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Large-scale cognitive GWAS meta-analysis reveals tissue-specific neural expression and potential nootropic drug targets

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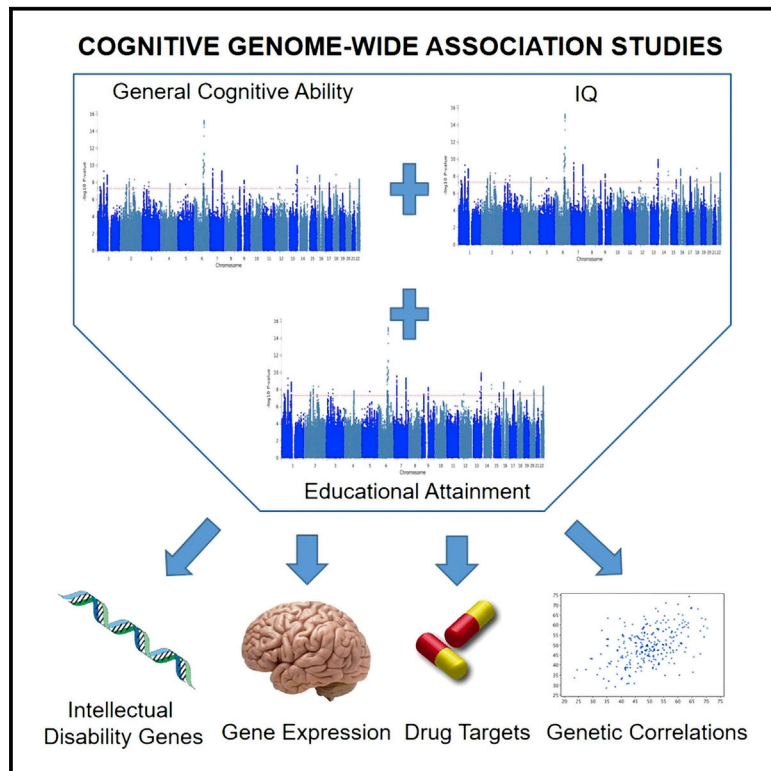
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Cell Reports

Large-Scale Cognitive GWAS Meta-Analysis Reveals Tissue-Specific Neural Expression and Potential Nootropic Drug Targets

Graphical Abstract



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In Brief

Lam et al. conduct a large-scale genome-wide association study of cognitive ability, identifying 70 associated loci. Results provide biological insights into the molecular basis of individual differences in cognitive ability, as well as their relationship to psychiatric and other health-relevant phenotypes.

Highlights

- Large-scale GWAS of cognitive performance, combined with GWAS of educational attainment
- 70 independent genomic loci associated with individual differences in cognition
- Implicated genes suggest potential treatment targets for cognitive enhancement
- Genetic overlap between cognitive ability and multiple health-related phenotypes



Large-Scale Cognitive GWAS Meta-Analysis Reveals Tissue-Specific Neural Expression and Potential Nootropic Drug Targets

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SUMMARY

Here, we present a large (n = 107,207) genome-wide association study (GWAS) of general cognitive ability (“g”), further enhanced by combining results with a large-scale GWAS of educational attainment. We identified 70 independent genomic loci associated with general cognitive ability. Results showed significant enrichment for genes causing Mendelian disorders with an intellectual disability phenotype. Competitive pathway analysis implicated the biological processes of neurogenesis and synaptic regulation, as well as the gene targets of two pharmacologic agents: cinnarizine, a T-type calcium channel blocker, and LY97241, a potassium channel inhibitor.

Transcriptome-wide and epigenome-wide analysis revealed that the implicated loci were enriched for genes expressed across all brain regions (most strongly in the cerebellum). Enrichment was exclusive to genes expressed in neurons but not oligodendrocytes or astrocytes. Finally, we report genetic correlations between cognitive ability and disparate phenotypes including psychiatric disorders, several autoimmune disorders, longevity, and maternal age at first birth.

INTRODUCTION

Genome-wide association studies (GWASs) have been highly successful at uncovering hundreds of genetic loci associated



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with heritable quantitative traits such as height (Wood et al., 2014) and weight/body mass index (BMI) (Locke et al., 2015). However, identifying genetic loci underlying cognitive ability has been much more challenging, despite heritability of 0.5 or greater, as determined by both classical twin studies (Deary et al., 2009) and molecular genetic studies (Davies et al., 2011a). In part, the difficulty with cognitive GWASs may be caused by the relative heterogeneity in the measurement of the cognitive phenotype. Traditionally, general cognitive ability (*g*) has been defined as a latent trait underlying shared variance across multiple subdomains of cognitive performance, psychometrically obtained as the first principal component of several

distinct neuropsychological test scores (Johnson et al., 2008). Using this approach, several cognitive GWASs with fewer than 20,000 subjects yielded no genome-wide significant (GWS) effects (Benyamin et al., 2013; Davies et al., 2011b; Lencz et al., 2014), while a few GWS loci were identified in larger GWAS of 35,298 (Trampush et al., 2017) and 53,949 (Davies et al., 2015) subjects, respectively. By contrast, two independent GWASs of height with sample sizes of approximately 30,000 subjects each yielded 20–30 GWS hits (Gudbjartsson et al., 2008; Weedon et al., 2008). Allelic effect sizes were ~2–5 times larger than the largest obtained in cognitive GWASs (Trampush et al., 2017).

Very recently, a cognitive GWAS (Sniekers et al., 2017) was able to leverage a very brief measure of fluid intelligence, highly correlated with psychometrically defined g , obtained in over 50,000 subjects. In combination with several traditional cognitive GWAS cohorts, total sample size was 78,308. This sample size permitted discovery of 18 independent GWS allelic loci, as well as numerous additional loci from gene-based analysis. This report was critical in demonstrating that signal could be enhanced by combining data from cohorts with brief measures of intelligence with data from more traditional cognitive GWASs.

A further approach to enhancing power in cognitive GWASs has focused on educational attainment as a proxy phenotype (Rietveld et al., 2014). It is acknowledged that this phenotype is “noisy”, as it is influenced by non-cognitive genetic (e.g., personality; Belsky et al., 2016) and environmental (e.g., socio-economic; Johnson et al., 2010) factors; consequently, observed allelic effect sizes have been even smaller than those obtained for GWASs of g (Rietveld et al., 2013). However, by utilizing a single-item measure (years of education completed), obtained incidentally in large studies of other phenotypes, this approach has allowed investigators to obtain extremely large sample sizes. A recent study of educational attainment in nearly 300,000 individuals identified 74 independent GWS loci (Okbay et al., 2016). Moreover, a new technique called multi-trait analysis of GWAS (MTAG) (Turley et al., 2017) has been developed which permits integration of GWAS data across related traits, accounting for the possibility of overlapping samples across studies and requiring only summary statistics. The developers of MTAG demonstrated its accuracy and utility in a study of traits (depression, neuroticism, and subjective well-being) that demonstrate genetic correlations in the range of $\sim .70$ – $.75$; importantly, the genetic correlation between cognitive performance and educational attainment has been consistently reported to be in the same range (Davies et al., 2015, 2016; Okbay et al., 2016; Trampush et al., 2017; Sniekers et al., 2017). MTAG is able to quantify the degree of “boost” to the signal of a single-trait GWAS, providing an estimate of observed sample size and providing summary statistics (allelic weights) that can then be utilized in all downstream annotation pipelines available for GWAS output.

In the present study, we first utilized GWAS meta-analysis to combine our prior Cognitive Genomics Consortium (COGENT) consortium GWAS (Trampush et al., 2017) of psychometrically defined g with the recently reported GWAS (Sniekers et al., 2017), relying primarily on the brief measure, resulting in a combined cohort of $n = 107,207$ non-overlapping samples measured for cognitive performance. Next, we utilized MTAG to combine these results with the large-scale GWAS of educational attainment, resulting in further enhanced power. At each step, we performed both allelic and gene-based tests. We then performed downstream analyses on the resulting MTAG summary statistics, including: (1) competitive gene set analyses to identify key biological processes and potential drug targets implicated, (2) stratified linkage disequilibrium score regression (LDSC) to identify differential cell type expression, (3) transcriptome-wide association study (TWAS) methods, to identify specific effects of altered gene expression in the brain on cognition, and (4) LDSC to identify genetic correlations with other anthropometric and biomedical phenotypes.

