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1	The long-term ecology and evolution of marine reptiles in a Jurassic seaway
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- 11 Marine reptiles flourished in the Mesozoic oceans, filling ecological roles today
- 12 dominated by crocodylians, large fish, sharks, and cetaceans. Many groups of these
- 13 reptiles coexisted for over 50 million years, through major environmental changes.
- 14 Little is known, however, about how the structure of their ecosystems or their ecologies
- 15 changed over millions of years. We use the most common marine reptile fossils—teeth—
- 16 to establish a quantitative system that assigns species to dietary guilds, and then track
- 17 the evolution of these guilds over the ca. 18 million year history of a single seaway, the
- 18 Jurassic Sub-Boreal Seaway of the United Kingdom. Groups did not significantly
- 19 overlap in guild space, indicating that dietary niche partitioning enabled many species
- 20 to live together. Although a highly diverse fauna was present throughout the history of
- 21 the seaway, fish and squid-eaters with piercing teeth declined over time while hard-
- 22 object and large-prey specialists diversified, in concert with rising sea-levels. High niche
- 23 partitioning and spatial variation in dietary ecology related to sea depth also
- 24 characterize modern marine tetrapod faunas, indicating a conserved ecological
- 25 structure of the world's oceans that has persisted for over 150 million years.

26 During the Mesozoic Era, when dinosaurs ruled the land, the oceans were teeming with a 27 diversity of large reptiles adapted to an aquatic lifestyle<sup>1-4</sup>. These included iconic groups such 28 as the long-necked plesiosauroids, big-headed pliosaurids, dolphin-like ichthyosaurs, and an 29 aberrant assemblage of crocodile relatives called thalattosuchians (subdivided into the 30 gharial-like teleosaurids and cetacean-like metriorhynchids). These groups were 31 exceptionally diverse and coexisted in the same environments for over 50 million years, from 32 the Early Jurassic (ca. 180 million years ago) to the Early Cretaceous (ca. 130 million years ago)<sup>5-12</sup>, through major changes in sea-level, climate, and ocean chemistry. They formed 33 34 complex ecosystems and filled a variety of ecological roles-ranging from fast-swimming fish-eaters to large-bodied apex predators<sup>7-9</sup>—that are today dominated by crocodylians (in 35 near-shore environments), sharks and other large fishes, and marine mammals<sup>1-2,13</sup>. As 36 modern oceans are undergoing rapid changes, understanding Mesozoic marine reptile 37 38 ecosystems may provide critical insight into how species at or near the top of the food chain 39 might respond to environmental shifts.

40 Although the phylogenetic relationships and taxonomic diversity patterns of Mesozoic marine reptiles have been the subject of intensive focus<sup>14-19</sup>, much less is known about their 41 42 ecology, about the structure of their ecosystems, and how their ecosystems changed over 43 millions of years of evolutionary time. Two main problems have hindered progress. First, it is 44 difficult to determine the ecological niches and feeding styles of extinct species, particularly when they are represented by limited fossil material. The pioneering work of Massare<sup>7-8</sup> 45 46 assigned marine reptiles to broad ecological guilds, but these are qualitative in nature and 47 have not been universally accepted<sup>9</sup>. Second, the marine reptile fossil record is notoriously patchy, dominated by fossil-rich localities (Lagerstätten) that register snapshots of 48 ecosystems but do not document long-term changes<sup>3</sup>. These issues hamper comparisons 49

between Mesozoic and modern marine faunas, and make it difficult to draw parallels between
the fossil record and contemporary environmental change.

52 Here, we use teeth-the most common marine reptile fossils, which are often 53 preserved in the absence of more complete skeletal remains-to evaluate feeding ecology. Our quantitative approach validates the guild system of Massare<sup>7-8</sup>, and allows species to be 54 55 placed in a functional morphospace that reflects the breadth of dietary niches. We then track 56 changes in how these niches were filled over time, focusing on the remarkable fossil record 57 of the Jurassic Sub-Boreal Seaway of the United Kingdom. Marine reptiles have long been 58 known from two Lagerstätten formed in this seaway, the Oxford Clay Formation (Callovian-59 early Oxfordian, ~165-161 Ma) and the Kimmeridge Clay Formation (Kimmeridgian-early Tithonian,  $\sim 157-148$  Ma)<sup>20-21</sup>, but recent work has clarified the fossil record of the 60 intervening Corallian Group (~161.5-157.3 Ma)<sup>17,22</sup>. Thus, we can now examine long-term 61 62 ecological changes in a single seaway spanning ca. 18 million years of the Middle-Late 63 Jurassic, a time of known environmental changes that began with recovery from a drop in 64 sea-level linked to a major glaciation, followed by a gradual rise in sea-level that culminated in a chaotic interval of rapid shifts in water depth<sup>23-30</sup>. 65

66

#### 67 **Results**

Principal Coordinates Analysis (PCO) of our dental morphology dataset ordinated all specimens into a morphospace, the first three axes of which describe over 63% of total variance (Fig. 1; Supplementary Figures 2-3; Supplementary Table 1). A linear discriminant analysis found that clusters of species in the morphospace correspond to the feeding guilds of Massare<sup>7</sup>, originally qualitatively defined based on comparisons of tooth shape, wear, and size and with modern marine tetrapods (Supplementary Table 6). When the 35 specimens in 74 our dataset (encompassing 25 taxa) that were also included in Massare's study were assigned 75 to Massare's guilds *a priori*, a jackknifing procedure found that our PCO axes successfully 76 discriminated these groups, with 84% of species correctly attributed to the guild Massare 77 assigned them to. This supports broad correspondence between our PCO morphospace and Massare's guilds, but 16% of species are mis-attributed, which may be because some species 78 79 previously known from limited fossil material (particularly metriorhynchids) were 80 misclassified when Massare introduced the guilds. We therefore used our PCO results to 81 establish a revised system of five guilds, which linear discriminant analysis finds to be 82 97.30% accurate. Along with the fact that characters in our dataset were chosen to represent 83 functionally significant features related to feeding, this indicates that the PCO morphospace 84 can be interpreted as a functional morphospace reflecting diet (Table 1). This link was previously demonstrated by Massare<sup>7</sup>, by assessing tooth morphology, shape and gut contents 85 of the same species, and validated with comparisons with modern marine tetrapods (for 86 87 which dietary data are available).

