneal epithelium [3, 88] but, as already noted, Langerhans cells in the limbus also share this

phenotype [56]. These observations are consistent with the hypothesis that the limbus contains stem cells but no more than that.

The discovery of the K3 difference between basal limbal and corneal epithelia, as a whole, was followed by a quest for a specific cell marker to identify the LESC within the limbal epithelium. Many candidate markers have been proposed based on differential expression studies (reviewed in reference [18]) or conventional immunostaining (Table 2) but no definitive marker has been found, that is known to be expressed in putative stem cells in the limbus but not in neighbouring early generation TACs.

Some of the markers expressed in the limbal but not the corneal epithelium have been identified as putative stem cell markers in other tissues. ATP-binding cassette transporters (ABC transporters) are a family of transmembrane proteins whose functions include the transport of (potentially harmful) metabolic products out of the cells [89]. Conceptually, they may form a component of the molecular mechanisms by which long-lived stem cells reduce the potential for genomic damage over their extended lives, and their expression has been correlated with stem cell activity [90]. ABCG2 expression in the limbus is one such example and cells expressing this marker can be isolated as a 'side population' by fluorescence-activated cell sorting (FACS) [3, 57, 91-95]. However, as noted above, some of the ABCG2-positive, label-retaining cells with a high nucleus to cytoplasm ratio cells in the rat limbus have been identified as Langerhans cells rather than epithelial stem cells [56]. It has recently been shown that ABCB5 appears to be a promising new marker for LESC and early TACs in both mice and humans, which should also allow enrichment by FACS sorting [54].

Despite the absence of a marker that is only expressed in the stem cells,  $\Delta Np63\alpha$  has proved useful for identifying cultures of human limbal cells with sufficient LESC and early TACs for clinical transplantation [37, 96].

## **RESOLVING THE LESC vs. CESC DEBATE**

Some of the evidence discussed so far (summarised in rows 1-15 of Table 1) is inconclusive. Evidence from holoclone experiments with pig and human tissues is inconsistent and, in any case, the critical thing is to understand how the corneal epithelium is maintained *in vivo* during normal homeostasis. Various studies have shown that some cells in the central corneal epithelium are capable of acting as long-term

progenitor cells. On the face of it, this favours the CESC and GBL hypotheses. However, these observations are also consistent with the LESC hypothesis if some basal corneal epithelial TACs have a latent proliferative potential that is only used if LESC function is compromised so homeostasis is disrupted. Drawing attention to this latent proliferative potential is an important outcome of Majo et al's investigations [1] even if the CESC hypothesis ultimately proves to be incorrect.

Other evidence provides better discrimination. There are several strong arguments against the GBL hypothesis. It is inconsistent with the consensus of results from label-retaining cell experiments and the evidence that mosaic patterns in the mouse cornea change after birth, when clones of cells emerge from the peripheral cornea and form radial stripes. The GBL hypothesis also has difficulty accounting for the convincing evidence that corneal epithelial cells move centripetally across the full radius. In our view, this evidence (summarised in Table 1) is sufficient to exclude the GBL hypothesis and it is, therefore, not considered further.

These same observations also argue against the CESC hypothesis. However, results of the label-retaining cell experiments could be accommodated by the CESC hypothesis if CESC cells were not slow cycling. Nevertheless, the CESC hypothesis is inconsistent with the developmental switch from randomly orientated patches to stripes that emerge from the periphery in various types of genetic mosaics in mice and then extend across the full radius to the centre. By the same token, the CESC hypothesis requires *ad hoc* assumptions to account for centripetal movement, if the same clone of cells moves across the full radius.

Conversely, the LESC hypothesis is inconsistent with the observation that when genetically marked limbal tissue was surgically transplanted to the limbus of immunocompromised mice, donor cells failed to move into the cornea unless the corneal epithelium was removed [1]. Thus, the LESC and CESC hypotheses each have difficulty accounting for one type of evidence. In each case, critical evidence is based on experiments with mice so there are no grounds for suggesting that maintenance of the corneal epithelium differs between mice and humans or other species.

Clinically, it may not matter whether the LESC or CESC hypothesis is correct as both agree that the limbus is a suitable source of stem cells for therapeutic use. However, we need to know where the stem cells that maintain the corneal epithelium during normal homeostasis are located in order to understand the biology of this process. The key issue

that needs to be resolved is why evidence from mouse mosaics and transfection with lentiviral markers conflict with results of surgical transplantation experiments. Analysis of mosaics show that, during normal homeostasis, clones of cells appear at the corneal periphery at around 5 weeks after birth and extend centripetally across the corneal radius, consistent with activation of LSCs. In contrast, surgical studies in mice show that transplantation of labelled cells to the limbus fail to colonise the cornea in a similar way.

One possible explanation is that the transplanted limbal tissue failed to colonise the corneal epithelium because the surgical manipulation or other aspects of the experimental procedure somehow perturbed normal homeostasis and affected the outcome. In principle, one way of testing this would be to label individual cells in the limbus of adult mice, using a genetic switch rather than surgical transplantation and test whether any of the labelled cells produce long-lived clones of cells that colonise the corneal epithelium. Similar genetic labelling of cells in the adult basal corneal epithelium would also allow investigation of whether these produce long-lived clones of cells in the corneal epithelium. This requires lineage tracing experiments and some possibilities are discussed below.

It should be borne in mind that some of the apparent contradictions between the different hypotheses may be more imagined than real, and arise as a result of different research groups attempting to subdivide and label what is, perhaps, a continuum of biological situations. It seems certain that limbal stem cells exist, that they can contribute to regeneration of the cornea, and yet that the basal corneal epithelial cells themselves also have massive regenerative potential. There is no reason why the balance between limbal-mediated and corneal-mediated corneal regeneration should not shift over the lifetime of the animal, with age, disease and injury, and no reason why the balance should necessarily be the same in different species.

Although we talk about wounded and unwounded corneas as if they are separate entities, in fact the corneal epithelium is, of necessity constantly regenerating, because of normal desquamation of cells. Desquamation rate is modulated by rate of blinking, tear film composition, irritants and abrasive dust in the environment, chronic abrasion caused by e.g., contact lens wear and diseases such as trachoma, acute minor scratches, ranging through to significant physical or chemical injuries and acute infections such as Herpes simplex keratitis. This continuum of insults to the corneal surface may require different levels of limbal response to support the regenerative potential of the corneal epithelial

cells. It may be that the genuinely uninjured cornea does not require limbal input, but the genuinely uninjured corneal epithelium does not exist. Experimentally, factors such as the abrasive nature of bedding and dust from food that laboratory mice are exposed to, may modulate corneal regeneration and be a source of variation between research institutes. Furthermore, the large circular central corneal wounds that are so widely used as models of induced regeneration do not really recapitulate any of the most common injuries that happen in life.

There may also be problems with the practical definition of stem cells, particularly with regard to their property of 'immortality'. We accept that TACs do not renew indefinitely and therefore have a finite lifespan, whereas stem cells do not have this constraint. However, outside the laboratory, a stem cell cannot outlive the individual. We do not know the maximum lifespan of a TAC but this is likely to vary stochastically. If the longer-lived TACs survive for 6 months or more, this will encompass the effective lifespan of most laboratory mice used experimentally. At that stage, the difference between a long-lived TAC and a stem cell becomes blurred. On the other hand, in larger animals such as humans, a lifespan of 6 months or a year for a TAC is utterly insignificant in terms of the lifespan of the individual. Differences such as this may start to explain why stem-like regenerative ability may be assigned to the corneal epithelium on experimental small animals, while at the same time the data do not reflect clinical experience in humans.

### **Lineage tracing experiments**

To test whether any cells in the limbal and/or corneal epithelia can generate long-lived clones of cells that colonise the corneal epithelium, an inducible lineage tracing method is required that can label some of the putative stem cells in the adult at a chosen time without surgical intervention or disturbing homeostasis. Sophisticated, inducible lineage tracing methods using *Cre/loxP* transgenic mice are now available that can be used to throw a genetic switch to label a chosen cell population with a fluorescent or histochemical marker that will identify them and all their mitotic progeny. Such methods have been used to trace stem cell lineages in other tissues, including the hair follicle [97], intestinal epithelium [98-100] and ovarian surface epithelium [101], and this approach could be used to help resolve the LESC vs. CESC debate. A similar approach has already been used, in conjunction with a multi-coloured reporter construct, to trace clonal lineages in the ocular

surface of zebrafish and demonstrate that, as in mice, the initial patchwork of cells established in the embryo is replaced by a radial pattern of clones that extend from the limbus [102].

We have begun to explore this Cre/*loxP* lineage tracing approach using transgenic mice in which a reporter transgene is ubiquitously expressed once the flanking *loxP* sites are recombined by active Cre recombinase to remove an upstream stop sequence. Cre recombinase is provided in the form of a CreER fusion protein, which is produced by another transgene under the control of a ubiquitous promoter. The CreER fusion protein is normally sequestered in the cytoplasm unless the mouse is treated with tamoxifen. This binds to the modified oestrogen receptor (ER) and translocates CreER to the nucleus, where it can recombine the *loxP* sites, so removing the stop sequence and activating the reporter transgene. Use of a ubiquitous promoter to drive expression of CreER provides an unbiased approach, which allows putative stem cells in the limbus or cornea (and any other tissue) to be labelled. However it is not specific for stem cells so initially most of the labelled cells will not be stem cells but, by including a chase period, short-lived clones founded by labelled TACs will be shed, leaving long-lived clones founded by labelled stem cells.

The genetic switch is activated when the mouse is injected with tamoxifen. Delaying tamoxifen treatment, until well after the adult stem cells are activated, should ensure that the genetic switch is thrown to label individual adult stem cells, during normal homeostasis. This avoids labelling ancestral cells, which could each generate multiple labelled stem cells. Furthermore, by titrating the dose of tamoxifen it should be possible to label a relatively small proportion of cells so only a few stem cells will be labelled per eye. Together, this will ensure that most of the clones of labelled cells that remain as stripes or patches after the chase period are individual clones produced by single stem cells. The predicted results for such an experiment are shown, for the LESC hypothesis and three versions of the CESC hypothesis, in Fig. 5.