RESULTS

Meta-Analysis: Cognitive Performance GWASs

Meta-analysis of all non-overlapping cohorts from the two GWASs of cognitive performance (total $n = 107,207$) identified 28 independent genomic loci reaching genome-wide significance (GWS, $p < 5E-8$) using default clumping parameters from the Functional Mapping and Annotation (FUMA) pipeline (Watanabe et al., 2017; Figure 1A), representing a 55.6% increase in loci compared to the previous GWAS (Sniekers et al., 2017) of cognitive performance. Two of these loci each contained two uncorrelated variants with independent effects, resulting in 30 independent lead SNPs. Evidence for spurious inflation of statistical tests was quite limited for a large study of a highly polygenic trait ($\lambda = 1.23$; $\lambda_{1000} = 1.001$; linkage disequilibrium (LD) score intercept = 1.03; see also PP plot in Figure S1), and overall SNP heritability was 0.168. Of the 28 GWS loci, 12 were not previously reported as GWS in published studies of cognitive or educational phenotypes (Table S1). The majority of the 5,610 markers reaching a nominal significance threshold were intronic SNPs followed by those in the intergenic regions (Table S2). As shown in Table S3, several of the GWS loci overlap with loci related to schizophrenia, bipolar disorder, and other neuropsychiatric phenotypes, as well as obesity/BMI and other traits.

The significant loci harbored 88 known protein-coding genes (Table S4), about half of which were in three large regions (Figure S2), including two well-characterized regions: the distal 16p11.2 region, in which deletions have been associated with schizophrenia and other neuropsychiatric phenotypes (Guha et al., 2013), and the 17q21 region, in which inversions have been associated with neuropsychiatric disorders (Cooper et al., 2011). Using MAGMA (Multi-marker Analysis of GenoMic Annotation; de Leeuw et al., 2015) gene-based tests, 73 genes were genome-wide significant (Table S5), of which 39 were overlapping with the 88 genes noted above, resulting in a total of 122 candidate genes with statistical evidence of association to cognitive performance.

MTAG: Combining Cognitive Performance and Educational Attainment GWASs

MTAG analysis combining the cognitive performance results obtained above with the large educational attainment GWAS previously reported (Okbay et al., 2016), resulted in a 75% enrichment of statistical power, effectively boosting the original sample size of $n = 107,207$ to a GWAS equivalent of $n = 187,812$. Default clumping procedures revealed that 70 independent genomic loci reached genome-wide significance, with 82 independent SNPs (Figure 1B). Similar to the GWAS results above, the PP plot (Figure S3) demonstrated polygenicity without evidence for artifactual inflation of statistical tests ($\lambda = 1.28$; $\lambda_{1000} = 1.001$; LD score intercept = 0.91), and overall SNP heritability was 0.336. Of the 70 GWS loci, 34 were not previously reported as GWS in published studies of cognitive or educational phenotypes (Figure 2; Table S1). All but two of the 30 loci identified in the meta-analysis remained genome-wide significant in the MTAG results. Even these two loci showed the same direction of allelic effects between cognitive meta-analytic GWASs and

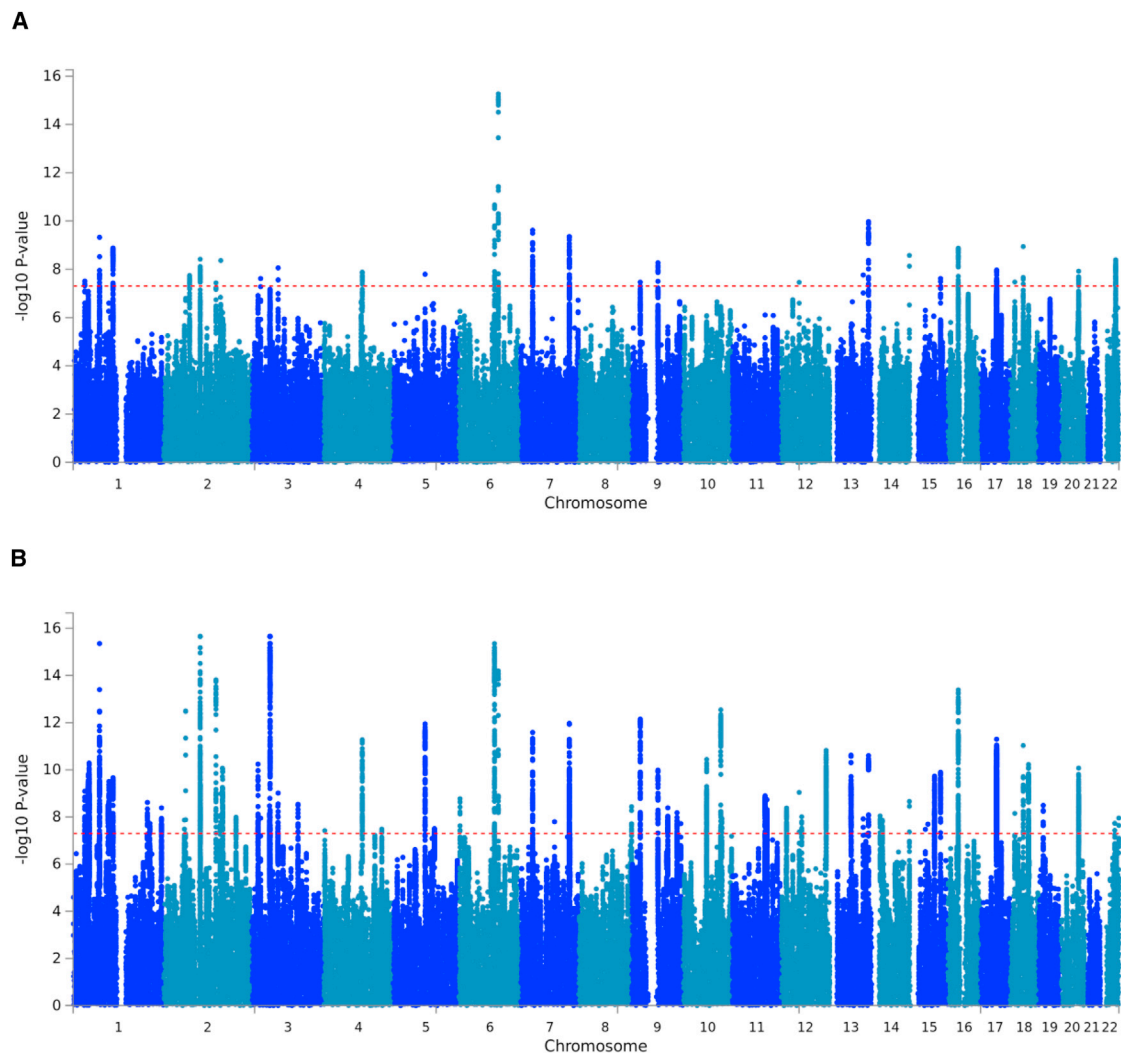


Figure 1. Manhattan Plots for GWAS Meta-Analysis and MTAG Analysis

(A) Manhattan plot depicting results of GWAS meta-analysis of cognitive performance. Dotted red line indicates threshold for genome-wide significance ($p < 5E-08$).

(B) Manhattan plot depicting results of MTAG of cognitive performance with educational attainment. Dotted red line indicates threshold for genome-wide significance ($p < 5E-08$).

the educational GWASs. The majority of the 13,549 SNPs reaching a nominal significance threshold in the MTAG analysis were intergenic or intronic (Table S2; Figure S4). GWAS catalog annotations are listed in Table S3. Within the GWS loci, 265 protein-coding genes were identified (Table S4). Additionally, 256 genes were significant in MAGMA gene-based tests (Table S6). Of these genes, 85 were non-overlapping with the 265 genes within SNP GWS loci, resulting in a total of 350 genes receiving GWS support from the MTAG results.

As a formal validation that the MTAG methodology successfully predicts phenotype variance for cognitive performance, MTAG was re-analyzed, excluding the COGENT cohorts (i.e., the IQ GWAS of Sniekers et al., 2017 was combined with the educational GWAS of Okbay et al. 2016). The ASPIS (Athens Study of Psychosis Proneness and Incidence of Schizophrenia)

and GCAP (NIMH Genes, Cognition and Psychosis Program) datasets were held out as target cohorts used for calculation of polygenic risk score modeling for “g.” Despite the relatively small size of these hold-out cohorts, results show strongly significant polygenic prediction of “g” using MTAG-derived allele weights (Figure 3A and 3C), accounting for more than 4% of the variance in the GCAP cohort. For both cohorts, polygenic prediction began to drop at P_T thresholds above 0.05, suggesting that there may be some degree of saturation of signal beyond the nominal 0.05 significance level at these sample sizes. Additional comparisons were made with IQ-only predictions (weights derived from Sniekers et al., 2017) and education-only predictions (weights derived from Okbay et al., 2016) for the same hold-out cohorts (Figure 3B and 3D), and we found that the MTAG-derived weights showed a 3.5 times and 3 times

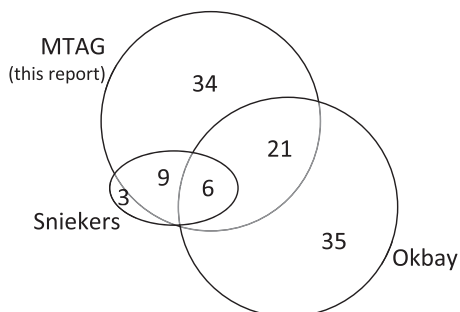


Figure 2. Overlap of Genome-wide Significant Loci in this Study with Other Recent Reports

Venn diagram depicting overlap and independence of genome-wide significant SNP loci observed in three studies: the MTAG analysis of the present report, the cognitive performance GWAS reported by Sniekers et al. (2017), and the educational attainment GWAS of Okbay et al. (2016).

improvement in R^2 variance explained in the ASPIS cohort, for IQ and education, respectively. For the GCAP cohort, there was a 5.1 times to 96 times improvement in R^2 variance relative to IQ or education alone.