88

Gu	ild	Apex shape	Ornamentation	Cutting edges	Tooth size and shape	Examples
Cu	t	Pointed	Normally largely absent (in geosaurines, except cf. ' <i>Metriorhynchus</i> ' <i>hastifer</i> )	Two functional cutting edges always present	Very large: generally > 4cm. Robust crowns, mid-to-high CR*	Dakosaurus, Plesiosuchus, <b>Geosaurus</b> , cf. 'Metriorhynchus' hastifer
Generalist	Generalist- Cut	Pointed	Present: high- relief with serration morphologies	Two or more cutting edges always present	Very large: generally > 5cm. Robust crowns, mid-to-high CR*	Pliosaurus spp.
	Generalist- Pierce	Pointed	Variable, it may be high relief	None, or non- prominent. If present they are smooth and non-serrated	Variably large: generally > 5cm. Normally conical. Variable CBR.	Liopleurodon, Simolestes, 'Pliosaurus' andrewsi

Pierce	Pointed	Variable, but never high relief	None, or non- prominent. In metriorhynchin es if they are present they are smooth, or not functionally serrated.	Normally small (<3cm), but can be large (>3cm). Mid- to-high CR*, lateral compression variable (weak or absent, in plesiosauroids and metriorhynchines; laterally compressed in geosaurines).	All plesiosauroids, <b>MJML K1885,</b> 'Steneosaurus' leedsi, Mycterosuchus, Gracilineustes, Peloneustes; <b>Metriorhynchus,</b> <b>Suchodus,</b> Tyrannoneustes, 'M.' cultridens
Smash	Pointed to Rounded	NA	None	Conical. Mid-CR*	Ophthalmosaurus, <b>Brachypterygius</b>
Crunch	Rounded	Strongly ornamented, with high-relief ridges and may have serration morphologies	Prominent carinae only on the top half of the crown.	Robust and conical, poorly to non- laterally compressed. Mid-to- low CR*.	Lemmysuchus, Machimosaurus, Torvoneustes

Table 1. Description of the main tooth features diagnosing the five guilds as used in this study. Note that this is
a key that can be used to assign specimens to guilds based on consideration of a few general features, but more
rigorous assignment can be made numerically using Linear Discriminant Analysis of PCO scores. Taxa in bold
are classified in a different guild compared to Massare's System<sup>7</sup> (see Supplementary Table 7, Appendix S3, S4).
The table is adjusted from Massare (1987)<sup>7</sup>. \*CR: Crown height to base Ratio.

94

95 Many thalattosuchians (teleosaurids and metriorhynchids), plesiosauroids, and small-bodied 96 pliosaurids cluster together in morphospace, due to their thin teeth that lack serrations (Pierce 97 Guild), similar to modern shallow-water river dolphins and gharials; they are inferred to be 98 fish and squid-eaters (Fig. 1, Table 1). An adjacent region of morphospace is occupied by the 99 soft prey-eating Smash Guild, which is almost exclusively filled by ichthyosaurs with 100 conical, non-carinated and non-serrated teeth (Fig. 1, Table 1). Strongly ornamented, blunt crowns of machimosaurin teleosaurids and the geosaurine Torvoneustes belong to the Crunch 101 102 Guild, inferred as hard-object (e.g. sea turtles) feeders (durophages) (Fig. 1, Table 1). There 103 are two clusters of cutting-type dentitions that correspond to different types of macrophagy 104 (feeding on large-bodied prey), analogous to modern deep-water cetaceans like killer whales: 105 the Cut Guild of geosaurine metriorhynchids (Geosaurus, Plesiosuchus, and Dakosaurus) 106 with large and serrated teeth, and the Generalist-Cut Sub-Guild (the macrophagous partition

of the Generalist Guild) of pliosaurids with huge, robust, heavily ornamented teeth
(*Pliosaurus*) (Fig. 1, Table 1). The Generalist-Pierce Sub-Guild (the other partition of the
Generalist Guild), characterised by large, conical and variably ornamented teeth and mostly
including large Callovian pliosaurids, fills a space between the Generalist-Cut and Pierce
Guilds (Fig. 1, Table 1).

112 We assessed changes in the dietary ecology of marine reptiles during the ca. 18 113 million year history of the Sub-Boreal Seaway by testing for changes in morphospace overlap 114 and morphological disparity (amount of occupied morphospace) between phylogenetic 115 groups. The major marine reptile groups do not significantly overlap in morphospace overall, 116 and within each time slice (Appendix S2), indicative of large-scale dietary niche partitioning, 117 but also a likely phylogenetic signal (e.g. through shared evolutionary ancestry). However, 118 individual groups show evidence of morphospace migration over time (Fig. 2). Pliosaurids 119 are the most extreme example, as they make a (nearly) significant jump (Supplementary 120 Table 2) between their scattered Callovian morphospace into a more restricted region in the 121 ensuing Oxfordian. This change is followed by another significant shift between the 122 Oxfordian and Kimmeridgian, as a consequence of the appearance of the genus *Pliosaurus*. 123 Other groups do not exhibit significant shifts between successive time intervals, but 124 do show significant changes between end points (e.g., Callovian vs. Tithonian, or Oxford Clay vs. Kimmeridge Clay Formations), which is indicative of gradual ecological changes. 125 126 There are no significant differences over time in the disparity of the pooled sample of 127 all marine reptile groups in the morphospace (as shown by permutation tests on range and 128 variance measures of morphospace occupation), indicating that an ecologically diverse fauna 129 persisted throughout the entire history of the seaway (Supplementary Table 5). There are few 130 instances of significant disparity changes within individual groups across successive time 131 intervals, despite turnover at the species level (Fig. 2). Metriorhynchid groups are the

132 exceptions, as geosaurines significantly increased in disparity from the Oxfordian to the 133 Kimmeridgian, as their variety of niches expanded, in concert with the decline of fish-eating 134 metriorhynchines. This implies that, although groups were migrating through morphospace 135 over time, most continued to fill a similar breadth of niches, albeit in different regions of the 136 morphospace (Fig.2). Partial disparity trends show that teleosaurids and metriorhynchids 137 make up a large proportion of overall disparity in the Callovian but decline over time, 138 whereas geosaurines and macrophagous pliosaurids (Pliosaurus) become relatively more 139 disparate (Fig. 3). When partial disparity is instead tracked by guild. Pierce taxa are highly 140 disparate early but decline into the Oxfordian and beyond, as Crunch, Smash, and Cut species 141 assume a larger share of overall disparity (Fig. 3).

142 These quantitative tests allow us to identify several major trends. Pliosaurids were 143 diverse in the Callovian, ranging across a broad span of fish-eating Pierce and apex-predator 144 Generalist/Generalist-Pierce guilds, but then in the Oxfordian atrophied into a more restricted 145 region of macrophagous morphospace (Generalist/Generalist Cut), where they diversified 146 further in the Kimmeridgian, without migrating back into the Pierce Guild. Similarly, 147 teleosaurids were highly diverse in the Callovian, including long-snouted fish-eaters in the 148 Pierce Guild and durophages in the Crunch Guild, but the former species mostly disappeared 149 from the Oxfordian onwards while the hard-object feeders persisted. Geosaurines began with 150 a large diversity of taxa spanning the Pierce Guild in the Callovian, and in the Oxfordian 151 bifurcated into a Crunch group that approached the durophagous teleosaurids in morphospace 152 and a Cut group of large-prey specialists, which were distinct from the macrophagous 153 Generalist-Cut pliosaurids in morphospace. Metriorhynchines included several Pierce Guild 154 piscivores in the Callovian, but then mostly disappeared afterwards. Ichthyosaurs were 155 relatively stable in almost exclusively occupying the soft-prey-eating Smash Guild from the 156 Callovian to Tithonian, but their share of partial disparity increases over time. Sample sizes

157 for plesiosauroids are too small to reasonably interpret, although they seem to stably occupy158 the Pierce Guild through time.

#### 159 **Discussion**

In agreement with previous studies 5,7,15, we argue that there is a tight link between tooth 160 161 morphology and dietary ecology in Jurassic marine reptiles. By assessing a limited number of 162 discrete features and measurements of the teeth, species can be placed in a functional 163 morphospace that distinguishes ecological guilds (Fig. 1). This morphospace differentiates 164 marine reptiles that fed in different ways, including soft-object feeders, piercers of fish and 165 squid, durophagous hard-object crushers, and apex predator macrophages that specialized on large vertebrate prey<sup>5,7,15</sup>. Our approach is a powerful tool for predicting the feeding habits of 166 167 long-extinct species, as it focuses on the most commonly preserved marine reptile fossils 168 (teeth), and can incorporate even minimally complete specimens. Our approach provides a quantitative method for projecting Massare's guilds<sup>7</sup>—which have long been used by marine 169 170 reptile workers-into a numerically-derived morphospace, meaning specimens can be 171 assigned to these guilds more objectively, and changes in guild occupation over time can be 172 analysed statistically.

173 Our study reveals the ecological structure of marine reptile faunas in the Sub-Boreal 174 Seaway during ca. 18 million years of the Middle-Late Jurassic. The positions of species in our functional morphospace give insight into niche occupation. The major marine reptile 175 176 groups do not significantly overlap with each other in morphospace, meaning that different 177 groups generally employed distinct dietary strategies (possibly in part due to shared 178 evolutionary ancestry). Partitioning niches in this way may explain why such a great diversity 179 of marine reptiles were able to coexist in the Mesozoic oceans<sup>13</sup>, and parallels the situation in 180 modern oceans, where aquatic tetrapods subdivide ecological space by feeding in different

ways and on different prey<sup>31-40</sup>. Evidently, Jurassic marine reptiles partitioned niches to a fine 181 182 degree, as testified by the development of two distinct and coexisting forms of macrophagy: 183 the robust-toothed pliosaurids in the Generalist-Cut Guild and the thinner, slicing-toothed 184 geosaurines of the Cut Guild. Other groups converged on similar niches-most notably machimosaurin teleosaurids and Torvoneustes-lineage geosaurines that independently 185 186 colonized the same durophagous area of morphospace-which speaks to the maintenance of key niches over long spans of time. We also find no significant differences in overall 187 188 morphological disparity of the Sub-Boreal assemblages over time, illustrating that a diverse 189 marine reptile fauna was present throughout the entire history of the seaway, and in general 190 terms was resilient to environmental change.

191 Nonetheless, the shifting distribution of species in the morphospace over time does 192 provide evidence for long-term ecological changes. Although the imprecise ages of fossils 193 make it difficult to untangle the tempo of these changes, they appear to be a mix of sudden 194 transformations at or near the Callovian-Oxfordian (Middle-Late Jurassic) boundary and 195 more gradual changes that occurred throughout the Oxfordian, Kimmeridgian, and Tithonian, 196 as sea-levels rose and water depth increased. Most severe was the decimation of the Pierce 197 Guild, which was filled by a diversity of teleosaurids, metriorhynchines, and small-bodied 198 pliosaurids in the Callovian, but was then largely vacated from the Oxfordian onwards. 199 Essentially, this reflects a drastic decline of nearshore, piscivorous species. Metriorhynchines 200 excelled in this niche and were the most abundant component of the Callovian Oxford Clay 201 ecosystem, but are almost unknown from the later Kimmeridge Clay. Replacing the Pierce 202 species was a greater variety of macrophagous taxa, particularly large pliosaurids and 203 geosaurines, and durophages, including the specialized machimosaurin lineage of 204 teleosaurids. At the same time, ichthyosaurs maintained their position in the Smash Guild and 205 diversified further, reflecting a blossoming of deep-diving species that fed on similar prey to

206 the more nearshore piercers that were devastated.

207 We hypothesize that these large-scale changes in marine reptile functional diversity 208 and niche occupation in the Jurassic Sub-Boreal Seaway were related to changes in water 209 depth over time, analogous to how modern aquatic tetrapod diversity and ecology are known to change from shallow water (nearshore) to deeper water (offshore) habitats<sup>31-40</sup>. It is well 210 211 established that the Sub-Boreal Seaway became generally deeper as global sea-levels rose 212 from the Callovian into the Oxfordian, and then across the Oxfordian, Kimmeridgian, and 213 early Tithonian. The main fossiliferous beds of the Callovian Oxford Clay Formation were deposited in nearshore waters ca. 50 metres deep<sup>25-26,30</sup>; the middle-late Oxfordian Corallian 214 215 Group rocks formed in deeper waters during a time of warming and sea-level transgression<sup>17,20-22,27-28,30</sup>; and the main fossil-bearing units of the Kimmeridge Clay 216 Formation, spanning the Kimmeridgian-Tithonian boundary, were deposited on continental 217 218 shelves 100-200 metres deep<sup>29,30</sup>. Although numerous factors (ranging from water 219 temperature and turbidity to productivity and seasonality) control the distribution of extant 220 marine tetrapods and their prey, water depth is widely recognized as one of the key drivers of ecological partitioning<sup>33-40</sup>. While small delphinoids such as bottlenose and spotted dolphins 221 222 can be found both nearshore and sometimes further shelf-ward, large delphinoids like killer whales predominately live in deeper, open waters and infrequently venture nearshore<sup>36-38</sup>, and 223 their abundance on continental shelves increases with water depth<sup>39</sup>. The diets of mid-shelf to 224 225 offshore delphinoids also becomes increasingly dominated by larger prey, and only off-shelf 226 species actively prey upon large tetrapods<sup>13,31,39</sup>.

In the Jurassic Sub-Boreal Seaway, a variety of marine reptiles shared environments by feeding in different ways, and nearshore fish-eating marine reptiles declined and openocean macrophages and durophages proliferated as sea-level increased over millions of years of evolutionary time. In the modern oceans, numerous species of large vertebrates coexist by partitioning dietary niches, and there is a noted ecological partitioning of shallow-water
small-object feeders and deep-water, large-prey specialists. These parallels between Jurassic
and modern oceans—separated in time by ca. 150 million years, occupied by different groups
of species, and shaped by vastly distinct paleogeography, temperature, and ocean chemistry—
may indicate a conserved ecological structure near the top of ocean food webs over time.

- 236
- 237 Methods
- 238

239 Dataset. We compiled a dataset of 22 functionally-relevant anatomical characteristics of the 240 teeth scored for 122 specimens of marine reptiles (representing ~50 species) that lived in the 241 Jurassic Sub-Boreal Seaway of the modern-day United Kingdom during the Callovian-242 Tithonian stages of the Jurassic (Appendix S1). The specimens come from a variety of localities and lithological facies within the Jurassic Sub Boreal Seaway basins<sup>21</sup>. It is possible 243 244 that the lower number of Oxfordian specimens is related to preservation (or sampling) bias of 245 the Corallian Group facies compared to the fossil-rich and better exposed Oxford Clay Formation and Kimmeridge Clay Formation<sup>21</sup>. However, by focusing on the most commonly 246 247 collected and preserved fossils (teeth), we minimize these biases. We focused on dental 248 morphology for the following reasons: 1) teeth are more commonly preserved as fossils than 249 any other part of the marine reptile skeleton; 2) teeth were the primary source of information 250 that Massare<sup>7</sup> used to assign marine reptiles into feeding guilds; 3) multivariate analyses of 251 tooth measurements, of the type encapsulated in our dataset, have been commonly applied to other reptile groups (most notably dinosaurs<sup>41,42</sup>) to determine feeding habits and track long-252 253 term trends in palaeoecology.

Included in our dataset are teeth belonging to the three major marine reptile clades
that lived in the Sub-Boreal Seaway—Plesiosauria, Ichthyosauria, and Thalattosuchia—and

256 all major subclades within these lineages. We designed our specimen sampling to maximise 257 the number of complete specimens for each taxon in each time bin. We selected an 258 approximately equal number of specimens for each taxon to avoid oversampling biases. All 259 tooth crowns in our dataset are from the anterior part of the tooth row, to avoid inflation of diversity caused by the marked heterodonty of some taxa<sup>15,43,44</sup>. Selecting the largest teeth 260 261 does not constitute a major problem in the interpretation of the final results because the signal 262 in the analyses is dominated by the discrete characters, which code features that do not 263 significantly change along the tooth row. Additionally, the largest 'fangs' are those that 264 undergo the largest stress during predation, and thus are ideal for investigating feeding ecology<sup>7,43,44</sup>. It is also worth noting that disarticulated teeth cannot always be oriented in 265 266 their correct mesiodistal orientation, so the distinction between upper or lower jaw teeth is 267 unnecessary.

Each specimen was scored for the 5 continuous and 17 discrete characters in our 268 269 dataset (Appendix S1), which were selected largely from published phylogenetic and 270 functional datasets<sup>12,18,19,45,46</sup>. The main sources for discrete characters were the analyses of Young et al. (2016)<sup>46</sup>, Foffa et al. (2017)<sup>45</sup>, and Fischer et al. (2015, 2016, 2017)<sup>12,18,19</sup>; as 271 272 these studies were designed for specific lineages, it was necessary to modify the definitions 273 and states of some characters, to make them broadly applicable to all marine reptiles. The 274 modified characters were specifically targeted to describe tooth shape, proportions, 275 ornamentation, and cutting edge (carina and denticle) morphologies (Table 1, 2) 276 (Supplementary Figure 1). Most continuous measurements were taken directly from 277 specimens with digital callipers, except for the curvature and crown angles (C4 and C5) that were measured using ImageJ<sup>47</sup>, and a small number that were taken from the literature 278 279 (Supplementary Information, Supplementary Figure 1).

General aspect	Carinae and	Ornamentation	Roots
	serrations		

C1. Apicobasal crown height – CH	<b>D1.</b> Crown mediolateral compression	<b>D2.</b> Carinae: presence or absence	<b>D7.</b> Enamel surface ornamentation: anastomosed pattern		
<b>C2.</b> Crown ratio – CR	D12. Crown cross-section	<b>D3.</b> Denticles: presence and size	<b>D8.</b> Enamel ornamentation presence and density: lingual side	D16. Root cross-section	
C3. Crown base ratio – CBR	<b>D13.</b> Trifaceted labial side	<b>D4.</b> 'Functionally serrated edges': presence or absence	<b>D9.</b> Enamel ornamentation presence and density: labial side		
C4. Labial- Lingual curvature – LLcufrv	<b>D15.</b> Tooth crown apex – shape	<b>D5.</b> Denticles: distribution along the carinae	<b>D10.</b> Enamel ridges, relief	<b>D17.</b> Bulbous	
C5. Crown Angle – Cang		<b>D6.</b> Presence or absence of denticle-like structures	D14. Enamel texture	root larger than the crown	
		<b>D11.</b> Ornamentation interfering with the carinae or cutting edges – false denticles			

Table 2. Table of continuous (C) and discrete (D) morphological characters used to represent marine reptile
 dentitions. See Supplementary Information for extended descriptions of each character.