Tamoxifen-inducible labelling of a low proportion of stem cells in adults is a significant advantage over analysis of other types of mosaics, where the cells are labelled early in development and many patches and stripes of labelled TACs are likely to be derived from multiple adjacent labelled stem cells. This is because labelling a cell in the embryo will produce a large clone of labelled cells, some of which will remain close

together throughout development. Later, when adult stem cells are specified, some adjacent stem cells will probably be clonally related making it difficult to identify TACs descended from a single stem cell in these mosaic systems.

Examples of results of a preliminary experiment of this type are shown in Fig. 6 and indicate that the strategy suggested in Fig. 5 is feasible. The cornea illustrated in Fig. 6A was stained for  $\beta$ -gal activity after chase period of 9<sup>1</sup>/<sub>2</sub> weeks. Stripe 1 has its peripheral end in the limbus and its more central end in the cornea so, using the stripe classification system described for mosaic *KRT5<sup>LacZ</sup>* transgenic mice [17], it is classified as a limbus-cornea (LC) stripe. This is consistent with expectations of the LESC hypothesis (Time 2 in Fig. 5A), particularly as the stripe does seem to extend into the limbus itself (arrow in Fig. 6A). A peripheral stripe could also be consistent with the original CESC hypothesis (Fig. 5B) or the centripetal version shown in Fig. 5D, although, in the latter case, peripheral stripes would be expected to be entirely within the cornea (CC stripes). This LC stripe fits less well with the hypothesis that all the CESC are located in the very centre of the cornea (Fig. 5C) unless additional assumptions are made (e.g. stripes have already extended to the periphery by 9<sup>1</sup>/<sub>2</sub> weeks).

Stripe 1 also appears to be radially aligned with other  $\beta$ -gal-stained tissue located more centrally so it may be a longer discontinuous stripe. Discontinuous stripes have been discussed elsewhere for *KRT5<sup>LacZ</sup>* transgenic mosaics [17] and also occur in mouse chimaeras and X-inactivation mosaics [27, 72, 103]. The discontinuities in  $\beta$ -gal staining shown in Fig. 6A could reflect (i) separate clones of cells derived from  $\beta$ -gal-positive and  $\beta$ -gal-negative CESC or TACs that are radially aligned, (ii) dispersal of a  $\beta$ -gal-positive clones by incursions from laterally adjacent  $\beta$ -gal-negative clones or (iii) alternating contributions of more than one stem cell to a single radial stripe if individual stem cells cycle through phases of activity and quiescence.

Stripe 2 in Fig. 6A is a cornea-cornea (CC) stripe with both ends in the cornea, consistent with the predictions of the original CESC hypothesis at Time 2 (Fig. 5B) or the centripetal version shown in Fig. 5D. However, it might also be consistent with the LESC hypothesis (Fig. 5A) because this stripe is radially aligned with a small  $\beta$ -gal-stained patch in the limbus, which could mark the location of a  $\beta$ -gal-positive LESC. If so, stripe 2 might be a discontinuous stripe formed in two phases. During the first phase a  $\beta$ -gal-positive region might have extended from a  $\beta$ -gal-positive LESC, which was active during the early

part of the chase period. This could have been followed by extension of a  $\beta$ -gal-negative region generated by an adjacent active  $\beta$ -gal-negative LESC, if the  $\beta$ -gal-positive LESC became inactive during the later part of the chase period.

The eye shown in Fig. 6B was stained after a 14-week chase period and has eight  $\beta$ -gal-positive stripes, all of which are LC stripes, consistent with the LESC hypothesis (Time 3 in Fig. 5A) and both the original CESC hypothesis (Fig. 5B) and the variant shown in Fig. 5C. However, the variant CESC hypothesis shown in Fig. 5D predicts that only a small proportion of stripes would extend right to the limbus. Some of these stripes span the full radius, consistent with the LESC hypothesis (Fig. 5A) and the variant CESC hypothesis shown in Fig. 5C. It could also be consistent with the original CESC hypothesis (Fig. 5B) and the centripetal version, shown in Fig. 5D, if full-radius stripes comprised several shorter stripes produced by  $\beta$ -gal-positive CESC<sub>s</sub> aligned on the same radius. However, most of the cornea is  $\beta$ -gal-negative so it seems unlikely that the *LacZ* reporter would be activated in radially aligned CESC<sub>s</sub> but not in many of the others CESC<sub>s</sub>, located elsewhere the corneal epithelium.

The preliminary results shown in Fig. 6 and the alternative explanations of discontinuous stripes indicate that the interpretation of the stripe patterns may be more complicated than predicted in Fig. 5, so detailed analysis of many more eyes and different chase periods will be required to resolve the LESC vs. CESC debate. Nevertheless, this lineage-tracing approach appears to be a promising way of resolving the conflicting evidence from transplantation experiments and mosaics. Although important, the evidence from conventional mosaics is limited because the time of stem cell labelling cannot be controlled and occurs early in development. Even lineage tracing with a GFP-tagged lentiviral marker (Figs. 4E,F) relied on marking one cell population before birth [79]. In contrast, a tamoxifen-inducible transgenic reporter system enables cells to be labelled at a specific time in the adult without the risk of disturbing homeostasis with surgical intervention.

While this review was in preparation, a similar lineage tracing study was published online, in a preliminary form, and this has already been mentioned in the section on cell movement [64]. This used K14-CreER<sup>T2</sup>;R26R-confetti mice, where the keratin 14 (K14 or *Krt14*) promoter, rather than a ubiquitous promoter, was used to drive tamoxifen-inducible CreER<sup>T2</sup>. The multi-colour 'R26R-confetti' fluorescent reporter [100] was used to



identify labelled cells. This reporter is based on an earlier 'rainbow' construct [104] and is also similar to that used for the zebrafish cornea [102]. According to the authors, immunofluorescence showed that K14 protein was present in basal epithelial cells in the mouse limbus but the central corneal epithelium had much lower levels. Thus, limbal epithelial cells will be preferentially targeted for labelling, making it difficult to test whether CESC exist. On the other hand, a chase period to remove short-lived clones formed by TACs rather than stem cells is less important than for the experimental design shown in Fig. 5, where CreER is expressed ubiquitously.

Mice were treated with tamoxifen at 6 weeks and corneas of the same mice were imaged repeatedly to allow changes in size and position of different clones to be tracked in real time. This elegant time-lapse study showed that labelled cells began to emerge from the limbus 5 weeks after tamoxifen treatment and the same stripes were tracked at 9, 13, 17 and 21 weeks, as they extended into the centre of the unlabelled cornea without overlapping. The frequency of labelled clones was higher than for the preliminary results illustrated for tamoxifen-treated CAGG-CreER;R26R-*LacZ* mice, shown in Fig. 6. However, because the multi-coloured R26R-confetti reporter randomly labelled cells with one of up to ten different colours, individual adjacent clones could be distinguished easily. In addition to the radial stripes, the authors reported the presence of some rare, small patches of labelled cells in the cornea, which they suggested might have arisen in the cornea itself.

This is an important study as it is the first lineage tracing investigation of the mouse cornea with an inducible marker that was activated in adults. In particular, the informative, real-time study with live mice proved that individual cells labelled in the adult limbal epithelium could form long-lived clones that extend centripetally across the complete radius of the cornea during normal homeostasis. This effectively provides a non-surgical equivalent of the limbal transplantation experiment described by Majo et al [1] and supports the interpretation that the outcome of the transplantation was adversely affected by the surgical procedures and homeostasis was perturbed. If so, this would undermine the evidence for the CESC hypothesis and reconcile the conflicting evidence from surgical transplantation experiments and conventional mosaics. In effect, all the available evidence would then be consistent with the LESC hypothesis. To further clarify the situation, additional investigations, driving CreER from a promoter expressed in the corneal

epithelium, are now required to investigate whether long-lived clones can arise in the adult cornea as well as the limbus.

## **CONCLUSION**

There is strong evidence that the corneal epithelium is maintained by stem cells rather than solely by proliferation of more differentiated cells in the basal corneal epithelium. Most evidence also favours the conventional LESC hypothesis, which proposes that limbal epithelial stem cells maintain the corneal epithelium during normal homeostasis. Although limbal transplantation experiments favour the alternative CESC hypothesis, this result could be reconciled with the LESC hypothesis if surgical transplantation perturbed normal homeostasis and affected the outcome. This possibility is supported by a recent non-surgical, lineage-tracing experiment, which demonstrated that clonal derivatives of cells in the limbal epithelium move into the corneal epithelium during normal homeostasis. Thus, the available evidence supports the conclusion that, during normal homeostasis, the corneal epithelium is maintained by stem cells in the limbus, which produce daughter TACs that migrate centripetally, rather than any stem cells in the corneal epithelium itself. However, if homeostasis is compromised, so the limbal epithelial stem cells are unable to maintain the corneal epithelium, it seems likely that TACs in the basal corneal epithelium can act as long-term progenitors and maintain the tissue for a considerable time in the absence of functional LESCs.

## **ACKNOWLEDGEMENTS**

We thank Dr Steven D. Morley for helpful discussion and critical comments on the manuscript and Mr. Ronnie Grant for assistance with illustrations. We also thank Dr Alan W. Flake, Dr Takayuki Nagasaki, Springer Science+Business Media, Developmental Dynamics, Molecular Vision and Molecular Therapy for permission to reproduce images as indicated in the figure legends. We are grateful to the Biotechnology and Biological Sciences Research Council and the Wellcome Trust for support for our own research.