Overlap with Intellectual Disability Genes

We compared the list of 350 genes emerging from MTAG with a list of 621 genes known to cause autosomal dominant or autosomal recessive Mendelian disorders featuring intellectual disability (Harripaul et al., 2017; Vissers et al., 2016). As shown in Table 1, a total of 23 genes identified by MTAG appeared on this list, representing a 2-fold enrichment over chance (hypergeometric probability $p = 0.001$). Examining autosomal dominant and recessive Mendelian genes demonstrated a somewhat stronger enrichment for autosomal dominant genes ($p = 0.0017$) than autosomal recessive genes ($p = 0.054$).

Tissue Expression Enrichment and Competitive Pathway Analysis

Downstream MAGMA expression profiles and competitive pathway analysis were conducted as part of the FUMA pipeline. MAGMA tissue expression profile analysis revealed that genes emerging from the MTAG analysis were significantly enriched for expression in nearly all central nervous system tissues (except for substantia nigra and spinal cord) and that this enrichment was exclusive to neural tissues (Figure 4A). Notably, the strongest enrichment was observed for genes expressed in the cerebellum, followed by the cortex, and slightly weaker (but still strongly significant) enrichment in subcortical and limbic structures. Competitive pathway analysis (based on gene ontology categories) for GWS MAGMA genes identified by MTAG revealed significant enrichment of neuronal and synaptic cellular components, as well as the biological processes of neurogenesis and regulation of synapse organization (Table 2, top). Because three MTAG loci (at chromosome 3q21.31, 16p11.2, and 17q21.31) were unusually large, each containing ≥ 15 genes that may have disproportionately impacted enrichment results, we re-ran the above tissue expression and pathway analyses excluding these three regions. Results were substantively un-

changed: all of the same neural tissues remained significantly enriched, in the same order of significance as shown in Figure 4A, and all of the same pathways remained significant (Bonferroni-corrected $p < .05$) as shown in Table 2, except for the cellular compartment “dendrite” (Bonferroni-corrected $p = 0.089$).

Competitive pathway analysis for drug pathways (Gaspar and Breen, 2017) revealed that the gene targets of two drugs were significantly enriched in the MTAG results (Table 2, bottom): Cinnarizine, a T-type calcium channel blocker, and LY97241, a potassium channel inhibitor. L-type calcium channel blockers and anti-inflammatories also showed suggestive evidence of enrichment. In a related analysis of drug classes, significant enrichment was observed for voltage-gated calcium channel subunits ($p = 9.28E-06$, Bonferroni-corrected $p = 5.38E-04$).

Stratified LD score regression (Finucane et al., 2017) also demonstrated an enrichment of cell type expression for neuronal tissues only. Notably, genes found in the neuronal expression list of Cahoy et al. (2008) were significantly enriched ($p = 0.0129$; Bonferroni-corrected $p = 0.0386$), whereas negative results were obtained for genes expressed in oligodendrocytes ($p = 0.4997$) and astrocytes ($p = 0.9057$). Additionally, using Roadmap annotations, epigenetic enrichment was strongest in fetal brain tissue DNase sites and H3K4me1 primed enhancers, followed by adult cortical H3K27ac active enhancer sites (see Table S7 for further details). No enrichment was observed for any non-neuronal tissue. Again, results were not substantively changed when the three large loci were removed from these analyses.

Gene Expression Analyses

In order to derive specific biological insights from the broad association loci implicated by MTAG, we performed a series of analyses designed to identify individual gene expression changes associated with cognition. First, we performed transcriptome wide analysis (TWAS) using MetaXcan (Barbeira et al., 2016) on MTAG SNP results in order to identify transcripts for which upregulation or downregulation in specific neural compartments was associated with cognition. Note that TWAS follows a similar logic to imputation, in that an external reference (in this case, publicly available GTEx eQTL data for 10 brain regions) is utilized to link SNP-based summary statistics to tissue-based expression levels. As shown in Figure 4B (and detailed in Table S8), most of the significant TWAS results are expressed across all neural tissues, involving genes such as *AMIGO3*, *RNF123*, and *RBM6*. Moreover, no individual tissue compartment was much more strongly enriched for associations compared to the others. However, a few strong transcriptomic associations were specific to individual brain regions. For example, the strongest result in hippocampus was with *DAG1*. TWAS demonstrated that greater expression of this gene in the hippocampus was associated with higher cognitive scores. However, this gene was not expressed in other neural tissue types in the Genotype-Tissue Expression (GTEx) database. Similarly, lower levels of *ACTR1A* were significantly associated with better cognition, but this transcript was observed only in the frontal cortex.

Second, we applied a Bayesian fine-mapping approach (CAVIAR-BF; Chen et al., 2015) to identify putative causal SNPs within each associated locus, as defined in Table S9.

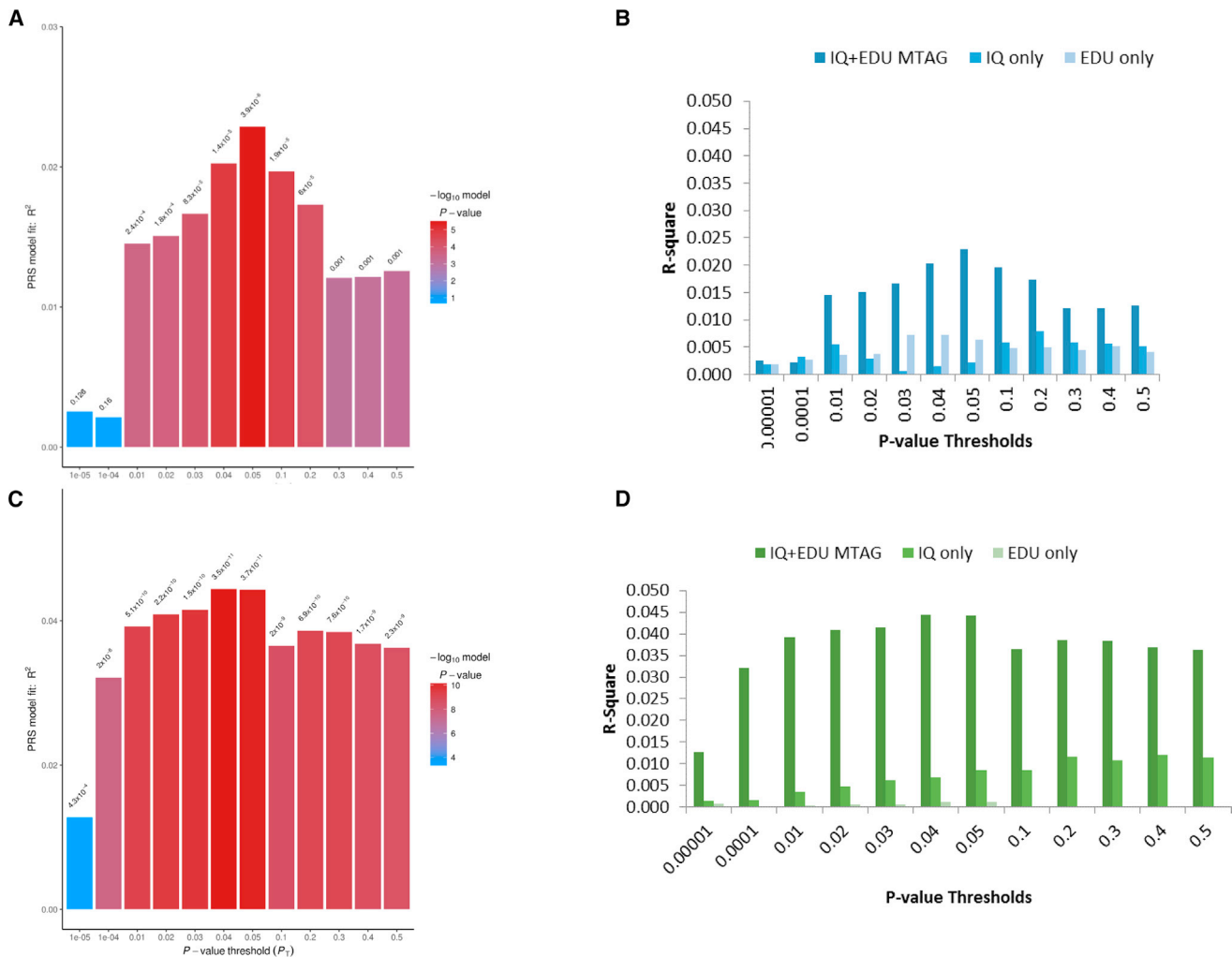


Figure 3. Leave-One-Out Analyses of Polygenic Risk Scores

(A) Polygenic risk score prediction for MTAG results against held-out ASPIS cohort.
 (B) Comparison of MTAG, cognitive (IQ) GWAS (Sniekers et al., 2017), and educational attainment (EDU) GWAS (Okbay et al., 2016) as source of weights for polygenic risk score prediction against held-out ASPIS cohort.
 (C) Polygenic risk score prediction for MTAG results against held-out GCAP cohort.
 (D) Comparison of MTAG, cognitive (IQ) GWAS (Sniekers et al., 2017), and educational attainment (EDU) GWAS (Okbay et al., 2016) as source of weights for polygenic risk score prediction against held-out GCAP cohort.