282

283 Multivariate analysis. The continuous characters were standardised using z-transformation 284 (distributions were equalized to the same mean value,  $\mu = 0$ , and standard deviation,  $\sigma =$ 1)<sup>11,48</sup>. The entire taxon-character matrix (Appendix S1) was then transformed into a Gower 285 286 distance matrix (necessary for the combination of ordered discrete and continuous characters)<sup>49</sup> and subjected to Principal Coordinates Analysis (PCO) in PAST v.3 and R 287 v3.4.3<sup>50,51</sup>, following the procedure in Stubbs and Benton (2015)<sup>11</sup>. The small number of 288 289 missing values (8.2% of the total scores) were automatically accounted for by pairwise 290 deletion. The scores of the first three PCO axes (which describe 28.54%, 22.16%, and 291 12.43% of total variance, respectively: Supplementary Table 1) were used to generate 292 morphospaces for data visualization (Figs. 1-2, Supplementary Figures 1-2), and the 15 PCO 293 axes that accounted for at least 0.5% of total variance were retained for discriminant function 294 analysis, statistical tests of morphospace overlap, and disparity calculations. We ran a Linear Discriminant Analysis (LDA) (Appendix S3) to test the ability of the 295

296	PCO scores to assign individuals to the feeding guild assignments of Massare <sup>7</sup> . The 35
297	specimens in our dataset that were also included in Massare's study <sup>7</sup> were a priori assigned to
298	the guilds that Massare originally assigned them to (four guilds total: Pierce [also tested for
299	separate Pierce I and Pierce II], Crunch, Smash, Cut) (for more details see the LDA Guild
300	Sensitivity Analyses in Appendix S4), and the percentages of correct specimen-guild matches
301	were determined using a jackknifing test in PAST v.3 <sup>50</sup> (see Supplementary Information,
302	Appendix S3). This revealed that 84% of specimens were correctly assigned to the Massare's
303	original guilds (the same percentage, 84%, if the Pierce I and II guilds of Massare are split
304	into two), indicating a broad correspondence between our quantitative PCO morphospace and
305	Massare's qualitative guilds (see Supplementary Information, Appendix S3, S4).
306	New fossil discoveries and reanalyses of key specimens suggest that some of the guild
307	assignments of Massare may be problematic, perhaps explaining why 16% of specimens are
308	incorrectly assigned by the LDA. Over the last few decades, many specimens used in
309	Massare's study have been re-evaluated (particularly metriorhynchids) and further details of
310	tooth morphology have been described (particularly regarding carinae and denticles)
311	(Appendix S3). Thus, we used the results of our PCO analysis—particularly the visual spread
312	of taxa in morphospace-to modify Massare's guilds into a new system of five guilds: Pierce,
313	Crunch, Smash, Cut, and Generalist (subdivided into Generalist-Pierce, and Generalist-Cut)
314	(Appendix S3, S4). We assigned the same 35 specimens from taxa shared with Massare's
315	study <sup>7</sup> to one of these guilds a priori, and then ran a second LDA, which correctly attributes
316	97.3% of the specimens to the proper guild. This indicates that our morphospace can be used
317	to sensibly cluster species into guilds, and thus be interpreted as a dietary function space.
318	This second LDA also serves to classify each of our 122 specimens into its best-fit guild. The
319	regions of our PCO morphospace occupied by each guild are denoted in Figure 1. This is the
320	first attempt to project Massare's <sup>7</sup> qualitative guilds into a numerically-derived morphospace.

Note that in creating our revised guild system, we preferred a unified Pierce guild over
Massare's<sup>7</sup> original subdivision of Pierce I and Pierce II guilds, because this distinction relied
on wear patterns and features of cranial morphology that are not captured in our dataset (but
see Appendix S4 for sensitivity analyses).

Statistical analyses. Specimens were binned by clade, guild, and time in order to conduct
statistical analyses of morphospace overlap and morphological disparity.

We employed a hierarchy of taxonomic bins. We broadly divided the specimens into 327 328 the three main order-level clades: Plesiosauria, Ichthyosauria, and Thalattosuchia. We then 329 ran additional analyses in which these large clades were subdivided into less inclusive 330 monophyletic groups of family-level, and subfamily-level. Plesiosauria was subdivided into 331 Plesiosauroidea and Pliosauridae. Thalattosuchia was split into Metriorhynchidae and 332 Teleosauridae, both of which were further divided (into Metriorhynchidae/Geosaurinae for 333 the former, and into Machimosaurini/non-Machimosaurini for the latter). As all ichthyosaur 334 specimens belong to Ophthalmosauridae, this group was not subdivided. Finally, for the 335 disparity analyses, we examined three further lineages within Pliosauridae (non-Pliosaurus 336 pliosaurids and Pliosaurus spp.) and Geosaurinae (the 'T-clade': Torvoneustes-'Metriorhynchus' hastifer-Mr. Passmore's specimen clade ; the 'GPD clade': Plesiosuchus-337 338 Dakosaurus-Geosaurus clade; and 'Basal Geosaurinae', which includes 'Metriorhynchus' 339 brachyrhynchus, with and without Tyrannoneustes lythrodectikos - because of the unstable position of the latter in the latest phylogenetic analyses<sup>17,45</sup>). 340 341 We employed four time bins when analysing trends over the Jurassic. These 342 correspond to stage level: Callovian (166.1–163.5 Ma), Oxfordian (163.5–157.3 Ma), 343 Kimmeridgian (157.3–152.1 Ma), and early Tithonian (152.1–148.5 Ma). The average length 344 of these time bins is 4.4 Ma, with a minimum of 2.6 Ma for the Callovian and maximum of

345 6.2 Ma for the Oxfordian. Given the age uncertainty of several specimens, equal-length bins

346 were deemed unreasonable because too many specimens would be assigned to multiple time 347 bins, decreasing the power of our analyses. We ran subsidiary analyses in which specimens 348 was also assigned to stratigraphic bins representing the main marine reptile faunas of the Sub-349 Boreal Jurassic Seaway: Oxford Clay Formation (~Callovian - early Oxfordian), 'Corallian Group' (middle - late Oxfordian), and Kimmeridge Clay Formation (Kimmeridgian - early 350 351 Tithonian). This binning scheme demonstrates changes in a series of temporally successive 352 Lagerstätten. We recognize that, for both binning schemes, each time bin includes a time-353 averaged sample, as is necessarily the norm in most palaeoecological analyses of long-term 354 change. Thus, these bins do not exactly represent individual ecosystems that existed at a 355 moment in time, but rather summarize the pool of species present during the finest temporal 356 resolution available.

357 We assessed whether taxonomic groups significantly overlapped in the overall 358 morphospace, and for each time interval, using non-parametric multivariate analysis of variance (NPMANOVA)<sup>51,11,48</sup> in R<sup>52</sup> (Appendix S2). The null hypothesis is that there is no 359 360 difference in the location of group centroids in PCO space; significant deviation from the null 361 indicates that the groups in question occupy significantly different areas of morphospace. The 362 p values were adjusted using the false-discovery rate (FDR) method to account for 'false discoveries' error that may be introduced by multiple comparisons<sup>53</sup>. Changes in 363 morphospace occupation over time, if confirmed by NPMANOVA, indicate the migration of 364 365 clades into different niche space.