## REFERENCES

- 1 **Majo F**, Rochat A, Nicolas M, Jaoude GA, Barrandon Y. Oligopotent stem cells are distributed throughout the mammalian ocular surface. *Nature* 2008; **456**: 250-255 [PMID: 18830243]
- 2 **Haddad A**, Faria-E-Sousa SJ. Maintenance of the corneal epithelium is carried out by germinative cells of its basal stratum and not by presumed stem cells of the limbus. *Braz J Med Biol Res* 2014; **47**: 470-477 [PMID: 24820068]
- 3 **Chen Z**, De Paiva CS, Luo LH, Kretzer FL, Pflugfelder SC, Li DQ. Characterization of putative stem cell phenotype in human limbal epithelia. *Stem Cells* 2004; **22**: 355-366 [PMID: 15153612]
- 4 **Schermer A**, Galvin S, Sun TT. Differentiation-related expression of a major 64K corneal keratin in vivo and in culture suggests limbal location of corneal epithelial stem cells. *J Cell Biol* 1986; **103**: 49-62 [PMID: 2424919]
- 5 **Cotsarelis G**, Cheng SZ, Dong G, Sun TT, Lavker RM. Existence of slow-cycling limbal epithelial basal cells that can be preferentially stimulated to proliferate: Implications on epithelial stem cells. *Cell* 1989; **57**: 201-209 [PMID: 2702690]
- 6 **Li W**, Hayashida Y, Chen YT, Tseng SCG. Niche regulation of corneal epithelial stem cells at the limbus. *Cell Res* 2007; **17**: 26-36 [PMID: 17211449]
- 7 **Xie H-T**, Chen S-Y, Li G-G, Tseng SCG. Limbal epithelial stem/progenitor cells attract stromal niche cells by SDF-1/CXCR4 signaling to prevent differentiation. *Stem Cells* 2011; **29**: 1874-1885 [PMID: 21948620 DOI: 10.1002/stem.743]
- 8 **Shortt AJ**, Secker GA, Munro PM, Khaw PT, Tuft SJ, Daniels JT. Characterization of the limbal epithelial stem cell niche: Novel imaging techniques permit in vivo observation and targeted biopsy of limbal epithelial stem cells. *Stem Cells* 2007; **25**: 1402-1409 [PMID: 17332511 DOI: 10.1634/stemcells.2006-0580]
- 9 **Dziasko MA**, Armer HE, Levis HJ, Shortt AJ, Tuft S, Daniels JT. Localisation of epithelial cells capable of holoclone formation *in vitro* and direct interaction with stromal cells in the native human limbal crypt. *PLoS One* 2014; **9**: e94283 [PMID: 24714106 DOI: 10.1371/journal.pone.0094283]
- 10 **Dua HS**, Shanmuganathan VA, Powell-Richards AO, Tighe PJ, Joseph A. Limbal epithelial crypts: A novel anatomical structure and a putative limbal stem cell niche. *Br J Ophthalmol* 2005; **89**: 529-532 [PMID: 15834076]

- 11 **Kulkarni BB**, Tighe PJ, Mohammed I, Yeung AM, Powe DG, Hopkinson A, Shanmuganathan VA, Dua HS. Comparative transcriptional profiling of the limbal epithelial crypt demonstrates its putative stem cell niche characteristics. *BMC Genomics* 2010; **11**:526 [PMID: 20920242 DOI: 10.1186/1471-2164-11-526]
- 12 **Pajooresh-Ganji A**, Pal-Ghosh S, Tadvalkar G, Stepp MA. Corneal goblet cells and their niche: Implications for corneal stem cell deficiency. *Stem Cells* 2012; **30**: 2032-2043 [PMID: 22821715 DOI: 10.1002/stem.1176]
- 13 **Lehrer MS**, Sun TT, Lavker RM. Strategies of epithelial repair: Modulation of stem cell and transit amplifying cell proliferation. *J Cell Sci* 1998; **111**: 2867-2875 [PMID: 9730979]
- 14 **Beebe DC**, Masters BR. Cell lineage and the differentiation of corneal epithelial cells. *Invest Ophthalmol & Vis Sci* 1996; **37**: 1815-1825 [PMID: 8759349]
- 15 **Marinari E**, Mehonic A, Curran S, Gale J, Duke T, Baum B. Live-cell delamination counterbalances epithelial growth to limit tissue overcrowding. *Nature* 2012; **484**: 542-546 [PMID: 22504180 DOI: 10.1038/nature10984]
- 16 **Sun TT**, Tseng SC, Lavker RM. Location of corneal epithelial stem cells. *Nature* 2010; **463**: E10 [PMID: 20182462 DOI: 10.1038/nature08805]
- 17 **Douvaras P**, Webb S, Whitaker DA, Dorà N, Hill RE, Dorin JR, West JD. Rare corneal clones in mice suggest an age-related decrease of stem cell activity and support the limbal epithelial stem cell hypothesis. *Stem Cell Res* 2012; **8**: 109-119 [PMID: 22099025 DOI: 10.1016/j.scr.2011.08.007]
- 18 **Mort RL**, Douvaras P, Morley SD, Dorà N, Hill RE, Collinson JM, West JD. Stem cells and corneal epithelial maintenance: Insights from the mouse and other animal models. In: Kubiak JZ *Results Probl Cell Differ*, **55**, "Mouse development: From oocyte to stem cells". Berlin & Heidelberg: Springer-Verlag, 2012: 357-394 [PMID: 2918816 DOI: 10.1007/978-3-642-30406-4\_19]
- 19 **Dor Y**, Brown J, Martinez OI, Melton DA. Adult pancreatic beta-cells are formed by self-duplication rather than stem-cell differentiation. *Nature* 2004; **429**: 41-46 [PMID: 15129273 DOI: 10.1038/nature02520]
- 20 **Clayton E**, Doupe DP, Klein AM, Winton DJ, Simons BD, Jones PH. A single type of progenitor cell maintains normal epidermis. *Nature* 2007; **446**: 185-189 [PMID: 17330052]

- 21 **Giangreco A**, Arwerta EN, Rosewell IR, Snyder J, Watt FM, Stripp BR. Stem cells are dispensable for lung homeostasis but restore airways after injury. *Proc Natl Acad Sci USA* 2009; **106**: 9286-9291 [PMID: DOI: 10.1073/pnas.0900668106]
- 22 **Miyajima A**, Tanaka M, Itoh T. Stem/progenitor cells in liver development, homeostasis, regeneration, and reprogramming. *Cell Stem Cell* 2014; **14**: 561-574 [PMID: 24792114 DOI: 10.1016/j.stem.2014.04.010]
- 23 **Tarlow BD**, Finegold MJ, Grompe M. Clonal tracing of Sox9<sup>+</sup> liver progenitors in mouse oval cell injury. *Hepatology* 2014; **60**: 278-289 [PMID: 24700457 DOI: 10.1002/hep.27084]
- 24 **Barrandon Y**, Green H. Three clonal types of keratinocyte with different capacities for multiplication. *Proc Natl Acad Sci USA* 1987; **84**: 2302-2306 [PMID: 2436229]
- 25 **Pellegrini G**, Golisano O, Paterna P, Lambiase A, Bonini S, Rama P, De Luca M. Location and clonal analysis of stem cells and their differentiated progeny in the human ocular surface. *J Cell Biol* 1999; **145**: 769-782 [PMID: 10330405]
- 26 **Chung EH**, Bukusoglu G, Zieske JD. Localization of corneal epithelial stem-cells in the developing rat. *Invest Ophthalmol & Vis Sci* 1992; **33**: 2199-2206 [PMID: 1607230]
- 27 **Collinson JM**, Morris L, Reid AI, Ramaesh T, Keighren MA, Flockhart JH, Hill RE, Tan SS, Ramaesh K, Dhillon B, West JD. Clonal analysis of patterns of growth, stem cell activity, and cell movement during the development and maintenance of the murine corneal epithelium. *Dev Dyn* 2002; **224**: 432-440 [PMID: 12203735]
- 28 **Tanifuji-Terai N**, Terai K, Hayashi Y, Chikama TI, Kao WWY. Expression of Keratin 12 and maturation of corneal epithelium during development and postnatal growth. *Invest Ophthalmol & Vis Sci* 2006; **47**: 545-551 [PMID: 16431949]
- 29 **Chang CYA**, McGhee JJ, Green CR, Sherwin T. Comparison of stem cell properties in cell populations isolated from human central and limbal corneal epithelium. *Cornea* 2011; **30**: 1155-1162 [PMID: 21849892 DOI: 10.1097/ICO.0b013e318213796b]
- 30 **Lavker RM**, Dong G, Cheng SZ, Kudoh K, Cotsarelis G, Sun TT. Relative proliferative rates of limbal and corneal epithelia - implications of corneal epithelial migration, circadian rhythm, and suprabasally located DNA-synthesizing keratinocytes. *Invest Ophthalmol & Vis Sci* 1991; **32**: 1864-1875 [PMID: 2032808]

- 31 **Ebato B**, Friend J, Thoft RA. Comparison of limbal and peripheral human corneal epithelium in tissue culture. *Invest Ophthalmol & Vis Sci* 1988; **29**: 1533-1537 [PMID: 3170124]
- 32 **Kenyon KR**, Tseng SCG. Limbal autograft transplantation for ocular surface disorders. *Ophthalmology* 1989; **96**: 709-722 [PMID: 2748125]
- 33 **Tseng SCG**. Concept and application of limbal stem cells. *Eye* 1989; **3**: 141-157 [PMID: 2695347]
- 34 **Pellegrini G**, Traverso CE, Franzi AT, Zingirian M, Cancedda R, DeLuca M. Long-term restoration of damaged corneal surfaces with autologous cultivated corneal epithelium. *Lancet* 1997; **349**: 990-993 [PMID: 9100626 DOI: 10.1016/s0140-6736(96)11188-0]
- 35 **Tsai RJ**, Li LM, Chen JK. Reconstruction of damaged corneas by transplantation of autologous limbal epithelial cells. *New Eng J Med* 2000; **343**: 86-93 [PMID: 10891515 DOI: 10.1056/nejm200007133430202]
- 36 **Sangwan VS**, Vemuganti GK, Iftekhar G, Bansal AK, Rao GN. Use of autologous cultured limbal and conjunctival epithelium in a patient with severe bilateral ocular surface disease induced by acid injury - a case report of unique application. *Cornea* 2003; **22**: 478-481 [PMID: 12827056 DOI: 10.1097/00003226-200307000-00016]
- 37 **Rama P**, Matuska S, Paganoni G, Spinelli A, De Luca M, Pellegrini G. Limbal stem-cell therapy and long-term corneal regeneration. *New Eng J Med* 2010; **363**: 147-155 [PMID: 20573916 DOI: 10.1056/NEJMoa0905955]
- 38 **Shortt AJ**, Tuft SJ, Daniels JT. Corneal stem cells in the eye clinic. *Br Med Bull* 2011; **100**: 209-225 [PMID: 21926089 DOI: 10.1093/bmb/ldr041]
- 39 **Huang AJW**, Tseng SCG. Corneal epithelial wound healing in the absence of limbal epithelium. *Invest Ophthalmol & Vis Sci* 1991; **32**: 96-105 [PMID: 1702774]
- 40 **Kawakita T**, Higa K, Shimmura S, Tomita M, Tsubota K, Shimazaki J. Fate of corneal epithelial cells separated from limbus in vivo. *Invest Ophthalmol & Vis Sci* 2011; **52**: 8132-8137 [PMID: 21896841 DOI: 10.1167/iovs.11-7984]
- 41 **Dua HS**, Miri A, Alomar T, Yeung AM, Said DG. The role of limbal stem cells in corneal epithelial maintenance testing the dogma. *Ophthalmology* 2009; **116**: 856-863 [PMID: 19410942 DOI: 10.1016/j.opthta.2008.12.017]
- 42 **Bi Y-L**, Bock F, Zhou Q, Cursiefen C. Central corneal epithelium self-healing after ring-shaped glycerin-cryopreserved lamellar keratoplasty in terrien marginal

- degeneration. *Int J Ophthalmol* 2013; **6**: 251-252 [PMID: 23638432 DOI: 10.3980/j.issn.2222-3959.2013.02.27]
- 43 **Castro-Munozledo F**, Gomez-Flores E. Challenges to the study of asymmetric cell division in corneal and limbal epithelia. *Exp Eye Res* 2011; **92**: 4-9 [PMID: 21056036 DOI: 10.1016/j.exer.2010.11.002]
- 44 **Bertalanffy FD**, Lau C. Mitotic rate and renewal time of corneal epithelium in rat. *Arch Ophthalmol* 1962; **68**: 546-50 [PMID: 13868376]
- 45 **Douvaras P**, Mort RL, Edwards D, Ramaesh K, Dhillon B, Morley SD, Hill RE, West JD. Increased corneal epithelial turnover contributes to abnormal homeostasis in the *Pax6<sup>+/-</sup>* mouse model of aniridia. *PLoS One* 2013 **8**: e71117 [PMID: 23967157 DOI: 10.1371/journal.pone.0071117]
- 46 **Urbanowicz MM**, Zhao J, Nagasaki T. Spatial distribution of cell divisions in the basal epithelium of mouse cornea. *Invest Ophthalmol & Vis Sci* 2011; (ARVO Meeting Abstracts April 22, 2011) **52**: E-abstract 320
- 47 **Hanna C**, O'Brien JE. Cell production and migration in the epithelial layer of the cornea. *Arch Ophthalmol* 1960; **64**: 536-539 [PMID: 13711262]
- 48 **Hanna C**, Bicknell DS, Obrien JE. Cell turnover in adult human eye *Arch Ophthalmol* 1961; **65**: 695-8 [PMID: 13711260]
- 49 **Haskjold E**, Bjerknes R, Bjerknes E. Migration of cells in the rat corneal epithelium. *Acta Ophthalmol* 1989; **67**: 91-96 [PMID: 2773642]
- 50 **Cenedella RJ**, Fleschner CR. Kinetics of corneal epithelium turnover in vivo - studies of lovastatin. *Invest Ophthalmol & Vis Sci* 1990; **31**: 1957-1962 [PMID: 2210991]
- 51 **Haddad A**. Renewal of the rabbit corneal epithelium as investigated by autoradiography after intravitreal injection of <sup>3</sup>H-thymidine. *Cornea* 2000; **19**: 378-383 [PMID: 10832703]
- 52 **Pajoohesh-Ganji A**, Pal-Ghosh S, Simmens SJ, Stepp MA. Integrins in slow-cycling corneal epithelial cells at the limbus in the mouse. *Stem Cells* 2006; **24**: 1075-1086 [PMID: 16282441]
- 53 **Zhao J**, Mo V, Nagasaki T. Distribution of label-retaining cells in the limbal epithelium of a mouse eye. *J Histochem Cytochem* 2009; **57**: 177-185 [PMID: 19001638 DOI: 10.1369/jhc.2008.952390]



- 54 **Ksander BR**, Kolovou PE, Wilson BJ, Saab KR, Guo Q, Ma J, McGuire SP, Gregory MS, Vincent WJB, Perez VL, Cruz-Guilloty F, Kao WWY, Call MK, Tucker BA, Zhan Q, Murphy GF, Lathrop KL, Alt C, Mortensen LJ, Lin CP, Zieske JD, Frank MH, Frank NY. *ABCB5* is a limbal stem cell gene required for corneal development and repair. *Nature* 2014; **511**: 353-357 [PMID: 25030174; DOI: 10.1038/nature13426]
- 55 **Chen WS**, Ishikawa M, Yamaki K, Sakuragi S. Wistar rat palpebral conjunctiva contains more slow-cycling stem cells that have larger proliferative capacity: Implication for conjunctival epithelial homeostasis. *Jap J Ophthalmol* 2003; **47**: 119-128 [PMID: 12738543 DOI: 10.1016/s0021-5155(02)00687-1]
- 56 **Chen W**, Hara K, Tian Q, Zhao K, Yoshitomi T. Existence of small slow-cycling Langerhans cells in the limbal basal epithelium that express ABCG2. *Exp Eye Res* 2007; **84**: 626-634 [PMID: 17254566 DOI: 10.1016/j.exer.2006.11.006]
- 57 **Budak MT**, Alpdogan OS, Zhou MY, Lavker RM, Akinci MAM, Wolosin JM. Ocular surface epithelia contain ABCG2-dependent side population cells exhibiting features associated with stem cells. *J Cell Sci* 2005; **118**: 1715-1724 [PMID: 15811951]
- 58 **Davanger M**, Evensen A. Role of the pericorneal papillary structure in renewal of corneal epithelium. *Nature* 1971; **229**: 560-561 [PMID: 4925352]
- 59 **Kinoshita S**, Friend J, Thoft RA. Sex chromatin of donor corneal epithelium in rabbits. *Invest Ophthalmol & Vis Sci* 1981; **21**: 434-441 [PMID: 7024181]
- 60 **Lemp MA**, Mathers WD. Corneal epithelial cell movement in humans. *Eye* 1989; **3**: 438-445 [PMID: 2606218]
- 61 **Auran JD**, Koester CJ, Kleiman NJ, Rapaport R, Bomann JS, Wirotko BM, Florakis GJ, Koniarek JP. Scanning slit confocal microscopic observation of cell morphology and movement within the normal human anterior cornea. *Ophthalmology* 1995; **102**: 33-41 [PMID: 7831039 ]
- 62 **Buck RC**. Measurement of centripetal migration of normal corneal epithelial cells in the mouse. *Invest Ophthalmol & Vis Sci* 1985; **26**: 1296-1299 [PMID: 4030257]
- 63 **Nagasaki T**, Zhao J. Centripetal movement of corneal epithelial cells in the normal adult mouse. *Invest Ophthalmol & Vis Sci* 2003; **44**: 558-566 [PMID: 12556383]
- 64 **Di Girolamo N**, Bobba S, Raviraj V, Delic NC, Slapetova I, Nicovich PR, Halliday GM, Wakefield D, Whan R, Lyons GJ. Tracing the fate of limbal epithelial progenitor cells

- in the murine cornea. *Stem Cells* 2014; **Epub ahead of print** [PMID: 24966117 DOI: 10.1002/stem.1769]
- 65 **Nagasaki T**, Zhao J. Uniform distribution of epithelial stem cells in the bulbar conjunctiva. *Invest Ophthalmol & Vis Sci* 2005; **46**: 126-132 [PMID: 15623764]
- 66 **Bron AJ**. Vortex patterns of corneal epithelium. *Trans Ophthalmol Soc UK* 1973; **93**: 455-472 [PMID: 4210604]
- 67 **Sharma A**, Coles WH. Kinetics of corneal epithelial maintenance and graft loss - a population balance model *Invest Ophthalmol Vis Sci* 1989; **30**: 1962-1971 [PMID: 2674050]
- 68 **Wolosin JM**, Xiong XL, Schutte M, Stegman Z, Tieng A. Stem cells and differentiation stages in the limbo-corneal epithelium. *Prog Retin Eye Res* 2000; **19**: 223-255 [PMID: 10674709 DOI: 10.1016/s1350-9462(99)00005-1]
- 69 **Foster JW**, Jones RR, Bippes CA, Gouveia RJ, Connon CJ. Differential nuclear expression of Yap in basal epithelial cells across the cornea and substrates of differing stiffness. *Exp Eye Res* 2014; **127**: 37-41 [PMID: 24992208 DOI: 10.1016/j.exer.2014.06.020]
- 70 **Jones MA**, Marfurt CF. Sympathetic stimulation of corneal epithelial proliferation in wounded and nonwounded rat eyes. *Invest Ophthalmol Vis Sci* 1996; **37**: 2535-2547 [PMID: 8977468]
- 71 **McCaig CD**, Rajnicek AM, Song B, Zhao M. Controlling cell behavior electrically: Current views and future potential. *Physiol Rev* 2005; **85**: 943-978 [PMID: 15987799 DOI: 10.1152/physrev.00020.2004]
- 72 **Mort RL**, Ramaesh T, Kleinjan DA, Morley SD, West JD. Mosaic analysis of stem cell function and wound healing in the mouse corneal epithelium. *BMC Dev Biol* 2009; **9**: 4 [PMID: 19128502 DOI: 10.1186/1471-213X-9-4.]
- 73 **Iannaccone S**, Zhou Y, Walterhouse D, Taborn G, Landini G, Iannaccone P. Three dimensional visualization and fractal analysis of mosaic patches in rat chimeras: Cell assortment in liver, adrenal cortex and cornea. *PLoS One* 2012; **7**: e31609 [PMID: DOI: 22347498 DOI:10.1371/journal.pone.0031609]
- 74 **Chong EM**, Campbell RJ, Bourne WM. Vortex keratopathy in a patient with multiple myeloma. *Cornea* 1997; **16**: 592-594 [PMID: 9294696]
- 75 **Dua HS**, Gomes JAP. Clinical course of hurricane keratopathy. *Br J Ophthalmol* 2000; **84**: 285-288 [PMID: 10684839]