CAVIAR-BF revealed that there was strong evidence ($BF = 3.71E+2$) for at least 1 causal SNP within each of the 70 independent MTAG loci. There is also evidence that there are at least 2 causal SNPs in 65 of the loci ($BF = 3E+6$) and at least 3 causal SNPs in 47 of the loci ($BF = 2.86E+6$). In the extended region analysis, there was evidence for at least 1 causal SNP ($BF = 3.45E+2$) and 2 causal SNPs ($BF = 2.89E+6$) for 70 and 63 loci, respectively. Model search revealed that there were 386 putative causal SNPs within the 70 independent loci (Table S10). Lookups of these SNPs in two brain expression quantitative trait loci (eQTL) databases (BrainEAC [Ramasamy et al., 2014] and CommonMind [Haugberg et al., 2017]) revealed several additional SNP-eQTL relationships that can explain variance in the cognitive phenotype (Tables S11 and S12). The most notable eQTL ef-

fect was observed for rs3809912 on chromosome 18. This SNP, which was GWS in the MTAG results ($p = 7.06E-09$), was a strong eQTL for *CEP192* ($p = 5.1E-38$, $FDR < 0.01$). This eQTL was confirmed in the CommonMind database ($FDR < .01$), which demonstrated that expression of 44 independent transcripts in the frontal cortex were significantly associated with MTAG SNPs at the $FDR < .01$ level. Combining annotation information from the Mendelian gene analysis, MetaXcan TWAS, Braineac, and CommonMind databases, we found supporting functional evidence for 112 of the 350 candidate genes nominated by MTAG (Table S13). The remaining 238 genes without functional support had statistical evidence for association to cognition but are considered to be “candidate genes” requiring further functional or experimental support.

Table 1. List of Candidate Genes Emerging from MTAG Analysis Associated with Mendelian Disorders Featuring an Intellectual Disability Phenotype

GENE	CHR	START	MAGMA P	Min MTAG P	OMIM	Mode	Phenotype
<i>AFF3</i>	2	100152323	6.53E-12	6.8834E-15	NA	AR	nonsyndromal intellectual disability
<i>AMT</i>	3	49444211	1.74E-09	8.5543E-09	605899	AR	glycine encephalopathy
<i>ARFGF2</i>	20	47528427	7.28E-10	4.1558E-10	608097	AR	periventricular heterotopia with microcephaly
<i>BCL11A</i>	2	60668302	8.5E-12	3.2174E-13	617101	AD	intellectual developmental disorder with persistence of fetal hemoglobin
<i>C12orf65</i>	12	123707463	1.48E-10	1.8088E-11	613559	AR	combined oxidative phosphorylation deficiency 7
<i>C12orf65</i>	12	123707463	1.48E-10	1.8088E-11	615035	AR	spastic paraplegia 55
<i>CLN3</i>	16	28467983	2.31E-08	1.9502E-08	204200	AR	ceroid lipofuscinosis, neuronal 3
<i>DPYD</i>	1	97533299	0.005108	4.4603E-08	274270	AR	dihydropyrimidine dehydrogenase deficiency
<i>DPYD</i>	1	97533299	0.005108	4.4603E-08	274270	AR	5-fluorouracil toxicity
<i>ERCC8</i>	5	60159658	2.96E-07	5.5002E-7	216400	AR	cockayne syndrome, Type A
<i>ERCC8</i>	5	60159658	2.96E-07	5.5002E-7	614621	AR	UV-sensitive syndrome 2
<i>FOXP1</i>	3	70993844	6.32E-07	3.5007E-09	613670	AD	mental retardation with language impairment and autistic features
<i>GMPPB</i>	3	49744277	1.75E-14	6.6613E-16	613530	AR	muscular dystrophy-dystroglycanopathy (congenital w/ brain, eye anomalies), type A, 14
<i>GMPPB</i>	3	49744277	1.75E-14	6.6613E-16	615351	AR	muscular dystrophy-dystroglycanopathy (congenital with mental retardation), type B, 14
<i>GMPPB</i>	3	49744277	1.75E-14	6.6613E-16	615352	AR	muscular dystrophy-dystroglycanopathy (limb-girdle), type C, 14
<i>KANSL1</i>	17	44097282	1.62E-08	5.0278E-12	610443	AD	Koolen-De Vries syndrome
<i>KCNH1</i>	1	210846555	1.04E-06	5.2513E-08	135500	AD	Zimmermann-Laband syndrome
<i>KMT2D</i>	12	49402758	1.69E-07	4.3422E-08	147920	AD	Kabuki syndrome, 1
<i>LARGE</i>	22	33548212	7.99E-07	5.4265E-07	613154	AR	muscular dystrophy-dystroglycanopathy (congenital w/ brain, eye anomalies), type A, 6
<i>LARGE</i>	22	33548212	7.99E-07	5.4265E-07	608840	AR	muscular dystrophy-dystroglycanopathy (congenital with mental retardation), type B, 6
<i>MEF2C</i>	5	88003975	1.74E-13	1.1304E-12	613443	AD	mental retardation, stereotypic movements, epilepsy, and/or cerebral malformations
<i>MEF2C</i>	5	88003975	1.74E-13	1.1304E-12	613443	AD	chromosome 5q14.3 deletion syndrome
<i>NFIX</i>	19	13096422	2.45E-06	5.3017E-09	602535	AD	Marshall-Smith syndrome
<i>NFIX</i>	19	13096422	2.45E-06	5.3017E-09	614753	AD	Sotos syndrome
<i>PDE4D</i>	5	58254865	9.13E-08	3.6537E-07	614613	AD	Acrodysostosis 2 with or without hormone resistance
<i>SHANK3</i>	22	51102843	2.7E-10	8.0006E-08	606232	AD	Phelan-McDermid syndrome
<i>ST3GAL3</i>	1	44161495	3.58E-13	1.6388E-10	611090	AR	mental retardation, autosomal recessive 12
<i>SUOX</i>	12	56380964	3.07E-05	4.1129E-08	272300	AR	sulfite oxidase deficiency
<i>TCF4</i>	18	52879562	1.02E-06	3.5713E-05	610954	AD	Pitt-Hopkins syndrome
<i>THRB</i>	3	24148651	0.000682	4.6883E-06	188570	AD	thyroid hormone resistance
<i>THRB</i>	3	24148651	0.000682	4.6883E-06	274300	AR	thyroid hormone resistance, autosomal recessive
<i>UBA7</i>	3	49832640	2.11E-13	6.6613E-16	NA	AR	nonsyndromal intellectual disability

AD, autosomal dominant; AR, autosomal recessive.

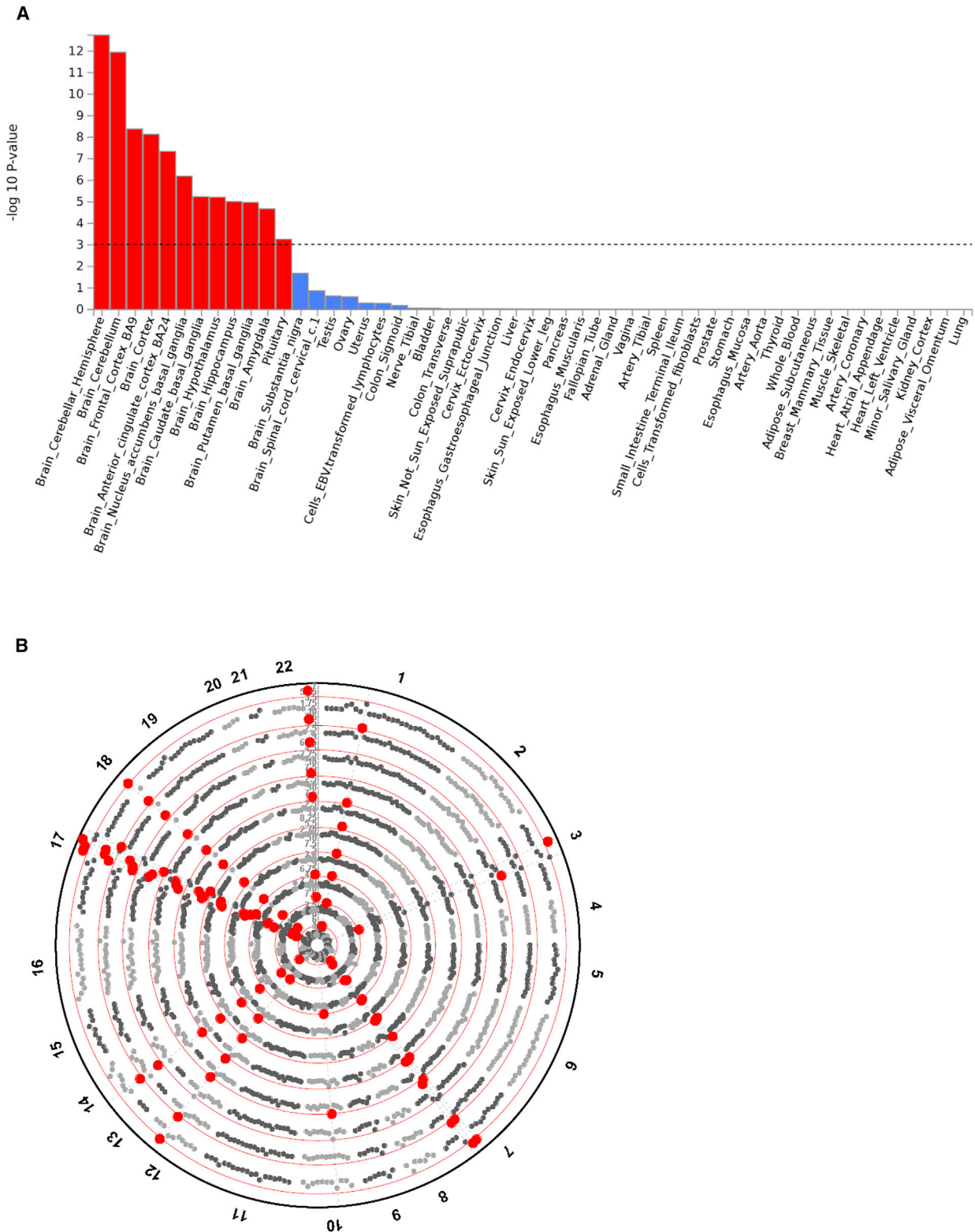


Figure 4. Tissue Expression and Transcriptome-wide Gene Expression Results

(A) Tissue expression profile analysis for genome-wide significant genes (as defined by MAGMA) emerging from the MTAG analysis. Gene results were significantly enriched for expression in nearly all central nervous system tissues (except for substantia nigra and spinal cord) but no tissues outside the central nervous system.

(legend continued on next page)

Table 2. Competitive Pathway Analyses of MTAG Results

GO Category Name	NGENES	BETA	BETA_STD	SE	P	Pbon
GO_cc:go_neuron_part	1204	0.155	0.0385	0.0304	1.84E-07	0.002008
GO_cc:go_neuron_projection	898	0.179	0.0388	0.0352	1.84E-07	0.002009
GO_bp:go_neurogenesis	1355	0.148	0.0388	0.0291	1.92E-07	0.002092
GO_cc:go_synapse	718	0.198	0.0386	0.0393	2.25E-07	0.002455
GO_cc:go_synapse_part	580	0.21	0.0369	0.0436	7.37E-07	0.008026
GO_cc:go_dendrite	430	0.229	0.0348	0.0501	2.49E-06	0.027087
GO_bp:go_regulation_of_synapse_organization	106	0.447	0.034	0.0987	2.94E-06	0.031982
GO_bp:go_regulation_of_synapse_structure_or_activity	223	0.291	0.032	0.0671	7.36E-06	0.080154
GO_bp:go_regulation_of_nervous_system_development	723	0.166	0.0325	0.0385	7.84E-06	0.085334
GO_bp:go_modulation_of_synaptic_transmission	291	0.253	0.0317	0.059	9.41E-06	0.102429
GO_bp:go_calcium_dependent_cell_cell_adhesion_via_plasma_membrane_cell_adhesion_molecules	26	1.06	0.0402	0.259	2.06E-05	0.224726
GO_cc:go_postsynapse	356	0.224	0.031	0.0553	2.64E-05	0.287583
GO_cc:go_neuron_spine	116	0.379	0.0302	0.0939	2.75E-05	0.299998
GO_cc:go_cell_projection	1710	0.103	0.0301	0.0258	3.36E-05	0.365381
GO_bp:go_regulation_of_cell_development	808	0.144	0.0297	0.0365	3.99E-05	0.434751
Drug Name	NGENES	BETA	BETA_STD	SE	P	Pbon
CINNARIZINE	9	1.62	0.036	0.355	2.61E-06	0.007071
LY97241	2	3.65	0.0382	0.842	7.59E-06	0.020535
CELECOXIB	45	0.632	0.0314	0.159	3.49E-05	0.094545
ISRADIPINE	8	1.59	0.0334	0.404	4.18E-05	0.11317
NITRENDIPINE	12	1.19	0.0305	0.323	1.19E-04	0.323151
ABT-639;ML218;TTA-A2;Z944	3	2.31	0.0297	0.641	1.59E-04	0.429388
NEUREGULIN-1;NEUREGULIN-2	2	2.39	0.0251	0.669	1.75E-04	0.473469
FLUNARIZINE	6	1.58	0.0287	0.457	2.67E-04	0.723503
GLUCOCORTICOIDS	2	3.68	0.0386	1.08	3.22E-04	0.872117

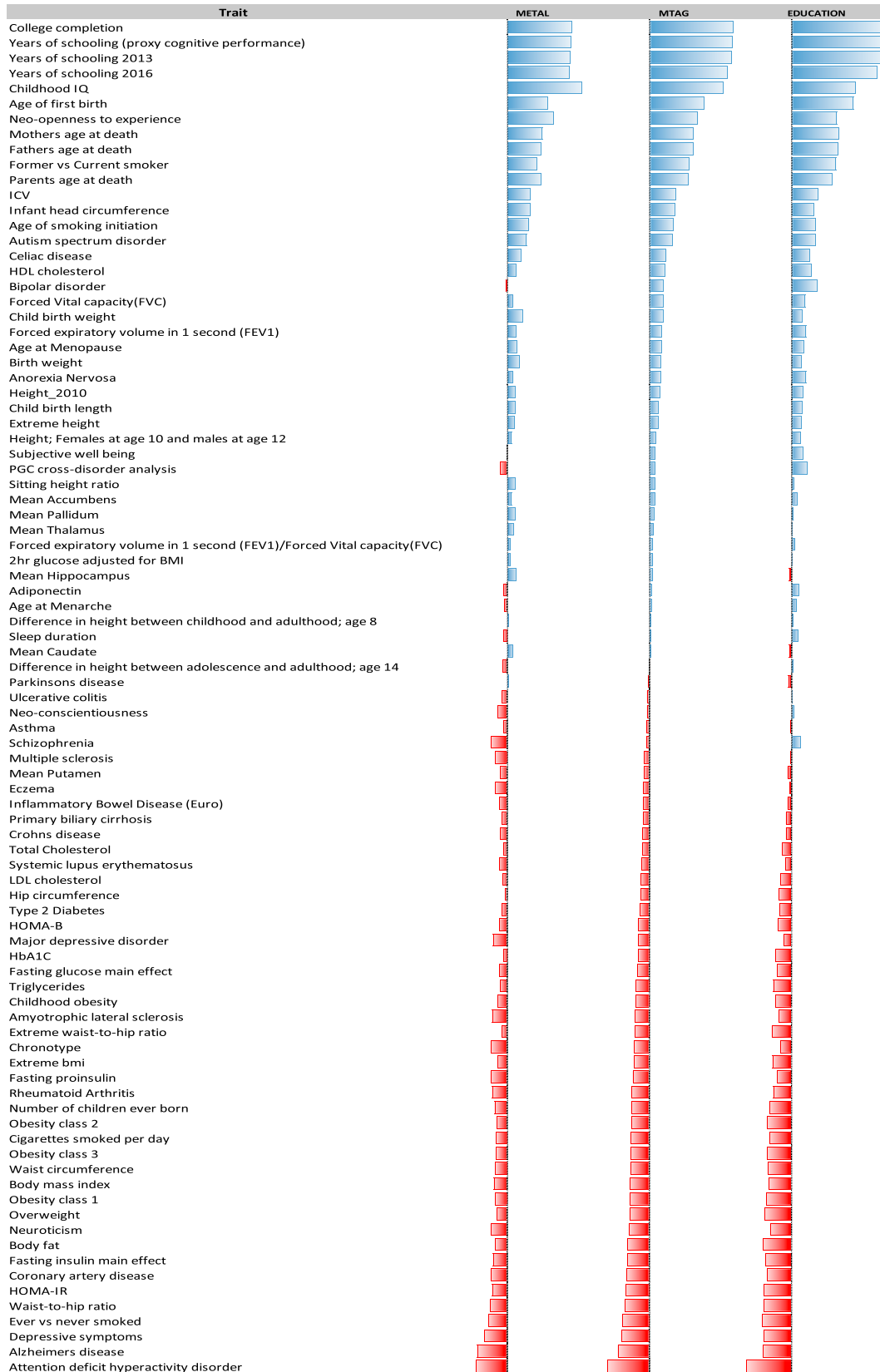
Genetic Correlations with Other Phenotypes

LD score regression was carried out across 89 traits in 15 broad phenotypic categories in LD Hub (Zheng et al., 2017): (1) aging, (2) anthropometric, (3) autoimmune, (4) brain volume, (5) cardiometabolic, (6) education, (7) glycemic, (8) lipids, (9) lung function, (10) neurological, (11) personality, (12) psychiatric, (13) reproductive behavior, (14) sleep, and (15) smoking behavior (Figure 5; Table S14). We performed LD score regression separately for the results of our initial meta-analysis and for the MTAG results. For comparison, we also present LD score regression results for the educational attainment GWAS of Okbay et al. (2016). It should be noted that only 14 phenotypes were examined for genetic correlation in that publication.

Cognition appeared to be strongly associated at the genetic level with aging, education, personality, neuropsychiatric disorders, reproductive behavior, and smoking behavior. Strong association with parental age at death was observed for both the GWAS meta-analysis and MTAG results. Meanwhile, moderate associations with anthropometric traits were observed,

although associations with brain volumes were surprisingly modest, except for total intracranial volume (r_g for MTAG results = 0.31, $p = 7.37E-19$). While many of these correlations have been described previously (Hagenaars et al., 2016; Okbay et al., 2016; Sniekers et al., 2017; Trampush et al., 2017), two results observed in the present study were not reported in those prior publications. First, we report a strong positive genetic correlation between cognitive performance and maternal age at first birth (r_g for MTAG results = 0.63, $p = 2.36E-163$) and inverse correlation with parental number of children ever born (r_g for MTAG results = -0.22 ; $p = 6.91E-13$). It is possible that these effects are mediated by years of higher education, insofar as correlations were even stronger with educational attainment (r_g for parental age at first birth = 0.72, $p = 2.24E-244$; r_g for number of children = -0.26 , $p = 3.34E-18$). As with any other regression relationship, a role for unmeasured mediators, such as propensity for delayed gratification, cannot be ruled out. Second, we observed modest, yet nominally significant, inverse correlations between cognition and autoimmune diseases such as eczema

(B) Circular Manhattan Plot for MetaXcan results based on MTAG of cognitive performance with educational attainment. From inner circle out, GTEX tissue order is as follows: ACC, Anterior Cingulate Cortex; CDBG, Caudate – Basal Ganglia; CRBHM, Cerebellar Hemisphere; CRBLM, Cerebellum; CRTX, Cortex; FCTX, Frontal Cortex; HIPP, Hippocampus; HYPO, Hypothalamus; NACMB, Nucleus Accumbens; PUTM, Putamen. GWAS threshold is set at Bonferroni-corrected $p < 0.05$.



(legend on next page)

and Crohn's disease, attaining Bonferroni significance for rheumatoid arthritis (r_g for MTAG results = -0.2086 ; $p = 1.60E-08$). There was also a Bonferroni-significant positive genetic correlation with celiac disease (r_g for MTAG results = 0.1922 ; $p = 0.0001$). While results of cross-trait analyses were largely consistent using either the GWAS results, the MTAG results, or the previously published educational attainment datasets, there were notable divergences in correlations with psychiatric phenotypes, especially schizophrenia and bipolar disorder.

DISCUSSION

Uncovering the molecular genetic basis of individual differences in cognitive performance can have a significant impact on our understanding of neuropsychiatric disorders, which are both phenotypically (Burdick et al., 2011; Ferreri et al., 2011; Keefe and Harvey, 2012; Snyder, 2013) and genetically (Lencz et al., 2014; Smeland et al., 2017; Stergiakouli et al., 2017) correlated with cognition, as well as numerous non-psychiatric health-relevant phenotypes (Hagenaars et al., 2016), which also demonstrate significant genetic correlations with cognitive function. Here, we have presented the largest GWAS of cognition to date, with 107,207 individuals phenotypically characterized for performance on standardized tests measuring general cognitive ability. Results were further enhanced by utilizing a relatively new approach to allow meta-analysis with a large-scale GWAS of educational attainment, which is highly (though not perfectly) correlated with cognitive ability at the genetic level. With this approach, we were able to identify 70 genomic loci significantly associated with cognition, implicating 350 candidate genes underlying cognitive ability. In total, we found that common SNPs were able to account for roughly half of the overall heritability of the phenotype as determined by prior family studies (Plomin and Deary, 2015).

Downstream analysis confirmed an important role for neurodevelopmental processes in cognitive ability, consistent with implications from the education GWAS (Okbay et al., 2016). Significant genes were more strongly enriched for expression in fetal brain tissue than adult tissue. Results were also enriched for genes implicated in early neurodevelopmental disorders, and neurogenesis was the most strongly enriched GO biological process. At the same time, it is important to emphasize that adult neural tissues were also strongly represented in the results, and multiple synaptic components were significant in the pathway analysis. In this context, it is noteworthy that many cellular processes necessary for early neurodevelopment are also involved in adult synaptic plasticity. This duality is represented by several significant genes emerging from our analysis. *CELSR3* encodes an atypical cadherin plasma membrane protein involved in long-range axon guidance in neurodevelopment through planar cell polarity signaling (Chai et al., 2015) but is also necessary for adult formation of hippocampal glutamatergic synapses (Thakar et al.,

2017). Similarly *SEMA3F* is a negative regulator of dendritic spine development in adult hippocampus (Tran et al., 2009) but embryonically serves as an endogenous chemorepellent, guiding septohippocampal fibers away from non-limbic regions of developing cortex (Pascual et al., 2005).

While synaptic mechanisms were strongly implicated by our results, it is noteworthy that there was no statistical evidence for enrichment of genes expressed in oligodendrocytes or astrocytes. While developmental disorders primarily affecting oligodendrocytes, such as metachromatic leukodystrophy, are marked by cognitive impairment (Faust et al., 2010), it is possible that individual variation in cognitive ability within the normal range is less directly under genetic control via white matter mechanisms. By contrast, strong evidence was provided for the involvement of genes expressed in the cerebellum. Converging evidence from functional imaging studies, lesion studies, structural connectivity, and evolutionary considerations strongly implicate a role for the cerebellum in higher cognitive functions (Buckner, 2013), possibly through the mechanism of prediction and error-based learning (Sokolov et al., 2017).

By utilizing TWAS methodology, we were able to isolate expression effects of specific genes within some of our broad GWAS loci. For example, *ACTR1A*, which lies near the GWAS peak at chromosome 10q24, encodes a microtubular dynactin protein involved in retrograde axon transport (Moughamian et al., 2013). Other genes at this locus were not significant in the TWAS analysis (although a role in cognition cannot be ruled out, given the limited sample size in the reference brain expression datasets in GTEx). However, most of the genes implicated by TWAS were clustered in a few "hot" genomic loci, which may represent topologically associated domains (TADs) under the control of a shared three-dimensional chromatin structure (Gonzalez-Sandoval and Gasser, 2016). Whether effects on cognition are driven by all differentially expressed genes within such loci or if specific effects can be disentangled through experimental means remains to be determined.

The overlap of 23 genes from our results with known genes for Mendelian disorders characterized by intellectual disability has several implications. First, this statistically significant enrichment provides partial validation of our MTAG results. Second, genes with known mutations of large effect, when combined with our data demonstrating SNPs with smaller regulatory effects on the same phenotype (cognition), can be considered an "allelic series" (Plenge et al., 2013)—a natural set of experiments powerfully demonstrating directional information (in the form of a dose-response curve) regarding gene function. Such information can be leveraged for the identification of novel drug targets. Third, converging evidence across the Mendelian and GWAS lists can aid interpretation of specific pathways and molecular processes that are necessary to normal neuronal function and vice versa. For example, two genes on both the Mendelian and GWAS lists (*GMPPB* and *LARGE*) are associated with

Figure 5. Genetic Correlations for GWAS Meta-analysis of Cognitive Performance, MTAG of Cognition and Educational Attainment, and GWAS of Educational Attainment

Genetic correlations (r_g) between cognitive phenotypes and other publicly available GWAS results, based on LD score regression. The first and second columns (labeled METAL and MTAG, respectively) refer to results of the cognitive meta-analyses in the present report. The third column displays correlations for the educational attainment GWAS of Okbay et al. (2016).

dystroglycanopathies with mental retardation. This information provides context for the observation that *DAG1*, which encodes dystroglycan 1, is the strongest TWAS result in the hippocampus. *DAG1* is necessary for GABAergic signaling in hippocampal interneurons (Früh et al., 2016). While dystroglycanopathies are most prominently characterized by muscular dystrophy and retinal abnormalities, it is possible that all of these genes play a role in hippocampal synapse formation that is relevant to normal cognitive ability.

As noted above, one of the most important aims of GWAS studies is the identification of novel drug targets, and it has been suggested that targets with supporting GWAS evidence may be twice as successful in clinical development compared to those without such evidence (Nelson et al., 2015). Our drug set enrichment analysis pointed to several potential nootropic mechanisms. Most notably, the strongest signal was for cinnarizine, a T-type calcium channel inhibitor typically prescribed for seasickness. In the present study, we discovered an association of cognition to *CACNA1I*, which encodes one component of the voltage-dependent T-Type Cav3.3 channel and has been previously associated with schizophrenia (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014). While cinnarizine has strong antihistamine activity and may be inappropriate for general cognitive enhancement, a novel agent targeting Cav3.3 has shown nootropic activity in preclinical models (Moriguchi et al., 2012). In addition to gene set results suggesting a potential role for calcium and potassium channel regulation, single-gene results also point toward a potential role for the metabotropic glutamate receptor encoded by *GRM3*. This gene is also implicated in schizophrenia (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014), and drugs targeting *GRM3* have been suggested as a potential treatment (Lencz and Malhotra, 2015); however, a large-scale trial of one such agent was unsuccessful in treating psychotic symptoms (Downing et al., 2014). Based on the present results, future studies may seek to examine a role for such compounds in cognitive remediation. It is also noteworthy that the present study identified genome-wide significant evidence implicating three phosphodiesterase genes: *PDE1C*, *PDE2A*, and *PDE4D*. In particular, there is growing interest in *PDE2A* inhibitors as potential agents for cognitive enhancement (Trabanco et al., 2016), and evidence suggests that these agents may enhance synaptic plasticity via presynaptic modulation of cAMP hydrolysis (Fernández-Fernández et al., 2015). *PDE4D* inhibition is also under investigation as a potential therapy for neurodegenerative disease (Ricciarelli et al., 2017).

It is important to emphasize that uncovering genetic variation underlying general cognitive ability in the healthy population does not have deterministic implications. As has been previously explicated in similar studies (Trampush et al., 2017), effect sizes for each allele are extremely small ($R^2 < 0.1\%$ for even the strongest effects), and the combined effects genome-wide predict only a small proportion of the total variance in hold-out samples (Figure 3). Thus, results of the present study do not hold the potential for individual prediction or classification. Nevertheless, the results may still have substantial impact on our understanding of molecular mechanisms underlying cognitive ability.

EXPERIMENTAL PROCEDURES

Subject Details

The cohorts included in the current study were described in detail in two prior reports on cognitive performance (Sniekers et al., 2017; Trampush et al., 2017) and one prior report on educational attainment (Okbay et al., 2016). Sample sizes for these three studies were $n = 78,308$, $n = 35,298$, and $n = 328,917$, respectively. For the present study, two cohorts reported in Trampush et al. (2017) were excluded, so that cohorts included will be independent from those reported in Sniekers et al. (2017): (1) Minnesota Center for Twin and Family Research (MCTFR) and (2) Lothian Birth Cohort 1936 Study. As a result, sample sizes decreased from the originally reported $n = 35,298$ to $n = 28,899$. All phenotypes included were as reported originally in the respective publications. All subjects provided written, informed consent to procedures that were approved by local review boards for the institutions at which each cohort was collected. Further details are available in the supplementary materials to those three publications.

GWAS Quality Control

Markers reported in the prior COGENT study (Trampush et al., 2017) were updated to build 37 coordinates but were originally imputed against the HRC (Haplotype Reference Consortium) reference panel (McCarthy et al., 2016) via the Sanger imputation server. To ensure that markers, allele frequencies, and alleles were aligned to the 1000 Genomes phase 3 reference panel (1000 Genomes Project Consortium et al., 2015), the COGENT summary statistics (Trampush et al., 2017) were checked using the EasyQC pipeline (Winkler et al., 2014), which allows summary statistics to be aligned and checked against a reference panel of choice. We used the default 1000 Genomes phase 3 reference panel (1000 Genomes Project Consortium et al., 2015), provided along with the EasyQC package. Markers were inspected for allele frequency outliers, presence of duplicated markers, and allele mismatches with the 1000 Genomes reference panel. Quality control filters for INFO score < 0.6 and $n < 10,000$ were additionally implemented. After EasyQC quality control, 8,040,131 SNPs were available for analysis. Only 87 SNPs were excluded due to allele mismatches, 13,276 SNPs were excluded due to allele frequency mismatches from the 1000 Genomes phase 3 reference panel, 283,163 were found to be duplicates and excluded, 104 SNPs were found on the HRC reference panel, but not on the 1000 Genomes phase 3 reference panel, and 2,723,493 SNPs had sample sizes $< 10,000$ individuals. None of the SNPs failed the INFO score < 0.6 cutoff. The same set of SNPs was utilized for subsequent reduced sample meta-analysis without the overlapping LBC1936 and MCTFR cohorts in Trampush et al. (2017). As the other prior studies of cognitive performance (Sniekers et al., 2017) and education (Okbay et al., 2016) were imputed to the 1000 Genomes phase 3 reference panel, summary statistics were used as provided (https://ctg.cncr.nl/software/summary_statistics; <https://www.thessgac.org/data>).

GWAS Meta-Analysis

Fixed-effect meta-analysis was conducted between Sniekers et al. (2017) and independent cohorts reported in Trampush et al. (2017) using the METAL package (Willer et al., 2010). To ensure that results of the meta-analysis were contributed by both studies, markers present only in Sniekers et al. (2017) or Trampush et al. (2017), but not in both, were excluded for further analysis. The number of available markers after QC filtering was 7,357,080. Because the GWAS of Sniekers et al. (2017) utilized the sample-size-weighted method to perform meta-analysis across its own cohorts and did not report variance terms, our meta-analysis was conducted using the sample-size-weighted method.

Multi-Trait Analysis for GWAS (MTAG)

To further enrich genetic signals, we employed a newly developed methodology that integrates LD score regression and meta-analysis techniques across related traits: MTAG (Turley et al., 2017). MTAG (v0.9.0) was applied to the METAL results described immediately above and combined with summary statistics from the recent, large-scale education GWAS (Okbay et al., 2016). MTAG analysis allows the boosting of genetic signals across related traits and has been found to be effective in resolving unknown sample overlaps,

generating trait-specific effect estimates weighted by bivariate genetic correlation. The MTAG QC pipeline aligned all alleles across both sets of summary statistics and ensured that SNPs were present across all datasets. SNPs that were not present in either dataset were removed. The final SNP count for MTAG was 7,333,576. The MTAG methodology proceeds by: (1) estimating the variance-covariance matrix of the GWAS estimation error, by using a series of LD score regressions, of which, under the known properties of LD score regression, captures relevant sources of estimation error, incorporating population stratification, unknown sample overlap, and cryptic relatedness, (2) estimating the variance-covariance of SNP effects using the maximum likelihood procedure reported in [Turley et al. \(2017\)](#), and (3) computing the MTAG estimator for each SNP and each trait. Summary statistics consisting of SNP, CHR, BP, per SNP sample size, BETA, and SE for each trait were entered to the MTAG python command line. The resulting effect estimates and p values are interpreted the same as single-trait GWAS, which allows standard downstream follow-up analysis on the summary statistics. The python code for MTAG is available at <https://github.com/omeed-maghzian/mtag>.

Functional Mapping and Annotation for GWAS

GWAS summary statistics from the METAL meta-analysis and MTAG analysis were separately entered into the FUMA pipeline ([Watanabe et al., 2017](#)). The FUMA pipeline enables fast prioritization of genomic variants and genes and permits interactive visualization of genomic results with respect to state-of-the-art bioinformatics resources. Manhattan and QQ plots are produced, and MAGMA gene-based analysis is performed, accounting for gene size and LD structure. FUMA was also utilized to perform competitive gene-set analyses for GO cell compartment and biological process categories using the Molecular Signature Database (MsigDB 5.2). A separate competitive gene-set analysis was also conducted for the drug-based pathways previously described by [Gaspar and Breen \(2017\)](#). The pipeline also generates aggregated statistics for independent loci, lead SNPs, tagged genes, and supplementary plots—including SNP and locus annotations. Default clumping parameters are: GWAS p value < 5E-08; r^2 threshold to define LD structure of independent SNPs > 0.1; maximum p value cutoff < 0.05; population for clumping = EUR; minor allele frequency filter > 0.01; maximum distance between LD blocks to merge into a single locus = 250 kb. Follow-up queries were then made for independent loci of the cognitive performance meta-analysis as well as the MTAG results and compared against summary statistics for the prior cognitive and education GWAS. For purposes of comparison, loci in which the lead SNPs were within 500kb of each other were considered overlapping.

We compared the list of genes resulting from the MTAG analysis (including all genes within GWS SNP loci, as well as GWS genes identified with MAGMA) with a list of 621 genes known to cause autosomal dominant or autosomal recessive Mendelian disorders featuring intellectual disability. This list is primarily derived from a recent comprehensive review ([Vissers et al., 2016](#)), supplemented by a subsequent large-scale study of consanguineous multiplex families ([Harripaul et al., 2017](#)). A total of 193 autosomal dominant genes were identified, and a total of 413 autosomal recessive genes were identified. Fifteen genes were annotated as causing both autosomal dominant and autosomal recessive disorders with intellectual disability. Statistical significance was determined by probabilities derived according to the hypergeometric distribution. For this purpose, the total pool of autosomal genes was set to 19,011 (per Gencode).

Polygenic Risk Prediction for Independent Datasets

To validate that the genetic architecture elucidated via the MTAG methodology, we attempted to predict the phenotypic variance of general cognitive function in two of the independent COGENT cohorts (ASPI and GCAP). MTAG analysis was conducted as above, but removing the COGENT cohorts. Polygenic score prediction across multiple thresholds of P_T was conducted using PRSice ([Euesden et al., 2015](#)). To compare the effectiveness of MTAG, we also conducted polygenic risk prediction using IQ-only and education-only summary statistics. Finally, R^2 across SNP thresholds is compared to obtain the degree of improvement in terms of the ratio of MTAG PRS R^2 values versus those of IQ or education PRS R^2 .

Stratified LD Regression: Cell Type Expression and Epigenomics

Functional characterization of GWAS summary statistics was carried out via stratified LD regression to investigate if heritability of cognitive performance is enriched in specific tissue or cell types. Summary statistics were first subjected to baseline partitioned heritability and thereafter passed through a cell-type-specific functional characterization pipeline ([Finucane et al., 2017](#)). Cell-type characterization includes the DEPICT tissue expression database, GTEx tissue expression, IMMGEN immune cell types, CAHOY brain level cell types, and the ROADMAP cell epigenomic marks.

Transcriptome-Wide Analysis and Brain Expression lookups

Transcriptome-wide analysis was carried out via MetaXcan ([Barbeira et al., 2016](#)), which allows for GTEx brain expression data to be integrated with GWAS summary statistics. MetaXcan computes downstream phenotypic associations of genetic regulation of molecular traits, using elastic, adjustment for model uncertainty, and colocalization of GWAS and eQTL signals ([Barbeira et al., 2016](#)). GTEx Version 6, brain tissue expression profiles and sample sizes include the anterior cingulate cortex (n = 72); caudate-basal ganglia (n = 100); cerebellar hemisphere (n = 89); cerebellum (n = 103); cortex (n = 96); frontal cortex (n = 92); hippocampus (n = 81); hypothalamus (n = 81); nucleus accumbens (n = 93); and putamen (n = 82).

Bayesian Fine-Mapping Analysis and Functional Annotations

To identify potential causal variants in each of the independent loci, CAVIAR-BF is implemented to a region ± 50 KB of a lead SNP identified in the MTAG analysis. We followed similar procedures setting prior effect distribution σ_a to 0.1 in the model, which was recommended for GWAS studies ([Chen et al., 2015](#); <https://bitbucket.org/Wenan/caviarbf>). The prior probability of being causal for each SNP is set to $1/m$, where m is the number of SNPs. Bayes factor was calculated for three model sets for independent loci, which modeled for 1, 2, and up to 3 causal SNPs within each independent region, after which a model search algorithm searches and identifies the putative causal SNPs. These SNPs were then annotated using the Ensembl Variant Effect Predictor ([McLaren et al., 2016](#)). The analysis was repeated for extended regions taking into account the length of the independent loci identified by earlier FUMA procedures modeling for either 1 or 2 causal SNPs. SNPs identified by the two stage CAVIARBF analysis were then examined for potential gene expression in the BrainEAC ([Ramasamy et al., 2014](#)) and CommonMind ([Haugberg et al., 2017](#)) databases. BrainEAC top SNP lookups were for the following tissue expression across n = 134 individuals: aveALL, all area combined; CRBL, cerebellum; FCTX, frontal cortex; HIPP, hippocampus; MEDU, medulla; OCTX, occipital cortex; PUTM, putamen; SNIG, substantia nigra; TCTX, temporal cortex; THAL, thalamus; and WHMT, white matter. Finally, the prefrontal cortex lookup was included as part of the CommonMind consortium brain expression profile in n = 467 genetically inferred Caucasian samples.

Linkage Disequilibrium Score Regression

LD score regression allows genetic correlations to be computed across traits ([Bulik-Sullivan et al., 2015a, 2015b](#)), which allows further insights to be drawn from understanding the degree to which genetic architecture are shared across traits. To further examine potential traits that overlap with the cognitive architecture from the cognition meta-analysis results and MTAG results, LD score regression was conducted via the LD-hub pipeline, a centralized trait database ([Zheng et al., 2017](#)). LD score regression was carried out across 89 traits in 15 broad phenotypic categories: (1) aging, (2) anthropometric, (3) autoimmune, (4) brain volume, (5) cardiometabolic, (6) education, (7) glyceric, (8) lipids, (9) lung function, (10) neurological, (11) personality, (12) psychiatric, (13) reproductive behavior, (14) sleep, and (15) smoking behavior. Very recent reported GWAS summary statistics for attention deficit hyperactivity disorder (ADHD; [Demontis et al., 2017](#)) and intracranial volume (ICV; [Adams et al., 2016](#)) were included as additional phenotypes. For comparison, we also present LD score regression results for the educational attainment GWAS of [Okbay et al. \(2016\)](#). It should be noted that only 14 phenotypes were examined for genetic correlation in that publication. It should be noted that the MHC (Major Histocompatibility Complex) region was redacted from all datasets prior to LD score regression analysis, as per standard protocol at LD-Hub.

SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures and fourteen tables and can be found with this article online at <https://doi.org/10.1016/j.celrep.2017.11.028>.

AUTHOR CONTRIBUTIONS

T.L. designed the study and supervised the data analysis. M.L. performed the primary data analysis, and J.W.T., J.Y., and E.K. provided additional statistical input. A.K.M., D.C.G., I.J.D., K.E.B., and G.D. provided the initial conceptual framework for the COGENT consortium. M.L. and T.L. drafted the manuscript. All other authors were involved in ascertainment, assessment, and analysis of individual cohorts, provided conceptual input to study design, and critically reviewed the manuscript.

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Cardiovascular Health Study (CHS): phs000287.v4.p1, phs000377.v5.p1, and phs000226.v3.p1

Framingham Heart Study (FHS): phs000007.v23.p8 and phs000342.v11.p8

Multi-Site Collaborative Study for Genotype-Phenotype Associations in Alzheimer's Disease (GENADA): phs000219.v1.p1

Long Life Family Study (LLFS): phs000397.v1.p1

Genetics of Late Onset Alzheimer's Disease Study (LOAD): phs000168.v1.p1

Minnesota Center for Twin and Family Research (MCTFR): phs000620.v1.p1

Philadelphia Neurodevelopmental Cohort (PNC): phs000607.v1.p1

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WEB RESOURCES

EasyQC, <http://www.uni-regensburg.de/medizin/epidemiologie-praeventivmedizin/genetische-epidemiologie/software/>
 METAL, http://genome.sph.umich.edu/wiki/METAL_Documentation
 MTag, <https://github.com/omeed-maghzian/mtag>
 LD-HUB, <http://ldsc.broadinstitute.org/>
 LDSC, <https://github.com/bulik/ldsc>
 METAXCAN, <https://github.com/hakyimlab/MetaXcan>
 FUMA, <http://fuma.ctglab.nl/>
 PRSice, <http://prsice.info/>
 BRAINEAC, <http://www.braineac.org/>
 CommonMind, <https://www.synapse.org/#!/Synapse:syn2759792/wiki/69613>
 MAGMA, <https://ctg.cncr.nl/software/magma>

REFERENCES

- Adams, H.H.H., Hibar, D.P., Chouraki, V., Stein, J.L., Nyquist, P.A., Rentería, M.E., Trompet, S., Arias-Vasquez, A., Seshadri, S., Desrivières, S., et al. (2016). Novel genetic loci underlying human intracranial volume identified through genome-wide association. *Nat. Neurosci.* **19**, 1569–1582.
- Barbeira, A., Shah, K.P., Torres, J.M., Wheeler, H.E., Torstenson, E.S., Edwards, T., Garcia, T., Bell, G.I., Nicolae, D., Cox, N.J., et al. (2016). MetaXcan: Summary Statistics Based Gene-Level Association Method Infers Accurate PrediXcan Results. *bioRxiv*. <https://doi.org/10.1101/045260>.
- Belsky, D.W., Moffitt, T.E., Corcoran, D.L., