Rings of Charge within the Extracellular Vestibule Influence Ion Permeation of the 5-HT$_3$A Receptor*

Matthew R. Livesey$^{1,2}$, Michelle A. Cooper, Jeremy J. Lambert$^1$, and John A. Peters$^{1,3}$

From the Centre for Neuroscience, Division of Medical Sciences, Ninewells Hospital and Medical School, University of Dundee, Dundee DD1 9SY, United Kingdom

The determinants of single channel conductance ($\gamma$) and ion selectivity within eukaryotic pentameric ligand-gated ion channels have traditionally been ascribed to amino acid residues within the second transmembrane domain and flanking sequences of their component subunits. However, recent evidence suggests that $\gamma$ is additionally controlled by residues within the intracellular and extracellular domains. We examined the influence of two anionic residues (Asp$^{113}$ and Asp$^{127}$) within the extracellular vestibule of a high conductance human mutant 5-hydroxytryptamine type-3A (5-HT$_3$A) receptor (5-HT$_3$A(QDA)) upon $\gamma$, modulation of the latter by extracellular Ca$^{2+}$, and the permeability of Ca$^{2+}$ with respect to Cs$^+$ ($P_{\text{Ca}}/P_{\text{Cs}}$). Mutations neutralizing (Asp $\rightarrow$ Asn), or reversing (Asp $\rightarrow$ Lys), charge at the 113 locus decreased inward $\gamma$ by 46 and 58%, respectively, but outward currents were unaffected. The D127N mutation decreased inward $\gamma$ by 82% and also suppressed outward currents, whereas the D127K mutation caused loss of observable single channel currents. The forgoing mutations, except for D127K, which could not be evaluated, ameliorated suppression of inwardly directed single channel currents by extracellular Ca$^{2+}$. The $P_{\text{Ca}}/P_{\text{Cs}}$ of 3.8 previously reported for the 5-HT$_3$A(QDA) construct was reduced to 0.13 and 0.06 by the D127N and D127K mutations, respectively, but clearer significant, effects caused by the D113N (1.04) and D113K (0.60) substitutions. Charge selectivity between monovalent cations and anions ($P_{\text{Na}}/P_{\text{Cs}}$) was unaffected by any of the mutations examined. The data identify two key residues in the extracellular vestibule of the 5-HT$_3$A receptor that markedly influence $\gamma$, $P_{\text{Ca}}/P_{\text{Cs}}$, and additionally the suppression of $\gamma$ by Ca$^{2+}$.

The pentameric ligand-gated ion channel (pLGIC)$^4$ superfamily in eukaryotes can be subdivided into two functional groups based upon the charge selectivity of the ion channel in the open state. The excitatory 5-hydroxytryptamine type 3 (5-HT$_3$) and nicotinic acetylcholine (nACh) receptors, together with the zinc-activated channel, are non-selective cation channels that are permeable to Na$^+$, K$^+$, and, to varying degrees, Ca$^{2+}$ and Mg$^{2+}$ (1–3). The inhibitory $\gamma$-aminobutyric acid type A (GABA$_A$) and glycine receptors are anion-selective and conduct mainly Cl$^-$ and HCO$_3^-$ under physiological conditions (4–6).

All eukaryotic pLGIC subunits possess three topologically and functionally homologous domains (7–11): (i) an extracellular domain (ECD), (ii) the transmembrane domain, and (iii) an intracellular domain. The ECD is rich in $\beta$-sheet and loop motifs and contributes to the ligand binding site, which is located at subunit interfaces, and the extracellular vestibule of the ion channel. The transmembrane domain comprises four $\alpha$-helical stretches (TM1–TM4) organized such that TM1, TM3, and TM4 shield TM2 from the lipid bilayer (8, 9). The TM regions are linked by extracellular (TM2-TM3) and intracellular (TM1-TM2 and TM3-TM4) loops. TM2 forms the lining of the transmembrane portion of the ion channel and contains the channel gate (9). The TM3-TM4 linker (the large intracellular loop) forms the bulk of the intracellular domain and is the least conserved region of the subunit peptide across the pLGIC receptor family. The structure of the intracellular domain is largely unresolved, apart from an $\alpha$-helical region (the MA stretch) that is located immediately N-terminal to TM4 in images of the Torpedo nACh receptor obtained by cryo-electron microscopy (cryo-EM) (9, 12).

It is axiomatic that residues located within TM2 and the TM1-TM2 linker influence profoundly (i) single channel conductance ($\gamma$), (ii) charge selectivity, and (iii) for cation selective channels, di- versus monovalent cation selectivity (13–15) (reviewed in Ref. 4). However, several recent studies have revealed that regions homologous to the MA stretch of the Torpedo nACh receptor are additional determinants of $\gamma$ in 5-HT$_3$, nACh, A$\alpha$B$\delta$, and $\alpha$ glycine receptors (16–19) (reviewed in Ref. 1). In the case of the 5-HT$_3$A receptor, this region also contributes to di- versus monovalent cation selectivity but not charge selectivity (20). The 4 Å resolution structure of the Torpedo nACh receptor obtained by cryo-EM and image reconstruction (8, 9) reveals that the MA stretch, with a contribution from the TM1-TM2 linker, frames intracellular fenestrations through which ions must flow to enter or exit the cytoplasmic vestibule of the channel, providing a structural basis for such observations.
The ECD of the 5-HT₃A Receptor Influences Channel Biophysics

Adding to the concept of an extended permeation pathway that influences the biophysical properties of pLGICs, the wide extracellular vestibule, surrounded by the pore lining regions of the ECD, has long been theorized to localize cations to the narrower transmembrane channel (21). Indeed, the cryo-EM images of the Torpedo nACh receptor depict an electronegative vestibule of ~60 Å in length and maximally 20 Å in width that has been predicted to strongly influence cation conduction (9).

Using a homology model of the closed state of the adult muscle nACh receptor ([α1]₂β₁γε), based upon the cryo-EM images of the Torpedo nACh receptor (9), Wang et al. (22) conducted in silico molecular dynamics simulations of the transition of Na⁺ ions through the conduction pathway over a time frame of 16 ns. The simulation revealed the stabilization of cations at a number of positions along the channel axis. Notably, the extracellular vestibule was highlighted to present negative charges corresponding to the α1 subunit residues Glu⁸³ and Asp⁹⁷, where cations appeared to reside before further translocation to the transmembrane portion of the channel. Numerous additional computational approaches applied to homology models of the nACh receptor indicate that the ECD acts to concentrate monovalent cations over anions (e.g. see Refs. 23 and 24). By contrast, a homology model of the GABA₂α receptor indicates the ECD to concentrate anions in preference to cations (25).

Direct experimental evidence for a role of the extracellular vestibule of human muscle nACh receptors in setting γ has been provided by Hansen et al. (26), who made mutations within a negatively charged ring corresponding to Asp⁹⁷ in the α1 subunit. Mutation of the residues at this locus to lysine in all subunits caused a maximal reduction of γ to ~20% of control, as observed in recordings performed on cell-attached patches.

In this study, we investigate the influence of Asp¹¹³ and Asp¹²⁷ of the 5-HT₃A receptor, which align with Glu⁸³ and Asp⁹⁷ of the α1 nACh receptor subunit, respectively, upon γ, single current channel rectification, ion selectivity, and modulation of γ by extracellular Ca²⁺. The analyses were performed in the background of a high γ mutant of the human 5-HT₃A receptor (5-HT₃A(QDA)), because the wild-type 5-HT₃A receptor has γ value too small to quantify by conventional single channel recording techniques (1, 16, 27). The mutations introduced into the large intracellular loop of the human 5-HT₃A receptor to generate the 5-HT₃A(QDA) receptor construct are R432Q, R436D, and R440A (16). We demonstrate that the extracellular channel vestibule within this model cationic pLGIC strongly influences γ and is also an important determinant of relative permeability to Ca²⁺ and the modulation of γ by extracellular Ca²⁺. A preliminary account of some of the results has appeared in abstract form (28).

EXPERIMENTAL PROCEDURES

5-HT₃A Receptor Constructs and Transfection of cDNAs—The methods employed to generate mutant constructs within the 5-HT₃A(QDA) cDNA background and to transiently transf ect tsA-201 cells with cDNA constructs were as detailed previously (20). The mutations generated in this study were 5-HT3A(QDA) D113N, 5-HT3A(QDA) D113K, 5-HT3A(QDA) D127N, and 5-HT3A(QDA) D127K. Cells were maintained in Dulbecco’s modified Eagle’s medium supplemented with 10% (v/v) fetal bovine serum, 2 mM L-glutamine, 1 mM sodium pyruvate, 100 μg/ml streptomycin, and 100 units/ml penicillin. Cells were incubated at 37 °C for 18–96 h in 95% air, 5% CO₂, at 100% humidity prior to electrophysiological recordings.

Electrophysiological Recordings—The whole-cell and outside-out patch configurations of the patch clamp technique were used to record macroscopic and single channel currents evoked by 5-HT, respectively. Patch electrodes for both modes were filled with a solution comprising 140 mM CsCl, 0.1 mM CaCl₂, 1.1 mM EGTA, 10 mM HEPES, pH 7.2 (adjusted with 1 mM CsOH; final [Cs⁺]ᵢ = 143 mM). The intracellular free calcium concentration of this solution was estimated to be 10 nM (29). The recording chamber comprised a 35-mm Petri dish on which the cells were cultivated. Using a gravity-feed system, the chamber was superfused routinely at ~5 ml/min, at room temperature (20–23 °C), with extracellular solutions of the compositions described below. Single channel currents evoked by pressure-applied 5-HT (10 μM) were recorded in an extracellular solution (E1) consisting of 142.8 mM NaCl, 0.1 mM MgCl₂, 0.1 mM CaCl₂, 10 mM glucose, 10 mM HEPES, pH 7.2 (adjusted by 1 M NaOH; final [Na⁺]ᵢ = 146 mM). Whole-cell recording was used to determine the reversal potentials (E₅-HT) of macroscopic currents elicited by 5-HT (10 μM) by means of a voltage-ramp protocol using WinWCP V3.9.6 electrophysiology software (J. Dempster, Department of Physiology and Pharmacology, University of Strathclyde, UK; available on the World Wide Web). Unless stated otherwise, E₅-HT was determined via a protocol wherein the membrane potential was initially set at ~60 mV, and a macroscopic current response to 5-HT was elicited. During the plateau of the current response, the membrane potential was stepped from ~60 to ~100 mV for 100 ms and subsequently ramped to +80 mV within 1 s. Care was taken to ensure that the current amplitude at the beginning and termination of the voltage ramp protocol were similar. An identical voltage ramp in the absence of 5-HT served to determine the leakage current and was subtracted from the currents evoked in the presence of 5-HT to generate the current-voltage (I-V) relationship specifically attributable to the 5-HT-evoked conductance. E₅-HT was determined from such leak-subtracted currents. In order to determine the permeability of Na⁺ relative to Ca²⁺ (P₅₆/P₅₋₉), E₅-HT was determined in solution E1, and the permeability ratio was calculated from the Goldman-Hodgkin-Katz (GHK) (voltage) equation,

\[ E_{5-HT} = \frac{RT}{F\ln\left(\frac{P_{Na}/P_{Ca}[Na^+]_i}{[Cs^+]_i}\right)} \]  

where R, T, and F have their usual meaning, and [Na⁺]ᵢ and [Cs⁺]ᵢ are the calculated activities of extracellular Na⁺ and internal Cs⁺ ions. This equation ignores the very small error anticipated due to the presence of low concentrations of permeant divalent ions (e.g. Ca²⁺) within the extra- or intracellular solutions.

To calculate the permeability of Ca²⁺ relative to Cs⁺ (P₅₋₉/P₅₋₀), E₅-HT was determined in solution E2, comprising CaCl₂...
The ECD of the 5-HT3A Receptor Influences Channel Biophysics

100 mM, glucose 10 mM, l-histidine 5 mM (pH 7.2), and the ratio was derived from a modified GHK (voltage) equation (20, 30),

\[ E_{5-HT} = \frac{RT}{F} \ln \left( \frac{P_{Ca}/P_{Cl}}{[Ca^{2+}]o} \right) \]  

(Eq. 2)

where \([Ca^{2+}]o\) is the external activity of Ca^{2+}, \([Cl^-]o\) is the internal activity of Cl^-, and \(P_{Ca}/P_{Cl}\) is a modified term relating the permeability of Ca^{2+} to Cl^- . Substituting for \(P_{Ca}/P_{Cl}\), the above can be rewritten as follows.

\[ \exp(E_{junc}/RT) = \frac{P_{Ca}[Ca^{2+}]o}{P_{Cl}(1 + \exp(E_{junc}/RT))[Cl^-]o} \]  

(Eq. 3)

For simplicity, we routinely refer to ion concentrations rather than activities throughout, although the latter were employed in the calculations of relative permeabilities.

To examine modulation of \(\gamma\) by extracellular Ca^{2+}, single channel recordings and determinations of \(E_{5-HT}\) were also made in extracellular solutions (E3, E4, and E5) containing variable \([CaCl_2]o\) (0.1, 1, and 10 mM, respectively) but constant \([NaCl]o\) (95 mM), l-histidine (5 mM), and glucose 10 (mM), pH 7.2. The osmolality of such solutions was held constant by the addition of appropriate amounts of sucrose. An extracellular solution (E6) containing 20 mM NaCl, 0.1 mM MgCl_2, 0.1 mM CaCl_2, 10 mM glucose, 239.4 mM sucrose, 5 mM l-histidine, pH 7.2 (adjusted by 1 M HCl), in addition to solutions E1 and E3, was used to perform dilution potential measurements in order to determine relative permeability to chloride. Liquid junction potentials between the pipette tip and extracellular solutions were measured according to Fenwick et al. (29) and were corrected post hoc. In brief, the zero current voltage was initially determined with the intracellular solution present within both the patch pipette and the bath. The extracellular solutions (E1–E6) used in the experiments then replaced the intracellular solution, and the change in zero current voltage (i.e. the liquid junction potential) was noted. A salt bridge consisting of 3 M KCl in 4% (w/v) agar connected the bath to the reference electrode, eliminating changes in the potential of the latter during this procedure.

Single channel currents were low pass-filtered offline at 1 kHz, digitized at 10 kHz via a DigiData 1302A (Axon Instruments) interface. Using the WinEDR V2 7.6 Electrophysiology Data Recorder (J. Dempster, Department of Physiology and Pharmacology, University of Strathclyde, UK; available on the World Wide Web), single channel current amplitude histograms were constructed from sections of single channel activity in which unitary events predominated, as described in detail previously (20). The \(\gamma\) value is routinely reported as the chord conductance (i.e. \(\gamma = i(V_{m} - E_{5-HT})\)), where \(i\) is single channel current amplitude, \(V_{m}\) is the holding potential (including liquid junction potential correction), and \(E_{5-HT}\) is the reversal potential of the agonist-evoked macroscopic response determined as described above under the appropriate ionic conditions. In the case of the 5-HT3A(QDA) D127K mutant, single channel currents could not be resolved directly in recording from outside-out membrane patches. Thus, we estimated \(\gamma\) by fluctuation analysis of whole-cell current evoked by 5-HT (1 \muM) as previously described by us (16). A rectification index (RI) is used to describe the degree of single channel current rectification. The RI was calculated as follows,

\[ RI = \frac{\text{slope}_{\gamma_{\text{positive V}}}}{\text{slope}_{\gamma_{\text{negative V}}}} \]  

(Eq. 4)

where slope \(\gamma_{\text{positive V}}\) and slope \(\gamma_{\text{negative V}}\) were obtained from obtaining the slope of the line of best fit plotted to data points from positive and negative holding potentials, respectively. An RI value of 1 indicates a linear/ohmic \(i-V\) relationship. An RI of \(>1\) indicates outward rectification, and RI \(<1\) indicates inward rectification.

Statistical Analysis—Data are presented as mean ± S.E. Statistical analysis was conducted using one-way analysis of variance (ANOVA) with the post hoc Dunnett’s or Dunn’s test as appropriate. \(p < 0.05\) was considered to be significant.

RESULTS

Asp^{113} Is a Determinant of \(\gamma\) and Single Channel Current Rectification—Asp^{113} is predicted from a homology model of the human 5-HT3A receptor (18) to reside approximately one-third of the distance from the entrance to the extracellular vestibule to the extracellular border of TM2. Asp^{113} is homologous to the Torpedo nACh receptor \(\alpha 1\) subunit Glu^{83} locus and is modeled as forming a ring of negative charge within the apical extracellular vestibule (Fig. 1). Asp^{113} was mutated to either a neutral asparagine or a positively charged lysine, within the 5-HT3A(QDA) construct, yielding 5-HT3A(QDA) D113N or 5-HT3A(QDA) D113K, respectively. At a holding potential of −80 mV in extracellular (E1) and intracellular solutions containing Na^+ and Cs^- as the principal charge carriers respectively, 5-HT (10 \muM) applied by pressure to outside-out patches excised from tsA-201 cells expressing 5-HT3A(QDA) D113N
The ECD of the $5$-HT$_3$A Receptor Influences Channel Biophysics

The influence of mutations within the extracellular domain (ECD) upon $\gamma$ and single channel current rectification

Single channel currents evoked by 5-HT (10 $\mu M$) were recorded from outside-out patches excised from tsA-201 cells transfected with the construct of interest. The Cs$^+$-based pipette solution and extracellular solution E1 were utilized. A, example single channel events recorded at $-80$ and $+80$ mV from the 5-HT$_A$(QDA), 5-HT$_A$(QDA) D113N, and 5-HT$_A$(QDA) D113K mutant receptors. B, i-V profiles for the 5-HT$_A$(QDA) (solid circles), 5-HT$_A$(QDA) D113N (open circles), and 5-HT$_A$(QDA) D113K (inverted triangles) receptors. Note that mutation of the Asp$^{113}$ residue does not impact upon outwardly directed single channel currents. Data points are the mean of a minimum of three observations made from separate patches, and error bars, where visible, indicate S.E. Data for the 5-HT$_A$(QDA) receptor are from Livesey et al. (20).

**TABLE 1**
The influence of mutations within the extracellular domain (ECD) upon $\gamma$ and single channel current rectification

<table>
<thead>
<tr>
<th>Receptor constructs</th>
<th>$\gamma$</th>
<th>$n$</th>
<th>Rectification index</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-HT$_A$(QDA)</td>
<td>41.2 $\pm$ 1.2$^a$</td>
<td>8</td>
<td>1.12</td>
</tr>
<tr>
<td>5-HT$_A$(QDA) D113N</td>
<td>22.2 $\pm$ 0.3$^b$</td>
<td>9</td>
<td>1.7</td>
</tr>
<tr>
<td>5-HT$_A$(QDA) D113K</td>
<td>17.1 $\pm$ 0.2$^c$</td>
<td>8</td>
<td>2.63</td>
</tr>
<tr>
<td>5-HT$_A$(QDA) D127K</td>
<td>7.4 $\pm$ 0.1$^d$</td>
<td>3</td>
<td>3.18</td>
</tr>
<tr>
<td>5-HT$_A$(QDA) D127K</td>
<td>1.6 $\pm$ 0.3$^e$</td>
<td>6</td>
<td>ND</td>
</tr>
</tbody>
</table>

*a Data reported previously by Livesey et al. (20).
*b Significantly different $(p < 0.001)$ from the 5-HT$_A$(QDA) mutant receptor, as determined by one-way ANOVA with post hoc Dunnett’s test.
+c Significantly different $(p < 0.001)$ from the 5-HT$_A$(QDA) D113N mutant receptor construct, as determined by unpaired $t$ test.

Mutant receptors elicited clearly resolvable single channel events. By comparison with the 5-HT$_A$(QDA) receptor construct previously characterized under identical ionic conditions (20), inwardly directed single channel currents mediated by the 5-HT$_3$A(QDA) D113N construct are effectively halved in amplitude, the chord conductance ($\gamma$) being reduced from 41.2 pS to 22.2 $\pm$ 0.3 pS (Fig. 2, A and B, and Table 1). The 5-HT$_3$A(QDA) D113K receptor displayed a $\gamma$ value that was further reduced to 17.1 $\pm$ 0.2 pS (Fig. 2, A and B, and Table 1). Although the $\gamma$ value of the D113N and D113K 5-HT$_3$A(QDA) constructs did not differ strikingly, the reduction caused by reversal, rather than neutralization, of charge was significantly greater (Table 1).

Single channel current-voltage ($i$-$V$) relationships were generated by recording unitary currents evoked by 5-HT (10 $\mu M$) from outside-out patches at holding potentials between $-100$ and $+100$ mV in increments of 20 mV. For the 5-HT$_3$A(QDA) receptor construct, the single channel current $i$-$V$ relationship is linear at negative potentials with a slope $\gamma$ of 41.2 pS (20). However, at positive potentials, $\gamma$ increases to 46 pS, indicative of modest outward rectification quantified as an RI value (see “Experimental Procedures”) of 1.12 (20) (Fig. 2B). Under identical ionic conditions, the $i$-$V$ relationships obtained for the 5-HT$_3$A(QDA) D113N and the 5-HT$_3$A(QDA) D113K receptor constructs reveal that both mutations enhance outward rectification, the RI values being 1.7 and 2.63, respectively (Table 1).

Notably, either mutation reduced only inwardly (and not outwardly) directed single channel currents (Fig. 2B). Such results are consistent with a scheme whereby the ring of negativity formed by the Asp$^{113}$ residues acts via simple coulombic attraction to raise the availability of cations within the receptor extracellular vestibule for translocation to deeper regions of the pore. Neutralization or reversal of charge at this locus would be anticipated to reduce the local concentration of cations within the extracellular vestibule, leading to the emergence of outward rectification. Notably, the effect of neutralizing or reversing the charge at the Asp$^{113}$ position was increased when [Na$^+$], was reduced to 95 mM (see below) (see Fig. 5). In comparison with the 46 and 58% reductions in $\gamma$ found for the D113N and D113K mutations, respectively, with [Na$^+$], set at 146 mM, the corresponding reductions in the Na$^+$-deficient solution were 66 and 72%. The enhanced effect of the mutations at the lower ionic strength is once more consistent with a predominantly electrostatic influence of the Asp$^{113}$ residue. This is due to the screening effect of oppositely charged ions upon amino acid residues that have fixed charges that would reduce the influence of the Asp$^{113}$ residue at high ionic strength.

Asp$^{277}$ Is a Major Determinant of $\gamma$ and Influences Single Channel Current Rectification—Asp$^{277}$ is predicted from a homology model of the human 5-HT$_3$A receptor (18) to reside approximately two-thirds of the distance from the entrance to the extracellular vestibule to the extracellular border of TM2 (i.e. the 20’ residue) and occupies a locus homologous to that of the Asp$^{97}$ residue of the Torpedo nACh receptor a1 subunit (Fig. 1). Application of 5-HT (10 $\mu M$) to outside-out patches expressing the human 5-HT$_3$A(QDA) D127N receptor construct yielded relatively small, yet clearly resolvable, single channel currents at a holding potential of $-80$ mV, yielding a chord $\gamma$ of 7.3 $\pm$ 0.1 pS (Fig. 3, A and B, and Table 1). Neutralization of the negative charge of the 127 residues thus reduces $\gamma$ by 82% in comparison with the 5-HT$_3$A(QDA) receptor. Consistent with a contribution of an electrostatic influence upon inwardly directed cation flux, the D127N mutation within the 5-HT$_3$A(QDA) construct confers a strong outwardly rectifying (RI = 3.18) $i$-$V$ profile, as determined from single channel currents recorded between the holding potentials of $-100$ and
The ECD of the 5-HT₃A Receptor Influences Channel Biophysics

FIGURE 3. The 5-HT₃A(QDA) D127N construct single channel i-V relationship. Single channel current events were recorded from excised outside-out patches in extracellular solution E1 and the Cs⁺-based pipette solution. A, exemplar currents recorded at ~80 mV from the 5-HT₃A(QDA), 5-HT₃A(QDA) D127N, and 5-HT₃A(QDA) D127K constructs. Note that the latter construct does not support resolvable single channel events in response to 5-HT. B, single channel i-V relationships for the 5-HT₃A(QDA) construct (filled squares) and the 5-HT₃A(QDA) D127N mutant receptor (open circles). Note that the 5-HT₃A(QDA) D127N receptor mediated single channel currents that rectify outwardly but are additionally substantially reduced at positive holding potentials. Data points are the mean of at least three independent determinations, and error bars, where visible, indicate S.E. Data for the 5-HT₃A(QDA) receptor are from Livesey et al. (20).

+100 mV at 20-mV increments (Fig. 3B). However, unlike mutations at the 113 locus, the 5-HT₃A(QDA) D127N receptor mediated outwardly directed single channel currents that were clearly depressed in comparison with the 5-HT₃A(QDA) construct (Fig. 3B). In addition, the effect of the D127N mutation was not enhanced when [Na⁺]₀ was reduced to 95 mM. It is likely that the role of Asp¹²⁷ extends beyond that of locally concentrating cations within the extracellular vestibule.

In contrast to the 5-HT₃A(QDA) D127N receptor, individual single channel events in response to 5-HT (10 μM) were not discernible when recording at a holding potential of ~80 mV from membrane patches expressing the 5-HT₃A(QDA) D127K construct (Fig. 3A). Instead, a noisy “macroscopic-like” inward current evoked by 5-HT was observed, which is reminiscent of that previously reported for the human wild-type 5-HT₃A receptor in a similar experimental paradigm (31). To estimate the γ value of the channels underlying such currents, we employed fluctuation analysis of 5-HT-evoked macroscopic currents from whole cells voltage-clamped at a holding potential of ~80 mV, as described previously (16, 27). Such an analysis yielded a γ of 1.65 ± 0.3 pS (Table 1). Outwardly directed single channel events mediated by the 5-HT₃A(QDA) D127K receptor construct in outside-out patches at holding potentials as positive as +80 mV were also unmeasurable by direct observation (data not shown). We were unable to obtain an estimate of γ by fluctuation analysis of whole-cell currents at such positive potentials due to a poor signal/noise ratio. Thus, the 127 locus appears to be a major determinant of both inward and outward conduction of cations.

The Influence of the 5-HT₃A Receptor Extracellular Vestibule upon Ion Selectivity—The suppression of the γ of the 5-HT₃A(QDA) receptor by the mutations at the 113 and 127 loci suggests that the residues naturally occupying these positions participate in the concentration of cations within the extracellular vestibule. In addition, residues within the 5-HT₃A receptor cytoplasmic portals, also distant from the TM2 region, have been shown to influence ion permeability, namely to Ca²⁺ (20). Thus, we sought to examine whether mutations in the extracellular vestibule influence the permeabilities of Ca²⁺ with respect to Cs⁺ (P_Ca/P_Cs), Na⁺ with respect to Cs⁺ (P_Na/Ca), and Na⁺ with respect to Cl⁻ (P_Na/Cl).

Asp¹¹³ and Asp¹²⁷ Affect neither Cation versus Anion Selectivity nor Relative Permeability to Monovalent Cations—Dilution potential experiments were performed in order to determine P_Na/P_Ca. In such experiments, Na⁺ replaced Cs⁺ as the permeant monovalent cation that dialyzed the cell interior. A whole-cell voltage ramp protocol (see “Experimental Procedures”) was employed to obtain E₅₋₅-HT values for the extracellular domain mutants in the presence of variable [Na⁺]₀ (146, 95, and 20 mM; solutions E1, E3, and E6, respectively), where Na⁺ was replaced with sucrose to maintain osmolarity. Plots of E₅₋₅-HT versus the logarithm of extracellular Na⁺ activity ([Na⁺]₀) were generated to examine the cation versus anion selectivity of each of the extracellular domain mutants. No appreciable perturbation of charge selectivity was observed in comparison with the 5-HT₃A(QDA) receptor (slope of 59 mV/decade change in [Na⁺]₀, 20) because the slopes were found to be 55.6, 59.5, 57.4, and 57.5 mV/decade change in (Na⁺)₀ for the 5-HT₃A(QDA) D113N, 5-HT₃A(QDA) D113K, 5-HT₃A(QDA) D127N, and 5-HT₃A(QDA) D127K receptors, respectively (data not shown). The GHK voltage equation predicts a slope of 58 mV/decade change in (Na⁺)₀ (at room temperature) for a channel that is perfectly selective for monovalent cations.

P_Na/P_Cs determined similarly was generally unaffected at 5-HT₃A(QDA) constructs carrying mutations to the Asp¹¹³ and Asp¹²⁷ residues, although the 5-HT₃A(QDA) D113N (0.78 ± 0.02) and 5-HT₃A(QDA) D127K (1.02 ± 0.03) constructs exhibited modestly shifted, but significantly different, ratios compared with the 5-HT₃A(QDA) construct (0.87 ± 0.03). However, the sign of the charge at the 113 and 127 loci had no consistent effect upon P_Na/P_Cs (Table 2).

Asp¹¹³ and Asp¹²⁷ Are Determinants of P_Ca/P_Cs—The preceding data suggest that Asp¹¹³ and Asp¹²⁷ are involved in the selection of cations within the extracellular vestibule, and they would be anticipated to favor the attraction of Ca²⁺ over monovalent cations, potentially increasing the relative permeability...
of the channel to the divalent. To evaluate this possibility, \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) values were derived from measurements of \( E_{55\text{-HT}} \) made from whole-cell currents activated during a voltage ramp by 5-HT (10 \( \mu M \)) in the presence of extracellular solution E2, containing 100 mM [Ca\(^{2+}\)]_o as the sole cationic charge carrier, and the Cs\(^+\)-based intracellular solution. The 5-HT\(_3\)A(QDA) D113N and 5-HT\(_3\)A(QDA) D113K receptors yielded \( E_{55\text{-HT}} \) values of \(-11.7 \pm 1.1\) and \(-22.7 \pm 1.8\) mV, which, when compared with the 5-HT3A(QDA) construct (+9.4 mV) (20), indicates a substantially decreased relative permeability to Ca\(^{2+}\) (Fig. 4A–C). Quantitatively, the corresponding \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) ratios calculated from a modified form of the GHK voltage equation (30) were 1.04 \( \pm \) 0.08 and 0.60 \( \pm \) 0.06 (Table 2), both significantly less than the value of 3.8 found for the 5-HT3A(QDA) receptor construct (20).

Pronounced negative shifts in \( E_{55\text{-HT}} \) were found for the 5-HT3A(QDA) Asp\(_{127}^{	ext{mut}}\) mutant constructs in the Ca\(^{2+}\)-based extracellular solution E2. As such, modifications to the conventional voltage-ramp protocol employed in this study were required. In brief, \( E_{55\text{-HT}} \) for the 5-HT3A(QDA) D127N construct was determined by ramping the membrane potential over a period of 1 s from +40 to -100 mV and returning to +40 mV during the peak of a macroscopic current evoked by 5-HT (10 \( \mu M \)). The resultant leak-subtracted current exhibited a large negative shift in \( E_{55\text{-HT}} \) (-55.6 \( \pm \) 1.2 mV) compared with that of the 5-HT3A(QDA) construct (+9.4 mV), yielding a significant reduction of \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) from 3.8 to 0.13 \( \pm \) 0.01 (Table 2 and Fig. 4D). For the 5-HT3A(QDA) D127K mutant, a similar voltage ramp protocol, but spanning the potential range -40 to -150 mV, yielded leak-subtracted whole-cell currents with an \( E_{55\text{-HT}} \) value (-74.6 \( \pm \) 5.4 mV) markedly negative to that of both the 5-HT3A(QDA) and 5-HT3A(QDA) D127N receptor constructs (Fig. 4E). The calculated \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) value is 0.06 \( \pm \) 0.01 (Table 2), which implies that the D127K mutant is effectively impermeable to Ca\(^{2+}\).

### The Effect of Neutralization and Charge Reversal Mutations at the 113 and 127 Loci upon Suppression of 5-HT3A(QDA) γ by Extracellular Ca\(^{2+}\)

The \( \gamma \) of the human wild-type 5-HT\(_3\)A receptor and the 5-HT3A(QDA) construct are depressed by extracellular Ca\(^{2+}\) (20, 27). For the 5-HT3A(QDA) construct, such suppression is concentration-dependent over the range 0.1–30 mM and voltage-independent over the range 0.3–100 mV in the presence of a [Na\(^{+}\)]_o fixed at 95 mM (20). The predominant suppression of \( \gamma \) by [Ca\(^{2+}\)]_o is chiefly observed over the concentration range 0.1–10 mM (extracellular solutions E3, E4, and E5; see Fig. 5). In view of the pronounced influence of the extracellular vestibule mutants upon the \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) ratio of the 5-HT3A(QDA) construct, the effect of extracellular Ca\(^{2+}\) on the \( \gamma \) of the 5-HT3A(QDA) receptor mutated at either the Asp\(_{113}^{	ext{mut}}\) or Asp\(_{127}^{	ext{mut}}\) position was investigated with [Ca\(^{2+}\)]_o set at 0.1, 1, or 10 mM in the presence of a [Na\(^{+}\)]_o of 95 mM.

In order to accurately calculate chord \( \gamma \) values, \( E_{55\text{-HT}} \) for each channel construct in each ionic condition was determined by a voltage ramp protocol (see “Experimental Procedures”) applied to macroscopic currents (individual values are reported in Fig.

#### Table 2

<table>
<thead>
<tr>
<th>Receptor constructs</th>
<th>( E_{55\text{-HT}} ) of Na(^{+}) extracellular solution E1 (mV)</th>
<th>( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} )</th>
<th>n</th>
<th>( E_{55\text{-HT}} ) of Ca(^{2+}) extracellular solution E2 (mV)</th>
<th>( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} )</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-HT(_3)A(QDA)</td>
<td>-2.1 ( \pm ) 0.7</td>
<td>0.87 ( \pm ) 0.03(^a)</td>
<td>21</td>
<td>10.1 ( \pm ) 1.2</td>
<td>3.85 ( \pm ) 0.3</td>
<td>17</td>
</tr>
<tr>
<td>5-HT(_3)A(QDA) D113N</td>
<td>-4.9 ( \pm ) 0.5</td>
<td>0.78 ( \pm ) 0.01(^b)</td>
<td>15</td>
<td>-11.6 ( \pm ) 1.1</td>
<td>1.04 ( \pm ) 0.08</td>
<td>6</td>
</tr>
<tr>
<td>5-HT(_3)A(QDA) D113K</td>
<td>-4.1 ( \pm ) 0.9</td>
<td>0.80 ( \pm ) 0.03</td>
<td>7</td>
<td>-22.7 ( \pm ) 1.8</td>
<td>0.60 ( \pm ) 0.06(^c)</td>
<td>8</td>
</tr>
<tr>
<td>5-HT(_3)A(QDA) D127N</td>
<td>-4.3 ( \pm ) 1.0</td>
<td>0.80 ( \pm ) 0.03</td>
<td>8</td>
<td>-55.8 ( \pm ) 1.2</td>
<td>0.13 ( \pm ) 0.01</td>
<td>6</td>
</tr>
<tr>
<td>5-HT(_3)A(QDA) D127K</td>
<td>2.1 ( \pm ) 0.9</td>
<td>1.02 ( \pm ) 0.03(^b)</td>
<td>8</td>
<td>-74.6 ( \pm ) 5.4</td>
<td>0.06 ( \pm ) 0.01(^c)</td>
<td>3</td>
</tr>
</tbody>
</table>

\(^a\) Includes data from Livesey et al. (20).

\(^b\) Significantly different \((p < 0.01)\) from the 5-HT\(_3\)A(QDA) construct, as determined by one-way ANOVA on ranks with post hoc Dunn’s test.

\(^c\) Significantly different \((p < 0.001)\) from the 5-HT\(_3\)A(QDA) construct, as determined by one-way ANOVA with post hoc Dunnett’s test.

**Figure 4.** Macroscopic I-V relationships for 5-HT\(_3\)A(QDA) receptor constructs harboring mutations to Asp\(_{113}^{	ext{mut}}\) and Asp\(_{127}^{	ext{mut}}\) within the ECD. I-V plots were generated by the voltage ramp protocols detailed under “Experimental Procedures,” and 5-HT (10 \( \mu M \))-evoked currents were recorded in an extracellular medium containing Ca\(^{2+}\) as the sole permeant ion species. Representative leak-subtracted whole-cell currents are shown for 5-HT\(_3\)A(QDA) A (A), 5-HT\(_3\)A(QDA) D113N (B), 5-HT\(_3\)A(QDA) D113K (C), 5-HT\(_3\)A(QDA) D127N (D), and 5-HT\(_3\)A(QDA) D127K (E) receptor constructs. Note that all mutations cause a negative shift in \( E_{55\text{-HT}} \) that is indicative of a reduced \( \frac{P_{\text{Ca}}}{P_{\text{Cs}}} \) ratio. Paralleling the latter is an enhancement of outward rectification of the macroscopic currents due to a progressively diminished ability to conduct Ca\(^{2+}\) inwardly.
The ECD of the 5-HT₃ₐ Receptor Influences Channel Biophysics

A. [Ca²⁺]₀ (mM)

<table>
<thead>
<tr>
<th>[Ca²⁺]₀</th>
<th>0.1</th>
<th>1</th>
<th>10</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-HT₃ₐ(QDA)</td>
<td>-15.0 mV</td>
<td>-14.4 mV</td>
<td>-10.2 mV</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D113N</td>
<td>-14.2 ± 0.7 mV (4)</td>
<td>-13.2 ± 0.5 mV (6)</td>
<td>-13.9 ± 0.5 mV (3)</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D113K</td>
<td>-14.3 ± 0.4 mV (3)</td>
<td>-13.7 ± 0.3 mV (5)</td>
<td>-11.6 ± 0.9 mV (7)</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D127N</td>
<td>-15.9 ± 0.2 mV (3)</td>
<td>-15.2 ± 0.4 mV (4)</td>
<td>-16.3 ± 0.2 mV (3)</td>
</tr>
</tbody>
</table>

B. γ (pS)

γ (pS) vs [Ca²⁺]₀

<table>
<thead>
<tr>
<th>[Ca²⁺]₀</th>
<th>0.03</th>
<th>0.3</th>
<th>3</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-HT₃ₐ(QDA)</td>
<td>50</td>
<td>40</td>
<td>30</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D113N</td>
<td>45</td>
<td>35</td>
<td>25</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D113K</td>
<td>40</td>
<td>30</td>
<td>20</td>
</tr>
<tr>
<td>5-HT₃ₐ(QDA) D127N</td>
<td>35</td>
<td>25</td>
<td>15</td>
</tr>
</tbody>
</table>

5A). Single channel current amplitudes were recorded at a holding potential of −80 mV from excised outside-out patches. The results of this study demonstrate that both Asp¹¹³ and Asp¹²⁷ within the ECD of the 5-HT₃ₐ(QDA) receptor have a profound effect upon the (i) amplitude of inwardly directed single channel currents, (ii) relative permeability to Ca²⁺, and (iii) modulation of γ by extracellular Ca²⁺. It should be emphasized that we have utilized the 5-HT₃ₐ(QDA) receptor as a convenient model system and that not all effects described here can readily be extrapolated to the human wild-type 5-HT₃ₐ receptor. For example, the reduction in amplitude of inwardly directed single channel currents caused by the neutralization or reversal of charge at the Asp¹¹³ and Asp¹²⁷ cannot be translated directly to the wild-type receptor whose γ is already severely limited by Arg⁴³², Arg⁴³⁶, and Arg⁴⁴⁰ within the large intracellular loop (16, 17).

All atom molecular dynamics simulations performed on a homology model of the adult human skeletal muscle ((α₁)₂β₁6ε) nACh receptor reveal that monovalent cations undergo stepwise changes in position as they translocate along the axis of the extracellular vestibule to the extracellular entrance of the transmembrane pore (22). Specifically, Na⁺ dwells for several ns in the vicinity of rings of acidic and polar residues that correspond to Glu⁸³ and Asp⁹⁷ of the α₁ subunit, and there is direct experimental evidence that α₁Asp⁹⁷ and residues at equivalent positions in the β₁, δ, and ε subunits influence γ in the adult human skeletal muscle nACh receptor (22, 26). Glu⁸³ and Asp⁹⁷ align with Asp¹¹³ and Asp¹²⁷ of the human 5-HT₃ₐ receptor (Fig. 1), providing the rationale for our mutagenesis program.

Effects upon γ—Neutralization of the negative charge at the 113 or 127 loci by the replacement of aspartate with asparagine produced a very substantial reduction in the amplitude of inwardly directed single channel currents corresponding to decreases in chord γ of 46 and 82%, respectively. It is notable that neutralization of the charge at the 127 position of the 5-HT₃ₐ(QDA) construct produces a percentage reduction in γ that closely approximates the effect of reversing the charge at the position corresponding to Asp⁹⁷ in all subunits of the adult skeletal muscle nACh receptor (26). When we mimicked the latter study more closely by the mutation D127K incorporated into the 5-HT₃ₐ(QDA) construct, 5-HT-evoked single channel currents could not be observed directly, but a noisy inward current was recorded from patches, confirming that the mutation did not ablate receptor expression. By fluctuation analysis of whole-cell currents mediated by the 5-HT₃ₐ(QDA) D127K mutant, we inferred a γ of ~1.6 pS. Fluctuation analysis applied to various 5-HT₃ₐ receptor mutants tends to underestimate...
the true γ revealed by single channel recording (17), and, extrapolating from previous correlative data, the γ of the 5-HT3A(QDA) D127K mutant could be closer to 3 pS. Nonetheless, by comparison with the D127K mutation, the substitution of aspartate by lysine at the 113 locus was better tolerated and caused only a modest decrease in γ further than that produced by charge neutralization (from 22 to 17 pS).

A parsimonious interpretation of the effect on γ of the mutations at the 113 position is through a simple electrostatic mechanism. First, the i-V data obtained with both the D113N and D113K constructs show that the mutations exclusively depress inwardly directed single channels in comparison with the 5-HT3A(QDA) receptor, introducing outward rectification to the i-V relationship. The degree of rectification increased as the charge at the 113 locus was neutralized (5-HT3A(QDA) D113N, RI = 1.70) or reversed (5-HT3A(QDA) D113K, RI = 2.63). Such an effect is consistent with Asp113 acting to increase the local concentration of cations within the outer extracellular vestibule, facilitating their outward translocation to deeper regions of the pore. Simple electrostatic interactions are sensitive to ionic strength, due to the screening of fixed charges by counterions (32). Although we did not examine the influence of ionic strength in a systematic manner (as elegantly performed by Kienker et al. (32) for the skeletal muscle nACh receptor harboring mutations in the extracellular (20°) and cytoplasmic (−4°) rings), we note parenthetically that the percent decrease in γ caused by the D113N and D113K mutations was more pronounced when [Na+]o was reduced from 146 to 95 mm. Attempts to evaluate the influence of lower concentrations of [Na+]o were unsuccessful due to reductions in γ that rendered quantitation unreliable.

The effect of mutations at the 127 locus was more complex. Although inwardly directed single channel currents were reduced to a greater extent than outwardly directed events by the D127N mutation (RI = 3.18), suppression of γ was apparent over a wide range of potentials (−100 to +100 mV). Unlike the D113 mutant, the percent decrease in the amplitude of inwardly directed currents was not increased at reduced ionic strength in a systematic manner (as elegantly performed in the aligned region of the pore. Simple electrostatic interactions are sensitive to ionic strength, due to the screening of fixed charges by counterions (32). Although we did not examine the influence of ionic strength in a systematic manner (as elegantly performed by Kienker et al. (32) for the skeletal muscle nACh receptor harboring mutations in the extracellular (20°) and cytoplasmic (−4°) rings), we note parenthetically that the percent decrease in γ caused by the D113N and D113K mutations was more pronounced when [Na+]o was reduced from 146 to 95 mm. Attempts to evaluate the influence of lower concentrations of [Na+]o were unsuccessful due to reductions in γ that rendered quantitation unreliable.

The affect of mutations at the 127 locus was more complex. Although inwardly directed single channel currents were reduced to a greater extent than outwardly directed events by the D127N mutation (RI = 3.18), suppression of γ was apparent over a wide range of potentials (−100 to +100 mV). Unlike the D113 mutant, the percent decrease in the amplitude of inwardly directed currents was not increased at reduced ionic strength, at least not within the limits examined. Strikingly, the D127K mutation reduced γ to a value that could not be determined directly at both negative and positive holding potentials. That the effect of mutations at the 127 locus upon inward currents is more pronounced than at the 113 position is not surprising because the former ring of charge is positioned closer to the entrance of the transmembrane region of the pore (Fig. 1). Furthermore, it is likely that the position of the 127 residue at the tip of the β4/5 loop (26, 33) causes it to protrude into the vestibule, forming a ring of charge that may be narrower than that present at the 113 locus. Indeed, this appears to be the case for the aligned region of the Torpedo nACh receptor (9). However, we refrain from attempting to specify the diameters of the forgoing rings because our homology model (Fig. 1) is based on the pseudosymmetrical structure of the Torpedo nACh receptor and probably does not reflect with sufficient accuracy the fine structure of the homo-oligomeric 5-HT3A receptor.

The data obtained with the 5-HT3A(QDA) D127N construct are inconsistent with solely a through-space electrostatic effect whereby the Asp127 residue simply elevates the local concentration of cations within the vestibule. Interestingly, Hansen et al. (26) demonstrated that suppression of γ in the adult human muscle nACh receptor is a non-linear function of the net charge upon the Asp127 ring and that two sets of mutations that resulted in the same change in net charge (i.e. the introduction of three lysine and two alanine residues versus the insertion of four lysine and retention of one aspartate residue) had significantly different effects upon γ. Thus, the side chain substitutions and their locations within the mutant subunits of the nACh receptor are at least as important as the net charge on the ring (26). In the present study, we observed the suppression of outwardly directed single channel currents by the D127N mutation, which would not be anticipated if the Asp127 residue served only to elevate the concentration of permeant cations within the extracellular vestibule. The molecular dynamics simulations performed by Wang et al. (22) upon the adult muscle nACh receptor reveal that cations are stabilized by the homologous Asp97 residue within the adult muscle nACh receptor. Such stabilization indicates the presence of an energy well that would be anticipated to affect both inward and outward ionic movements. Energy wells within the extracellular vestibule are also apparent as negative vestibular pore potentials computed for the Torpedo nACh receptor (23, 24). We speculate that the decrease or possibly reversal in pore potential caused by the D127N and D127K mutations, respectively, removes a force that facilitates both inwardly and outwardly directed ion fluxes or introduces a barrier against such movements.

Effects upon Charge Selectivity—None of the mutations examined in this study had a significant impact upon the charge selectivity of the 5-HT3A(QDA) receptor construct, as measured by the ratio PNa/PK. Such results are not surprising. First, it has been shown that mutating the inner (−1°) ring of charge of the mouse 5-HT3A receptor from glutamate to alanine renders the channel essentially non-selective between cations and anions (PNa/PK = 0.89) (34). When combined with the 19° mutation Ser → Arg, the channel gained modest anion selectivity (PNa/PK = 0.37) (34). Second, extracellularly applied anionic methanethiosulfonate reagents react with cysteine residues engineered into the mouse 5-HT3A receptor at multiple locations within the TM2 domain, demonstrating that anions can penetrate the extracellular vestibule (35, 36). Finally, Brownian dynamics simulations conducted upon the Torpedo nACh receptor indicate that although the extracellular vestibule contains a much higher concentration of Na+ than Cl−, the latter is not excluded (24).

Effects upon Valence Selectivity—In sharp contrast to the negative results found for charge selectivity, both the Asp113 and Asp127 residues were found to have a marked influence upon relative permeability to Ca2+, as reflected in the ratio Pca/PNa previously reported to be 3.8 for the 5-HT3A(QDA) receptor construct (20). For the D113N, D113K, D127N, and D127K mutations, Pca/PNa ratios were calculated to be 1.04, 0.60, 0.13, and 0.06, respectively. It is notable that the decreases in Pca/PNa parallel the decrements in γ caused by such mutations. At each locus, the suppression of Pca/PNa increased as the negative charge was neutralized or reversed, consistent with a simple through-space electrostatic effect. We attribute the greater influence of the Asp127 residue, in common with its
The ECD of the 5-HT₃A Receptor Influences Channel Biophysics

effect upon γ, as probably being due to its deeper location within the pore of the extracellular vestibule and inclusion within a narrower ring of charge. Permeant ion species would thus be concentrated in proximity to the entrance of the transmembrane pore.

For the α7 nACh receptor, it has long been known that mutations at the -1’ (inner ring), 16’, and 17’ loci exert a profound influence upon \(P_{Ca}/P_{Na}\) (15). Similarly, reductions in the negative charge at the -1’ locus of the α4β2 and α3β4 nACh receptors greatly reduces \(P_{Ca}/P_{Na}\) (37). Placing the present data in context, the D127K and D127N mutations produced effects upon relative permeability to Ca\(^{2+}\) of a magnitude comparable with or greater than those reported in the above studies. Asp\(^{127}\) appears to be a residue that is critical for the translocation of Ca\(^{2+}\) into the transmembrane pore of the 5-HT₃A(QDA) receptor construct. To the best of our knowledge, this is the first report to identify residues within the N-terminal outer vestibule of a pLGIC that influence valence selectivity. Moreover, the data add to those of our previous report illustrating relative permeability to Ca\(^{2+}\) in the 5-HT₃A(QDA) receptor to be influenced at multiple sites that include the extracellular (20’) ring of charge and residues within the intracellular MA stretch (i.e. Arg\(^{236}\)) (20). Others (34) have inferred a role for the -1’ (inner ring) in Ca\(^{2+}\) permeation based on Ca\(^{2+}\) imaging assays conducted upon the mouse wild-type 5-HT₃A receptor. Collectively, the data are consistent with a processional movement of Ca\(^{2+}\) from the extracellular to the intracellular environment, where multiple negatively charged residues along the axis of the permeation pathway facilitate the inward translocation of the ion. It should be noted that the occurrence of positive charge (i.e. lysine) at a position most likely homologous to Asp\(^{127}\) does not prevent permeability to Ca\(^{2+}\) in α1 glycine receptors mutated within TM2 and flanking sequences to impart cation selectivity (38).

Effects upon Suppression of γ by Extracellular Ca\(^{2+}\)—Extracellular Ca\(^{2+}\), within the concentration range 0.1–10 mM, causes a concentration-dependent reduction of the γ of the 5-HT₃A(QDA) receptor construct, amounting to a suppression of ~50% (20). By contrast, [Ca\(^{2+}\)]\(_o\) over the same concentration range had no significant effect upon the γ of either the 5-HT₃A(QDA) D113N (range 13.9 to 11.4 pS), or 5-HT₃A(QDA) D127N (range 6.6 to 4.8 pS), receptor constructs. An elevation of [Ca\(^{2+}\)]\(_o\) caused a statistically significant depression of the γ of the 5-HT3A(QDA) D113K mutant, but the effect was modest (range 11.5 to 9.8 pS). We could not evaluate the influence of [Ca\(^{2+}\)]\(_o\) upon the 5-HT3A(QDA) D127K construct due to its low γ. Collectively, the data indicate that neutralization or reversal of charge at the Asp\(^{113}\) or the Asp\(^{127}\) loci alleviates suppression of γ by extracellular Ca\(^{2+}\).

As noted previously, the molecular dynamics stimulations of Wang et al. (22) indicate that monovalent cations are stabilized at residues homologous to Asp\(^{113}\) and Asp\(^{127}\) in the adult muscle nACh receptor. From simple considerations of coulombic attraction, an even greater degree of stabilization of Ca\(^{2+}\) would be anticipated. The outer vestibule is sufficiently wide (9) to accommodate multiple Ca\(^{2+}\) ions, which, stabilized therein, would exert a repulsive force upon monovalent cations moving either inwardly or outwardly, thereby depressing γ. The location of the Asp\(^{113}\) and Asp\(^{127}\) residues outside of the transmembrane voltage field is consistent with the voltage-independent nature of the block by extracellular Ca\(^{2+}\) (20). We envisage that mutations that neutralize or reverse charge at the 113 and 127 loci reduce the stabilization of Ca\(^{2+}\) within the vestibule and thus alleviate block by extracellular Ca\(^{2+}\). It is notable that mutations at either location appear sufficient to reduce the influence of Ca\(^{2+}\) upon γ. It is possible that the primary site at which Ca\(^{2+}\) is stabilized is the Asp\(^{127}\) position, with the Asp\(^{113}\) residue acting to facilitate the translocation of the divalent to the former site. Such an effect would be consistent with the observation that mutations at the 113 position have a more modest effect on \(P_{Ca}/P_{Cs}\) than those at the 127 locus, which in the case of the D127K substitution renders the channel virtually impermeable to Ca\(^{2+}\). However, it is evident that for either mutation, the data show a tendency to a residual suppression of γ by Ca\(^{2+}\), perhaps suggesting additional site(s) of action within the conduction pathway. We have previously excluded the extracellular (20’) ring of charge as such a candidate (20).

Conclusions—The extracellular vestibule within the pLGIC family forms the initial obligatory pathway in the translocation of ions from the extracellular to intracellular environment (9). The results herein indicate that the extracellular vestibule of the 5-HT₃A(QDA) receptor contains at least two rings of negative charge that strongly impact upon γ and \(P_{Ca}/P_{Cs}\), and suppression of γ by extracellular Ca\(^{2+}\). By inference, homologous residues in other subunits within the pLGIC family might be anticipated to have similar functions, with the caveat that such extrapolation is from a receptor construct already modified by mutations within the large intracellular loop. In particular, virtually all cation-selective eukaryotic pLGICs contain aspartate at a position equivalent to the 127 locus (exceptions are the nACh α3 and α6 subunits), whereas the aligned residue or an adjacent residue in the anion-selective members is lysine (3).

Thus, the sign of the residue at this position is appropriate to the charge selectivity of the pLGIC and indeed the simultaneous mutation of Lys\(^{104}\) and Gly\(^{105}\) to alanine and aspartate (the aligned residues in the nACh α1 subunit) in the α1 glycine receptor has very recently been shown to depress γ (39). By contrast, the residue aligned to the 113 locus in cation-selective pLGICs is poorly conserved and presents as either a neutral, polar, acidic, or basic, but not hydrophobic, residue, suggesting that the importance of this position may vary across the receptor family. It is probable that other residues within the ECD influence receptor biophysics, a prominent candidate being α1 Asn\(^{47}\) of the nACh receptor of adult skeletal muscle highlighted in the molecular dynamics simulations of Wang et al. (22). In addition, it has already been mentioned by Bernal et al. (40) that mutation of the bovine nACh α7 Lys\(^{145}\) residue (aligned with 5-HT3A(QDA) Val\(^{154}\)) to alanine is associated with a modest but significant increase in γ. Moreover, it has recently been reported that the dual mutation D57I and R59T within the α1 glycine receptor significantly reduces γ (39). In aggregate, it is now evident, for both cation- and anion-selective pLGICs, that to fully account for the biophysical properties of ion permeation, amino acids residing within the TM2/TM1-TM2 linker region, the cytoplasmic portals, and the extracellular vestibule must be considered collectively.
The ECD of the 5-HT\textsubscript{3}A Receptor Influences Channel Biophysics

Acknowledgment—We are grateful to Tim G. Hales for critical comments upon a draft of the manuscript.

REFERENCES