- 76 **Bradshaw JJ**, Obritsch WF, Cho BJ, Gregerson DS, Holland EJ. Ex vivo transduction of corneal epithelial progenitor cells using a retroviral vector. *Invest Ophthalmol & Vis Sci* 1999; **40**: 230-235 [PMID: 9888447 ]
- 77 **Zhang W**, Zhao J, Chen L, Urbanowicz MM, Nagasaki T. Abnormal epithelial homeostasis in the cornea of mice with a destrin deletion. *Mol Vis* 2008; **14**: 1929-1939 [PMID: 18958303]
- 78 **Hayashi Y**, Watanabe N, Ohashi Y. The “replacement hypothesis”: Corneal stem cell origin epithelia are replaced by limbal stem cell origin epithelia in mouse cornea during maturation. *Cornea* 2012; **31**, Suppl 1: S68-S73 [PMID: 23038039]
- 79 **Endo M**, Zoltick PW, Chung DC, Bennett J, Radu A, Muvarak N, Flake AW. Gene transfer to ocular stem cells by early gestational intraamniotic injection of lentiviral vector. *Mol Therap* 2007; **15**: 579-587 [PMID: 17245352]
- 80 **Waring GO III**, Roth AM, Ekins MB. Clinical and pathologic description of 17 cases of corneal intraepithelial neoplasia. *Am J Ophthalmol* 1984; **97**: 547-559 [PMID: 6720832]
- 81 **Reya T**, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001; **414**: 105-111 [PMID: 11689955]
- 82 **Burkert J**, Wright NA, Alison MR. Stem cells and cancer: an intimate relationship. *J Pathol* 2006; **209**: 287-297 [PMID: 16770755 DOI: 10.1002/path.2016]
- 83 **Wagers AJ**. The stem cell niche in regenerative medicine. *Cell Stem Cell* 2012; **10**: 362-369 [PMID 22482502: DOI: 10.1016/j.stem.2012.02.018]
- 84 **Culver JC**, Vadakkan TJ, Dickinson ME. A specialized microvascular domain in the mouse neural stem cell niche. *PLoS One* 2013; **8**: e53546 [PMID: 23308251 DOI: 10.1371/journal.pone.0053546]
- 85 **Barker N**. Adult intestinal stem cells: critical drivers of epithelial homeostasis and regeneration. *Nat Rev Mol Cell Biol* 2014; **15**: 19-33 [PMID: 24326621 DOI: 10.1038/nrm3721]
- 86 **Chaloin-Dufau C**, Pavitt I, Delorme P, Dhouailly D. Identification of Keratin-3 and Keratin-12 in corneal epithelium of vertebrates. *Epithelial Cell Biol* 1993; **2**: 120-125 [PMID: 7688259]
- 87 **Liu CY**, Zhu G, Westerhausenlarsen A, Converse R, Kao CWC, Sun TT, Kao WWY. Cornea-specific expression of K12 keratin during mouse development. *Curr Eye Res* 1993; **12**: 963-974 [PMID: 7508359]

- 88 **Romano AC**, Espana EM, Yoo SH, Budak MT, Wolosin JM, Tseng SCG. Different cell sizes in human limbal and central corneal basal epithelia measured by confocal microscopy and flow cytometry. *Invest Ophthalmol & Vis Sci* 2003; **44**: 5125-5129 [PMID: 14638707 DOI: 10.1167/iovs.03-0628]
- 89 **Rees DC**, Johnson E, Lewinson O. ABC transporters: the power to change. *Nat Rev Mol Cell Biol* 2009; **10**: 218-227 [PMID: 19234479 DOI: 10.1038/nrm2646]
- 90 **Bunting KD**. ABC transporters as phenotypic markers and functional regulators of stem cells. *Stem Cells* 2002; **20**: 11-20 [PMID: 11796918 ]
- 91 **Watanabe K**, Nishida K, Yamato M, Umemoto T, Sumide T, Yamamoto K, Maeda N, Watanabe H, Okano T, Tano Y. Human limbal epithelium contains side population cells expressing the ATP-binding cassette transporter ABCG2. *FEBS Lett* 2004; **565**: 6-10 [PMID: 15135043 DOI: 10.1016/j.febslet.2004.03.064]
- 92 **de Paiva CS**, Chen Z, Corrales RM, Pflugfelder SC, Li DQ. ABCG2 transporter identifies a population of clonogenic human limbal epithelial cells. *Stem Cells* 2005; **23**: 63-73 [PMID: 15625123 DOI: 10.1634/stemcells.2004-0093]
- 93 **Umemoto T**, Yamato M, Nishida K, Kohno C, Yang J, Tano Y, Okano T. Rat limbal epithelial side population cells exhibit a distinct expression of stem cell markers that are lacking in side population cells from the central cornea. *FEBS Lett* 2005; **579**: 6569-6574 [PMID: 16297384 DOI: 10.1016/j.febslet.2005.10.047]
- 94 **Umemoto T**, Yamato M, Nishida K, Yang J, Tano Y, Okano T. Limbal epithelial side-population cells have stem cell-like properties, including quiescent state. *Stem Cells* 2006; **24**: 86-94 [PMID 16150918: DOI: 10.1634/stemcells.2005-0064]
- 95 **Krulova M**, Pokorna K, Lencova A, Fric J, Zajicova A, Filipiec M, Forrester JV, Holan V. A rapid separation of two distinct populations of mouse corneal epithelial cells with limbal stem cell characteristics by centrifugation on percoll gradient. *Invest Ophthalmol & Vis Sci* 2008; **49**: 3903-3908 [PMID: 18469183 DOI: 10.1167/iovs.08-1987]
- 96 **Di Iorio E**, Barbaro V, Ruzza A, Ponzin D, Pellegrini G, De Luca M. Isoforms of  $\Delta$ Np63 and the migration of ocular limbal cells in human corneal regeneration. *Proc Natl Acad Sci USA* 2005; **102**: 9523-9528 [PMID: 15983386 DOI: 10.1073/pnas.0503437102]
- 97 **Morris RJ**, Liu YP, Marles L, Yang ZX, Trempus C, Li SL, Lin JS, Sawicki JA, Cotsarelis G. Capturing and profiling adult hair follicle stem cells. *Nat Biotech* 2004; **22**: 411-417 [PMID: 15024388 DOI: 10.1038/nbt950]

- 98 **Barker N**, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegebarth A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene *lgr5*. *Nature* 2007; **449**: 1003-1007 [PMID: 17934449]
- 99 **Sangiorgi E**, Capecchi MR. *Bmi1* is expressed *in vivo* in intestinal stem cells. *Nat Genet* 2008; **40**: 915-920 [PMID: 18536716]
- 100 **Snippert HJ**, van der Flier LG, Sato T, van Es JH, van den Born M, Kroon-Veenboer C, Barker N, Klein AM, van Rheenen J, Simons BD, Clevers H. Intestinal crypt homeostasis results from neutral competition between symmetrically dividing *Lgr5* stem cells. *Cell* 2010; **143**: 134-144 [PMID: 20887898 DOI: 10.1016/j.cell.2010.09.016]
- 101 **Ng A**, Tan S, Singh G, Rizk P, Swathi Y, Tan TZ, Huang RY-J, Leushacke M, Barker N. *Lgr5* marks stem/progenitor cells in ovary and tubal epithelia. *Nat Cell Biol* 2014; **Epub ahead of print**: [PMID: 24997521]
- 102 **Pan YA**, Freundlich T, Weissman TA, Schoppik D, Wang XC, Zimmerman S, Ciruna B, Sanes JR, Lichtman JW, Schier AF. Zebrafish: multispectral cell labeling for cell tracing and lineage analysis in zebrafish. *Development* 2013; **140**: 2835-2846 [PMID: 23757414 DOI: 10.1242/dev.094631]
- 103 **Collinson JM**, Chanas SA, Hill RE, West JD. Corneal development, limbal stem cell function, and corneal epithelial cell migration in the *Pax6<sup>+/-</sup>* mouse *Invest Ophthalmol & Vis Sci* 2004; **45**: 1101-1108 [PMID: 15037575]
- 104 **Livet J**, Weissman TA, Kang HN, Draft RW, Lu J, Bennis RA, Sanes JR, Lichtman JW. Transgenic strategies for combinatorial expression of fluorescent proteins in the nervous system. *Nature* 2007; **450**: 56-62 [PMID: 17972876]
- 105 **Stepp MA**, Zhu L, Sheppard D, Cranfill RL. Localized distribution of  $\alpha 9$  integrin in the cornea and changes in expression during corneal epithelial cell differentiation. *J Histochem Cytochem* 1995; **43**: 353-362 [PMID: 7534781]
- 106 **Pajooresh-Ganji A**, Ghosh SP, Stepp MA. Regional distribution of  $\alpha 9\beta 1$  integrin within the limbus of the mouse ocular surface. *Dev Dyn* 2004; **230**: 518-528 [PMID: 15188436]
- 107 **Pellegrini G**, Dellambra E, Golisano O, Martinelli E, Fantozzi I, Bondanza S, Ponzin D, McKeon F, De Luca M. p63 identifies keratinocyte stem cells. *Proc Natl Acad Sci USA* 2001; **98**: 3156-3161 [PMID: 11248048]

- 108 **Schlötzer-Schrehardt U**, Kruse FE. Identification and characterization of limbal stem cells. *Exp Eye Res* 2005; **81**: 247-264 [PMID: 16051216]
- 109 **Yoshida S**, Shimmura S, Kawakita T, Miyashita H, Den S, Shimazaki J, Tsubota K. Cytokeratin 15 can be used to identify the limbal phenotype in normal and diseased ocular surfaces. *Invest Ophthalmol & Vis Sci* 2006; **47**: 4780-4786 [PMID: 17065488]
- 110 **Hayashi R**, Yamato M, Sugiyama H, Sumide T, Yang J, Okano T, Tano Y, Nishida K. N-cadherin is expressed by putative stem/progenitor cells and melanocytes in the human limbal epithelial stem cell niche. *Stem Cells* 2007; **25**: 289-296 [PMID: 17008425 DOI: 10.1634/stemcells.2006-0167]
- 111 **Barbaro V**, Testa A, Di Iorio E, Mavilio F, Pellegrini G, De Luca M. C/EBP $\delta$  regulates cell cycle and self-renewal of human limbal stem cells. *J Cell Biol* 2007; **177**: 1037-1049 [PMID: 17562792 DOI: 10.1083/jcb.200703003]
- 112 **Moore JE**, McMullen CBT, Mahon G, Adamis AP. The corneal epithelial stem cell. *DNA and Cell Biology* 2002; **21**: 443-451 [PMID: 12167247]
- 113 **Ramaesh T**, Ramaesh K, Collinson JM, Chanas SA, Dhillon B, West JD. Developmental and cellular factors underlying corneal epithelial dysgenesis in the *Pax6*<sup>+/-</sup> mouse model of aniridia. *Exp Eye Res* 2005; **81**: 224-235 [PMID: 16080917]
- 114 **Goldberg MF**, Bron AJ. Limbal palisades of Vogt. *Trans Am Ophthalmol Soc* 1982; **80**: 155-171 [PMID: 7182957]
- 115 **Townsend W**. The limbal palisades of Vogt. *Trans Am Ophthalmol Soc* 1991; **89**: 721-756 [PMID: 1808821]
- 116 **Hayashi S**, McMahan AP. Efficient recombination in diverse tissues by a tamoxifen- inducible form of Cre: A tool for temporally regulated gene activation/inactivation in the mouse. *Dev Biol* 2002; **244**: 305-318 [PMID: 11944939]
- 117 **Soriano P**. Generalized lacZ expression with the ROSA26 Cre reporter strain. *Nature Genetics* 1999; **21**: 70-71 [PMID: 9916792]

**Table 1: Evidence discriminating between alternative hypotheses**

Observations	Species	References	Consistent with hypothesis?		
			LESC	CESC	GBL
1. Holoclone-producing cells are present in limbus but not cornea	human	[25]	++	±	±
2. Holoclone-producing cells are present in limbus and cornea	pig	[1]	±	++	±
3. Production of clonogenic spheres from limbus and central cornea	human	[29]	±	++	±
4. Limbal epithelial cells are superior to corneal epithelial cells for corneal repair.	human	[32, 33]	++	+	±
5. The central corneal epithelium can maintain itself when isolated from the limbus	rabbit, mouse, human	[1, 39-42]	+ <sup>a</sup>	++	++
6. Label-retaining cells are present in the limbus but not the corneal epithelium during normal homeostasis	mouse, rabbit	[13, 52-54, 57]	++	+	0
7. After a 40-day chase <sup>3</sup> H-TdR-labelled cells are present in both the limbus and the corneal epithelium during normal homeostasis	rabbit	[2]	+ <sup>b</sup>	+ <sup>b</sup>	+ <sup>b</sup>
8. Cells move centripetally from the periphery of the corneal epithelium during normal homeostasis	human, mouse	[58, 61-63]	++	± <sup>c</sup>	± <sup>c</sup>
9. Mosaic patterns change after birth and clones of labelled cells emerge from the limbus as radial stripes	mouse	[27, 72, 77-79]	++	0	0
10. Distribution of rare stripes in corneas of <i>KRT5-LacZ</i> <sup>+/-</sup> transgenic mice	mouse	[17]	++	±	±
11. Transplanted limbal tissue contributes to replacing debrided corneal epithelium	rabbit, mouse	[1, 76]	++	++	+

12. Transplanted limbal tissue does not contribute to the unwounded corneal epithelium	mouse	[1]	0	++	++
13. More tumours arise from the limbal epithelium than corneal epithelium	human	[80]	++ <sup>d</sup>	+	+
14. Diverse cell types and blood supply makes limbus a likely stem cell niche	all species	[6]	++ <sup>d</sup>	+	+
15. Distribution of markers associated with undifferentiated or stem cell phenotype	human, mouse, rabbit	See Table 2	++	±	±
16. Lineage tracing studies show that limbal cells contribute to the unwounded corneal epithelium during normal homeostasis	mouse	[64]	++	0	0

---

++, expected for hypothesis; + consistent with hypothesis; ±, not consistent with hypothesis unless specific assumptions are made or technical issues compromise the interpretation of the experiment; 0, not consistent with hypothesis. <sup>a</sup>This is compatible with the LESC hypothesis if self-maintenance of the central corneal epithelium is a back-up mechanism that is used when homeostasis is compromised and LSCs are unable to maintain the corneal epithelium. <sup>b</sup>The chase period may not have been sufficient to identify label-retaining cells (see text). <sup>c</sup>The CESC hypothesis, as originally stated <sup>[1]</sup>, proposed that corneal epithelial cell movement was centrifugal but this assumption is not necessary. However, it requires *ad hoc* assumptions to account for centripetal movement across the full radius. <sup>d</sup>Evidence is circumstantial.



**Table 2: Examples of marker gene expression differences between the basal limbal and corneal epithelia during normal homeostasis identified by immunostaining**

Positive Markers			Negative Markers		
Marker	Species	References	Marker	Species	References
Integrin $\alpha 9$	mouse human	[3, 105, 106]	Keratin 3	rabbit	[4]
$\Delta Np63^*$	human	[3, 107, 108]	Keratin 3/Keratin 12	human	[3, 108]
$\Delta Np63\alpha$	human	[96]	NGF receptor (p75NTR)	human	[3]
ABCG2	human, rat, rabbit	[3, 57, 91-93, 108]	Involucrin	human	[3]
Vimentin	human	[108]	Connexin 43	human	[3, 108]
Keratin 19	human, mouse	[108, 109]	E-cadherin	human	[3]
Keratin 15	human, mouse	[109]	Nestin	human	[108]
N-cadherin	human	[110]			
Bmi1	human	[111]			
C/EBP $\delta$	human	[111]			
ABCB5	human, mouse	[54]			

Positive markers are expressed in basal limbal but not basal corneal epithelium. Negative markers are expressed in basal corneal but not basal limbal epithelium.

\* $\Delta Np63$  is not expressed in the human basal corneal epithelium <sup>[107]</sup> but it is expressed in mouse and rat corneal epithelia <sup>[27, 112, 113]</sup>.

## FIGURE LEGENDS

**Figure 1: Diagrammatic representation of the tissues of the mouse ocular surface and eyelid**

**Figure 2: Palisades of Vogt**

Diagram showing the arrangement of the palisades of Vogt (upward projections of limbal stroma into the limbal epithelium) in the human limbus in plan view (left) and how they might appear in differently orientated sections through the limbus (right). Only 16 palisades are shown but in reality there are many more. For anatomy see references [114, 115].

**Figure 3: LESC vs. CESC hypotheses**

(A) Diagram of human corneal epithelial maintenance according to the limbal epithelial stem cell (LESC) hypothesis showing active LESC in the limbal epithelium in both a limbal crypt and a limbal epithelial crypt. The LESC divide slowly replacing themselves and producing daughter transient (or transit) amplifying cells (TACs), which divide more quickly and move centripetally from the basal layer of the limbal epithelium to the basal layer of the corneal epithelium. After a final cell division TACs leave the basal layer, move through the suprabasal layers and are shed from the surface as terminally differentiated cells (TDCs). (B) Histological section showing mouse cornea, limbus and part of the conjunctiva immunohistochemically stained for keratin 12 (K12; dark brown staining) to show the border between the corneal epithelium (K12 positive) and limbal epithelium (K12 negative). (C) Drawing of photograph shown in (B) with different tissues labelled. The boxed area shows part of the limbal and corneal epithelia, equivalent to that represented in (D & E). (D) Diagram of mouse corneal epithelial maintenance according to the limbal epithelial stem cell (LESC) hypothesis. The principles are the same as described for (A). (E) Diagram of mouse corneal epithelial maintenance according to the corneal epithelial stem cell (CESC) hypothesis. The CESC divide slowly replacing themselves and producing daughter TACs, which divide more quickly and move centrifugally as originally proposed [1]. After a final cell division TACs leave the basal layer, move through the suprabasal layers and are shed from the surface. *Abbreviations:* cb, ciliary body; ce, corneal epithelium; cj, conjunctiva; cs, corneal stroma; ir, iris; le, limbal epithelium; re, retina; sc, sclera. Photograph (B) is reproduced from Mort et al [18] with kind permission of Springer Science+Business Media.

**Figure 4: Transition from randomly orientated patches to radial stripes in corneal epithelia of different types of mosaic mice between 3 weeks and adulthood.**

(A, B)  $\beta$ -gal staining in *XLacZ* X-inactivation mosaics [27]. (C, D) GFP fluorescence in CAG-GFP transgenic mosaics [77]. (E, F) GFP fluorescence in corneal epithelium after transfecting conceptuses with lentiviral vectors encoding green fluorescent protein (GFP) at embryonic day 9 or 10 [79]. Photographs (A & D) are reproduced from *Developmental Dynamics* [27] with kind permission of John Wiley and Sons, (C & D) are reproduced from *Molecular Vision* [77] with kind permission of the authors and editors, and photographs (E & F) are reproduced from *Molecular Therapy* [79] with kind permission of the authors and the Nature Publishing Group. This combination of photographs was first published by Mort et al [18].

**Figure 5: Hypothetical results from a lineage tracing experiment to distinguish between the LESC and CESC hypotheses**

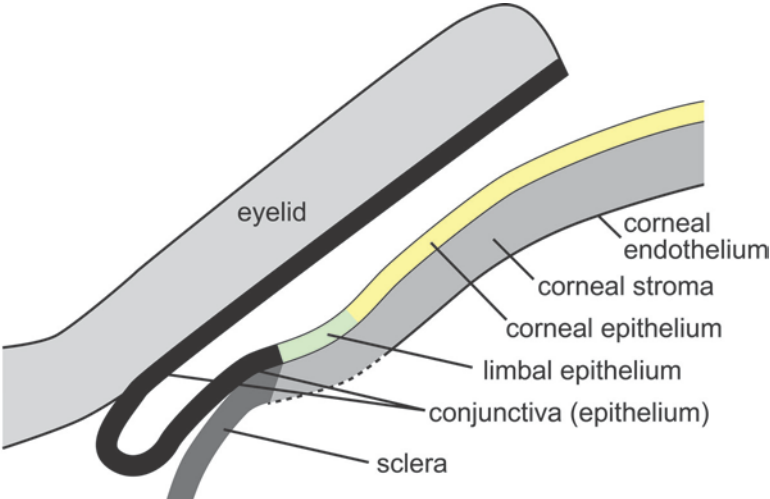
In each figure the inner disc represents the corneal epithelium and the outer ring represents the limbal epithelium. If a reporter transgene is driven by a tamoxifen-inducible, ubiquitous promoter, a proportion of all the cell types in the ocular surface (and other tissues) will be labelled shortly after tamoxifen treatment. The frequency of labelled cells will depend partly on the dose of tamoxifen, which could be titrated to ensure only a few stem cells are labelled per eye. Time 1 is shortly after tamoxifen-treatment and the labelled cells may have divided to produce a small clone of labelled cells. By Time 2, the short-lived labelled clones produced by labelling TACs should have been shed from the corneal epithelium but long-lived labelled clones produced by long-lived labelled stem cells will remain. Expectations for distributions of labelled cells at Times 2 and 3 vary for the different hypotheses. (A) The LESC hypothesis predicts clones of labelled cells will extend radially from the limbus and, by Time 3, clones of labelled cells will span the full radius. (B) The original CESC hypothesis predicts clones of labelled cells produced by labelled stem cells may also extend radially but will extend centrifugally from stem cells located throughout the corneal epithelium itself. Clones of labelled cells that do not arise from the centre of the cornea will not span the full radius. (C) If the CESC hypothesis is modified so that all the CESC are at the very centre of the cornea, centrifugal movement will produce clones of labelled cells that span the full radius by Time 3 but at Time 2 there should be no labelled peripheral cells produced by stem cells. (D) If the CESC hypothesis

is modified so that the CESC's are located throughout the corneal epithelium but movement is centripetal, clones of labelled cells that do not arise from the periphery of the cornea will not span the full radius. To distinguish between the various hypotheses it will be necessary to compare patterns of labelled clones at different times after tamoxifen treatment.

**Figure 6: Preliminary results from a lineage tracing experiment to distinguish between the LESC and CESC hypotheses**

Eyes from CAGG-CreER;R26R-*LacZ* mice that were injected with tamoxifen to induce *LacZ* reporter gene expression and stained for  $\beta$ -galactosidase ( $\beta$ -gal) activity after different chase periods. The pigmented iris is visible through the cornea and appears grey, whereas the  $\beta$ -gal staining is blue. (A) Side view of a  $\beta$ -gal-stained eye, after a chase period of 9 weeks and 4 days, with several radial  $\beta$ -gal-positive stripes and small patches in the cornea and numerous  $\beta$ -gal-positive patches in the conjunctiva. (The conjunctiva is torn near the limbus and hangs down at the bottom right of the photograph, so the sclera is visible between the limbus and conjunctiva.) Stripe 1 is a limbus-cornea (LC) stripe, with its more peripheral end in the limbus (arrow), consistent with expectations of the LESC hypothesis (see Fig. 5). It also appears to be aligned with other  $\beta$ -gal-positive patches towards the centre of the cornea so it may be part of a longer discontinuous stripe. Stripe 2 is a cornea-cornea (CC) stripe with both ends in the cornea, consistent with the CESC hypothesis. However, it is radially aligned with a small  $\beta$ -gal-positive patch in the limbus (arrow), which could be the location of a  $\beta$ -gal-positive LESC. If so, stripe 2 might be a discontinuous stripe, which extended from a LESC that was not continuously active (consistent with the LESC hypothesis). (B) Anterior (frontal) view of a  $\beta$ -gal-stained eye, after a 14-week chase period, with eight radial  $\beta$ -gal-positive stripes. All eight stripes are LC stripes with one end at the limbus and many extend the full radius and have a curved end, consistent with a central spiral pattern, as reported for other chimaeric and mosaic eyes [27, 72, 73]. CAGG-CreER;R26R-*LacZ* mice were produced by crossing CAGG-CreER and R26R-*LacZ* mice [full names Tg(CAG-cre/Esr1\*)5Amc and B6.129S4-*Gt(ROSA)26Sor<sup>tm1Sor</sup>/J*; references [116, 117].] *LacZ* reporter gene expression was induced at 12 weeks by 3 injections of tamoxifen (100  $\mu$ g /g body weight per injection). Scale bar = 0.5 mm.

**FIGURES**



**Figure 1.**

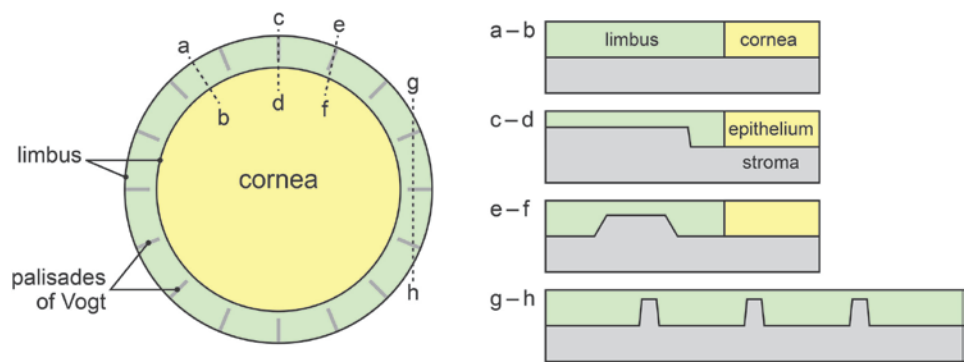
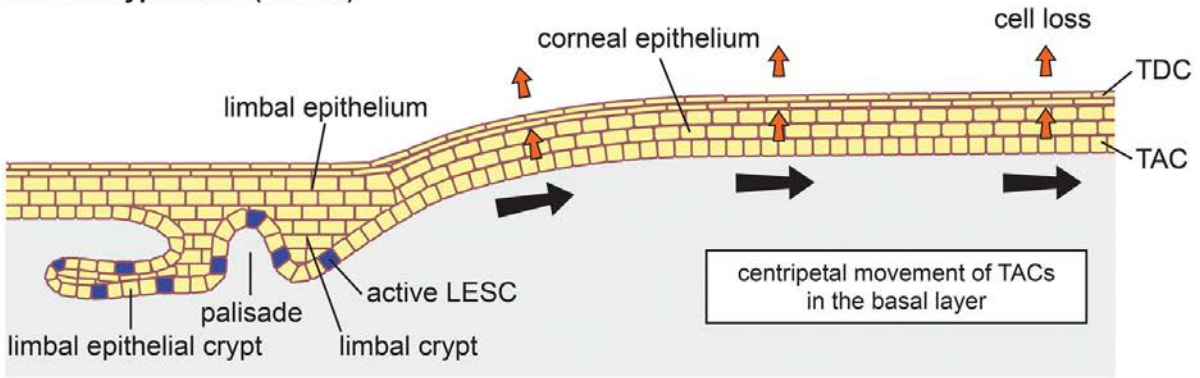


Figure 2.

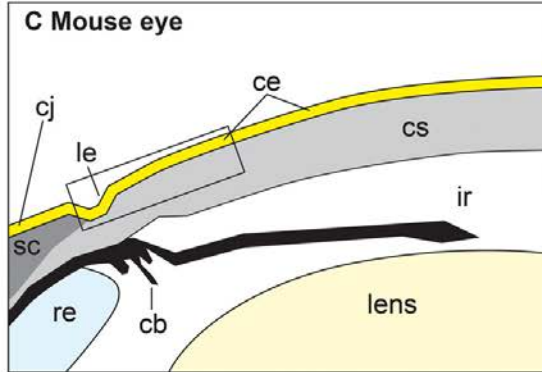
**A LESC hypothesis (human)**



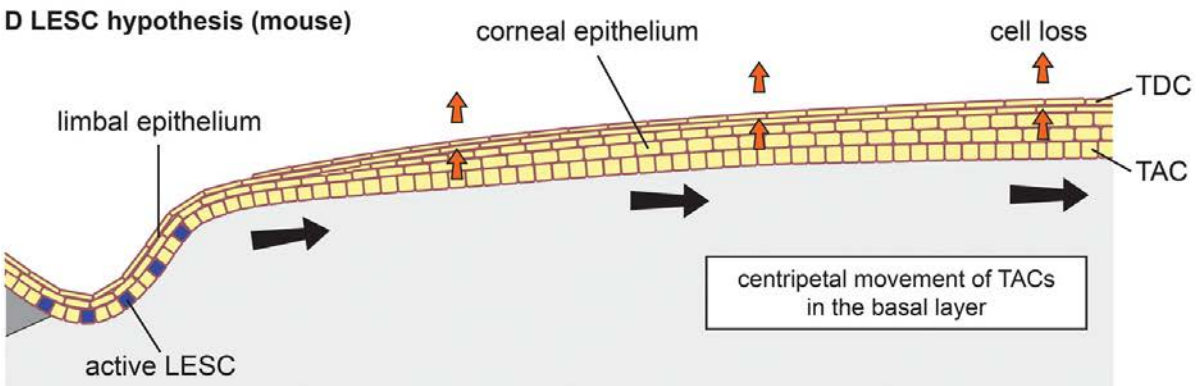
**B Mouse eye**



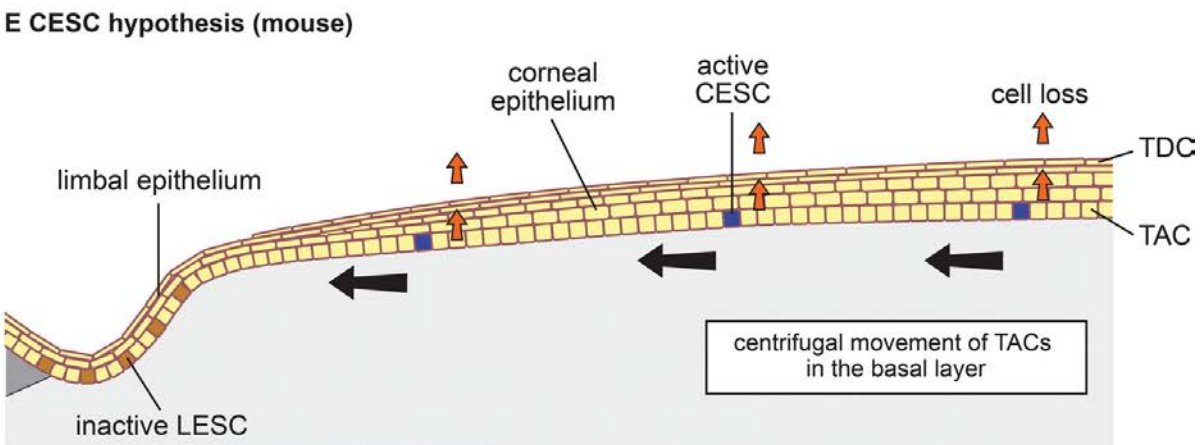
**C Mouse eye**



**D LESC hypothesis (mouse)**



**E CESC hypothesis (mouse)**



**Figure 3.**

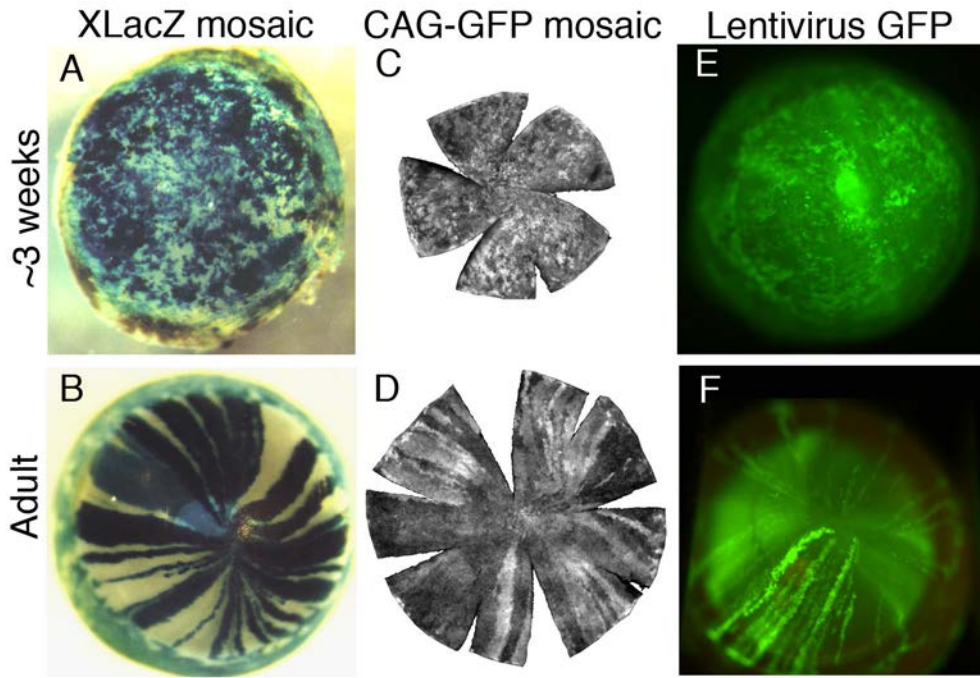
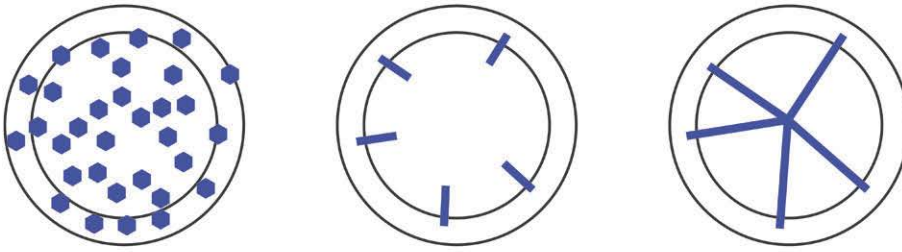


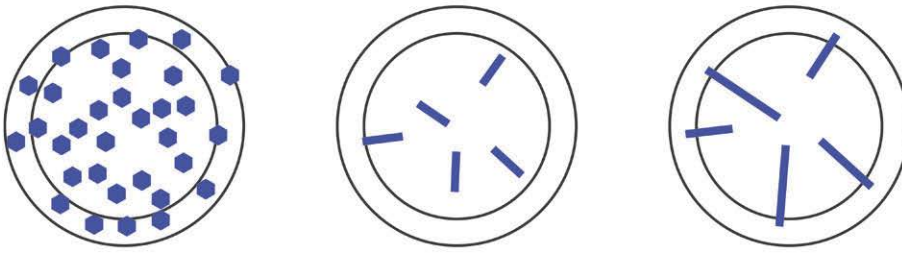
Figure 4.



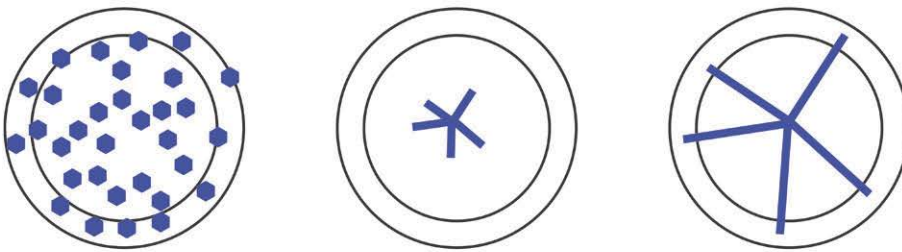
A LESC hypothesis (centripetal movement)



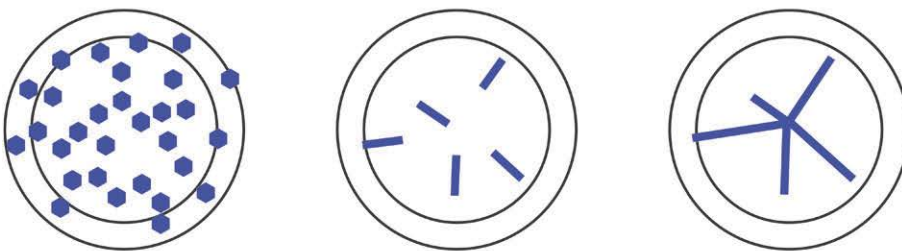
B Original CESC hypothesis (centrifugal movement)



C Central CESC and centrifugal movement



D CESC throughout cornea and centripetal movement



Time 1

Time 2

Time 3

Figure 5.

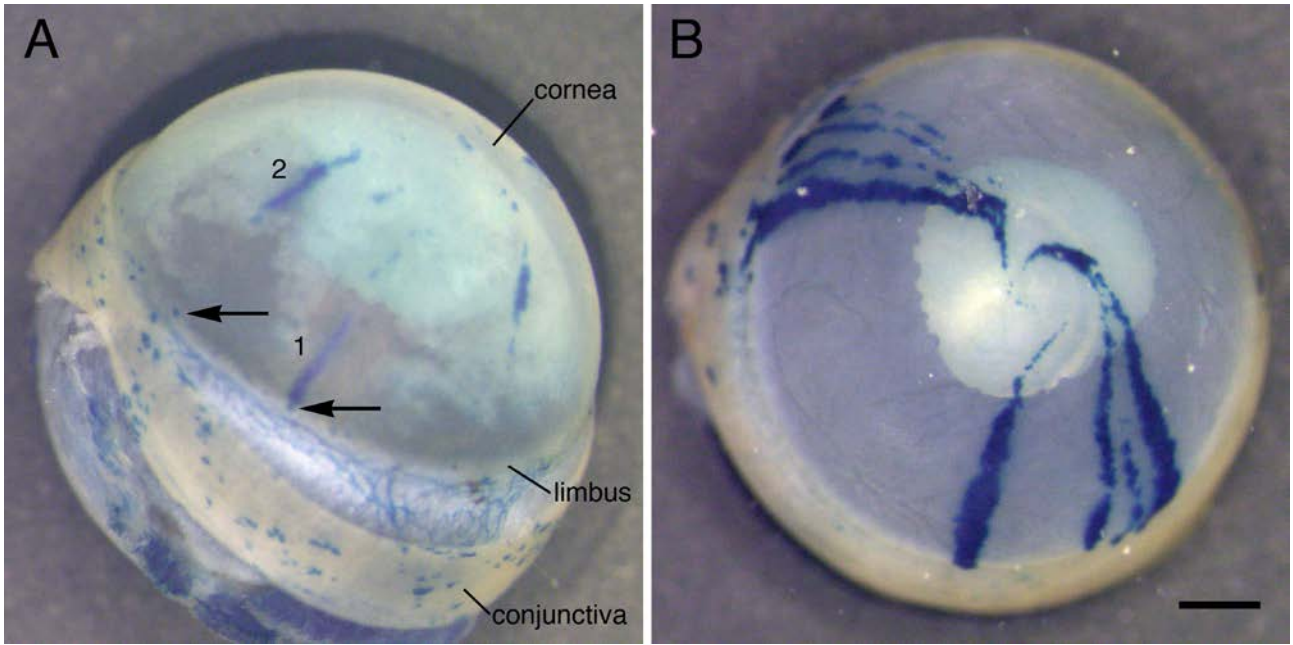


Figure 6.