We measured the breadth of morphospace occupied by taxa by calculating morphological disparity. We used two metrics to quantify disparity: the sum of variances and the sum of ranges on the PCO axes for each group in question<sup>54</sup>. Variance is a measure of the spread of taxa in morphospace relative to the group centroid, whereas range denotes the overall volume of morphospace occupied by the group. These disparity metrics were calculated in R using functions in the 'dispRity' package<sup>55</sup> Statistical significance of disparity
differences between groups, or within groups across time, were assessed by a permutation
test developed by Steve Wang<sup>56</sup>, which determined whether there was a significant difference
in the observed disparity value between the two comparisons and the expected disparity
difference between them. As the expected difference is generated based on the size of each
group, this procedure accounts for sample size differences between the groups, which
otherwise could be a major source of bias (particularly for sum of ranges).

We also tracked partial disparity over time, for taxonomic groups and guilds (Fig. 3). The contribution of each group/guild to overall disparity in each time bin was calculated as the average squared distance of each group/guild member point from the overall time bin centroid, with the resulting value weighted by a factor taking into account the sample size of the group/guild compared to the overall sample<sup>57</sup>. This procedure was conducted with the Morphological Disparity Analyses (MDA) package for MATLAB® R2016a v9.0.0<sup>58</sup>.

#### **Data availability**

The authors declare that all the data supporting the findings of this study are available within the paper and its supplementary information files. The R-codes used to perform statistical tests are available from the corresponding author upon reasonable request.

### 388 References

389 1. Pyenson, N.D., Kelley, N.P., and Parham, J.F. 2014. Marine tetrapod macroevolution:

390 Physical and biological drivers on 250 Ma of invasions and evolution in ocean

391 ecosystems. *Palaeogeography, Palaeoclimatology, Palaeoecology*. 400: 1–8.

392 2. Kelley, N.P., and Pyenson, N.D. 2015. Evolutionary innovation and ecology in marine

- tetrapods from the Triassic to the Anthropocene. *Science* 348, aaa3716.
- 394 DOI:10.1126/science.aaa3716

- 395 3. Benson, R.B.J., Butler, R.J., Lindgren, J., and Smith, A.S. 2010. Mesozoic marine
- 396 tetrapod diversity: mass extinctions and temporal heterogeneity in geological megabiases
- 397 affecting vertebrates. *Proceedings Royal Society B* 277: 829–834.
- 398 4. Ciampaglio, C.N., Wray, G.A., and Corliss, B.H. 2005. A Toothy Tale of Evolution:
- 399 Convergence in Tooth Morphology among Marine Mesozoic Cenozoic Sharks,
- 400 Reptiles, and Mammals. *The Sedimentary Record* 3(4): 4–8.
- 401 5. Andrews, C.W. 1909. A descriptive catalogue of the marine reptiles of the Oxford Clay,
- 402 Part One. London: British Museum (Natural History).
- 403 6. Andrews, C.W. 1913. A descriptive catalogue of the marine reptiles of the Oxford Clay,
- 404 Part Two. London: British Museum (Natural History).
- 405 7. Massare, J.A. 1987. Tooth morphology and prey preference of Mesozoic marine reptiles.
  406 *Journal of Vertebrate Paleontology* 7: 121–137.
- 407 8. Massare, J.A. 1988. Swimming capabilities of Mesozoic marine reptiles: implications for
  408 method of predation. *Paleobiology* 14: 187–205.
- 409 9. Buchy, M.-C. 2010. Morphologie dentaire et régime alimentaire des reptiles marins du
- 410 Mésozoïque: revue critique et réévaluation. Oryctos, 9: 49-82.
- 411 10. Chiarenza, A.A., Foffa, D., Young, M.T., Insacco, G., Cau, A., Carnevale,
- 412 G., and Catanzariti, R. 2015. The youngest record of metriorhynchid crocodylomorphs,
- 413 with implications for the extinction of Thalattosuchia. *Cretaceous Research*, 56: 608–616.
- 414 11. Stubbs, T.L., and Benton, M.J. 2015. Ecomorphological diversifications of Mesozoic
- 415 marine reptiles: the roles of ecological opportunity and extinction. *Paleobiology* 1-27.
- 416 doi:10.1017/pab.2016.15
- 417 12. Fischer, V., Bardet, N., Benson, R.J.B., Arkhangelsky, M.S., and Friedman, M. 2016.
- 418 Extinction of fish-shaped marine reptiles associated with reduced evolutionary rates and
- 419 global environmental volatility. *Nature Communications*. 7: 10825.

- 420 13. Kelley, N.P., and Motani, R. 2015. Trophic convergence drives morphological
- 421 convergence in marine tetrapods. *Biology Letters* 11: 20140709.
- 422 14. Young, M.T., Brusatte, S.L., Andrade, M.B., Desojo, J.B., Beatty, B.L., Steel, L.,
- 423 Fernández, M.S., Sakamoto, M., Ruiz-Omeñaca, J.I., and Schoch, R.R. 2012a. The
- 424 cranial osteology and feeding ecology of the metriorhynchid crocodylomorph genera
- 425 *Dakosaurus* and *Plesiosuchus* from the Late Jurassic of Europe. *PLoS ONE* 7, e44985.
- 426 15. Young, M.T., Brusatte, S.L., Andrade, M.B., Beatty, L., and Desojo, J.B. 2012b. Tooth-
- 427 On-Tooth Interlocking Occlusion Suggests Macrophagy in the Mesozoic Marine
- 428 Crocodylomorph *Dakosaurus*. *The Anatomical Record* 295: 1147–1158.
- 429 16. Benson, R.B.J., and Druckenmiller, P.S. 2014. Faunal turnover of marine tetrapods during
  430 the Jurassic–Cretaceous transition. *Biological Reviews* 89: 1–23.
- 431 17. Young, M.T. 2014. Filling the 'Corallian Gap': re-description of a metriorhynchid
- 432 crocodylomorph from the Oxfordian (Late Jurassic) of Headington, England. *Historical*
- 433 *Biology: An International Journal of Paleobiology* 26(1): 80–90.
- 434 18. Fischer, V., Arkhangelsky, M.S., Stenshin, I.M., Uspensky, G.N., Zverkov, N.G., and
- 435 Benson, R.B.J. 2015. Peculiar macrophagous adaptations in a new Cretaceous pliosaurid.
- 436 *Royal Society Open Science* 2: 150552.
- 437 19. Fischer, V., Benson, R.B.J., Zverkov, N.G., Soul, L.C., Arkhangelsky, M.S., Stenshin,
- 438 I.M., Lambert, O., Stenshin, I.M., Uspensky, G.N., and Druckenmiller, P.S. 2017.
- 439 Plasticity and Convergence in the Evolution of Short-Necked Plesiosaurs. *Current*
- 440 *Biology* 27: 1667–1676.
- 441 20. Cox, B.M. 2001. In Wright, J.K., and Cox, B.M. British Upper Jurassic Stratigraphy,
- 442 Geological Conservation Review Series, No. 21, Joint Nature Conservation Committee,
- 443 Peterborough, 266pp.
- 444 21. Cope, J.C.W. 2006. Jurassic: the returning seas. In Brenchley, P.J., and Rawson, P.F.,

445	(Ed.) The geology of England and Wales. 2nd ed. London: Geological Society 559pp.
446	22. Foffa D., Young, M.T., and Brusatte, S.L. 2018. Filling the Corallian gap: new
447	information on Late Jurassic marine reptile faunas from England. Acta Palaeontologica
448	Polonica 63(2).
449	23. Cecca, F., Garin, B. M., Marchand, D., Lathuiliere, B., and Bartolini, A. 2005.

- 450 Paleoclimatic control of biogeographic and sedimentary events in Tethyan and peri-
- 451 Tethyan areas during the Oxfordian (Late Jurassic). *Palaeogeography, Palaeoclimatology,*452 *Palaeoecology* 222(1–2): 10–32.
- 453 24. Armstrong, H.A., Wagner, T., Herringshaw, L.G., Farnsworth, A.J., Lunt, D.J., Harland,
- 454 M., Imber, J., Loptson, C., and Atar, E.F.L. 2016. Hadley circulation and precipitation
- 455 changes controlling black shale deposition in the Late Jurassic Boreal Seaway.

456 *Paleoceanography* 31: 1041–1053. doi:10.1002/2015PA002911.

- 457 25. Cox, B.M., Hudson, J.D., and Martill, D.M. 1992. Lithostratigraphic nomenclature of the
- 458 Oxford Clay (Jurassic). *Proceedings of the Geologists' Association* 103: 343–345.
- 459 26. Mettam, C., Johnson, A.L.A, Nunn, E.V. and Schöne, B.R. 2014. Stable isotope ( $\delta^{18}$ O and
- 460  $\delta^{13}$ C) sclerochronology of Callovian (Middle Jurassic) bivalves (*Gryphaea*
- 461 (*Bilobissa*) *dilobotes*) and belemnites (*Cylindroteuthis puzosiana*) from the Peterborough
- 462 Member of the Oxford Clay Formation (Cambridgeshire, England): Evidence of
- 463 palaeoclimate, water depth and belemnite behaviour. *Palaeogeography*,
- 464 *Palaeoclimatology, Palaeoecology* 399: 187-201.
- 465 https://doi.org/10.1016/j.palaeo.2014.01.010
- 466 27. Dromart, G., Garcia, J.P., Gaumet, F., Picard, S., Rousseau, M., Atrops, F., Lecuyer, C.,
- 467 and Sheppard, S.M.F. 2003a. Perturbation of the carbon cycle at the Middle–Late Jurassic
- 468 transition: geological and geochemical evidence. *American Journal of Science* 303: 667–
- 469 707.

- 470 28. Dromart, G., Garcia, J.P., Picard, S., Atrops, F., Lécuyer, C., and Sheppard, S.M.F. 2003b.
- 471 Ice age at the Middle–Late Jurassic transition? *Earth and Planetary Science Letters* 213:
  472 205–220
- 473 29. Gallois, R.W. 2004. The Kimmeridge Clay: the most intensively studied formation in
- 474 Britain. Open University Geological Journal 25: 33–38.
- 475 30. Haq, B.U., Hardenbol, J., and Vail, P.R., 1988, Mesozoic and Cenozoic
- 476 chronostratigraphy and cycles of sea-level change, in *Sea Level Changes An Integrated*
- 477 Approach, eds. C.K. Wilgus, B.S. Hastings, C.A. Ross, H. Posamentier, J. Van Wagoner
- 478 and C.G.S.C. Kendall: SEPM Spec. Publ., v. 42, p. 71-108.
- 479 31. Pauly, D., Trites, A.W., Capuli, E., and Christensen, V. 1998. Diet composition and
- 480 trophic levels of marine mammals. *ICES Journal of Marine Science* 55: 467–481.
- 481 (doi:10.1006/jmsc.1997.0280)
- 482 32. Esteban, R., Verborgh, P., Gauffier, P., Giménez, J., Afán, I., Cañadas, A., García, P.,
- 483 Murcia, J., Magalhães, S., Andreu, E., and de Stephanis, R., 2013. Identifying key habitat
- 484 and seasonal patterns of a critically endangered population of killer whales. *Journal of the*
- 485 *Marine Biological Association of the United Kingdom* 94: 1317–1325.
- 486 33. Forney, K.A. 2000. Environmental models of cetacean abundance: reducing uncertainty
- 487 in population trends. *Conservation Biology* 14:1271–1286.
- 488 34. Yen, P.P.W., Sydeman, W.J., and Hyrenbach, K.D. 2004. Marine birds and cetacean
- 489 associations with bathymetric habitats and shallow-water topographies:implications for
- 490 trophic transfer and conservation. *Journal of Marine Systems* 50: 79–99.
- 491 35. Balance, L.T., Pitman, R.L., and Fiedler, P.C. 2006. Oceanographic influences on
- 492 seabirds and cetaceans in the eastern tropical Pacific: A review. *Progress in*
- 493 *Oceanography* 69 (2–4): 360–390.
- 494 36. MacLoed, C.D., Weit, C.R., Pierpoint, C, and Harland, E.J. 2007. The habitat preferences

- 495 of marine mammals west of Scotland (UK). *Journal of the Marine Biological Association*496 *of the United Kingdom* 87: 157–164.
- 497 37. Spitz, J. Cherel, Y., Bertin, S., Kiszka, J., Dewez, A. and Ridoux V. 2011. Prey
- 498 preferences among the community of deep-diving odontocetes from the Bay of Biscay,
- 499 Northeast Atlantic. *Deep Sea Research Part I: Oceanographic Research Papers* 58(3):
- 500 273–282.
- 501 38. Weir, C.R., MacLeod, C.D., and Pierce, G.J. 2012. Habitat preferences and evidence for
- 502 niche partitioning amongst cetaceans in the waters between Gabon and Angola, eastern
- 503 tropical Atlantic. *Journal of the Marine Biological Association of the United Kingdom*

504 92(8): 1735–1749.

- 505 39. Roberts, J.J., Best, B.D., Mannocci, L., Fujioka, E., Halpin, P.N., Palka, D.L., Garrison,
- 506 L.P., Mullin, K.D., Cole, T.V.N., Khan, C.B., McLellan, W.A., Pabst, D.A., and Lockhart,
- 507 G.G. 2016. Habitat-based cetacean density models for the U.S. Atlantic and Gulf of
- 508 Mexico. Scientific Reports 6, doi:10.1038/srep22615
- 509 40. Fossete, S. Abrahams, B., Hazen, E.L., Bogard, S.J., Zilliacus, K.M., Calambokidis, J.,
- 510 Burrows, J.A., Goldbogen, J.A., Harvey, J.T., Marinovic, B., Tershy, B., and Croll, D.A.
- 511 2017. Resource partitioning facilitates coexistence in sympatric cetaceans in the
- 512 California Current. *Ecology and Evolution* doi: 10.1002/ece3.3409.
- 513 41. Larson, D.W., Brown, C.M., and Evans, D.C. 2016. Dental Disparity and Ecological
- 514 Stability in Bird-like Dinosaurs prior to the End-Cretaceous Mass Extinction. *Current*
- 515 *Biology* 26(10): 1325–1333. doi: 10.1016/j.cub.2016.03.039.
- 516 42. Smith, J.B., Vann, D.R., and Dodson, P. 2005. Dental morphology and variation in
- 517 theropod dinosaurs: Implications for the taxonomic identification of isolated teeth. The
- 518 Anatomical Record Part A Discoveries in Molecular Cellular and Evolutionary Biology
- 519 285(2):699-736. 10.1002/ar.a.20206

- 43. Noè, L.F. 2001. A taxonomic and functional study of the Callovian (Middle Jurassic)
  Pliosauroidea (Reptilia, Sauropterygia). Unpublished PhD thesis, University of Derby,
  UK
- 523 44. Sassoon, J., Foffa, D., and Marek, R. 2015. Dental ontogeny and replacement in
- 524 Pliosauridae. *Royal Society Open Science* 2 150384. DOI: 10.1098/rsos.150384.
- 525 45. Foffa, D., Young, M.T., Brusatte, S.L., Graham, M.R., and Steel, L. 2017. A new
- 526 metriorhynchid crocodylomorph from the Oxford Clay Formation (Middle Jurassic) of
- 527 England, with implications for the origin and diversification of Geosaurini. *Journal of*

528 Systematic Palaeontology 10.1080/14772019.2017.1367730

- 529 46. Young, M.T., Hastings, A.K., Allain, R., Smith, T.J. 2016. Revision of the enigmatic
- 530 crocodyliform *Elosuchus felixi* de Lapparent de Broin, 2002 from the Lower–Upper
- 531 Cretaceous boundary of Niger: potential evidence for an early origin of the clade
- 532 Dyrosauridae. Zoological Journal of the Linnean Society doi:10.1111/zoj.12452
- 47. Schneider, C.A., Rasband, W.S., and Eliceiri, K.W. 2012. NIH Image to ImageJ: 25 years
  of image analysis. *Nature Methods* 9: 671–675.
- 535 48. Anderson, P.S.L., Friedman, M., Brazeau, M.D., and Rayfield, E.J. 2011. Initial radiation
- of jaws demonstrated stability despite faunal and environmental change. *Nature* 476:
  206–209.
- 49. Gower, J.C. 1971. A general coefficient of similarity and some of its properties. *Biometrics* 27: 857–871.
- 540 50. Hammer, Ø., Harper, D.A.T., and Ryan, P.D. 2001. PAST: Paleontological statistics
- 541 software package for education and data analysis. *Palaeontologia Electronica* 4(1): 1–9.
- 542 51. Anderson, M.J. 2001. A new method for non-parametric multivariate analysis of
- 543 variance. *Austral Ecology* 26(1): 32–46. <u>doi:10.1111/j.1442-9993.2001.01070.pp.x</u>
- 544 52. R Core Team. 2012. R: A language and environment for statistical computing. R

- 545 Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0,
- 546 URL <u>http://www.R-project.org/</u>
- 547 53. Benjamini, Y., and Hochberg, Y. 1995. Controlling the false discovery rate: a practical
- and powerful approach to multiple testing. *Journal of the Royal Statistical Society, Series B.* 57(1): 289–300.
- 550 54. Wills, M.A., Briggs, D.E.G., and Fortey, R.A. 1994. Disparity as an evolutionary index: a
  - 551 comparison of Cambrian and Recent arthropods. *Paleobiology* 20: 93–130.
  - 552 55. Guillerme, T. 2016. dispRity: a package for measuring disparity in R. Zenodo.
  - 553 10.5281/zenodo.55646
  - 554 56. Brusatte, L.S., Lloyd, G.T., Wang, S.C., and Norell, M.A. 2014. Gradual assembly of
  - avian body plan culminated in rapid rates of evolution across the dinosaur-bird transition.

556 Current Biology 20: 2386–2392.

- 557 57. Foote, M. 1993. Contributions of individual taxa to overall morphological disparity.
- 558 *Paleobiology* 19: 403–419.
- 559 58. Navarro, N. 2003. MDA: a MATLAB-based program for morphospace-disparity analysis.
- 560 *Computers & Geosciences* 29: 655–664.

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#### 574 Author contributions

575 DF led the project, conceived the study and wrote the initial draft manuscript. MTY and SLB 576 helped develop the project, edited drafts, and provided guidance on statistical analyses. SLB 577 wrote the final manuscript, which was revised by all authors. DF designed and performed the 578 analyses with technical support from TLS and KGD.

### 579 Competing interests

580 The authors declare no competing interests.

#### 581 Figure Captions

Figure 1. Morphospace plots showing the distribution of marine reptile specimens based on
tooth morphology. a. 3D plot of PCO1-PCO2-PCO3, derived from Principal Coordinates
Analysis of our dental dataset; b. approximate position of the 'feeding guilds' (originally
outlined by Massare<sup>7</sup> and modified here) in the PCO morphospace. Tooth drawings depict the
general tooth morphologies of key regions of the morphospace.

587

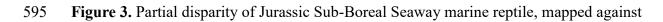
588 Figure 2. Morphospace plots showing the distribution of marine reptile clades based on tooth

589 morphology through time, only statistically significant ecological and evolutionary shifts are

- 590 noted. The morphospaces are 3D plots of PCO1-PCO2-PCO3, derived from Principal
- 591 Coordinates Analysis (Fig. 1). The radiations and declines of groups through time gradually

changed the morphospace/ecosystem composition and partitioning. Note that some groups
may have been limited to certain parts of ecomorphospace by evolutionary constraints.

594



- 596 global sea-level. **a.** Partial disparity of taxonomic groups, **b.** Partial disparity of dietary
- 597 guilds; **c.** sea-level curve (modified from Haq 1987)<sup>30</sup>, with fossil rich-intervals from the Sub-
- 598 Boreal Seaway noted.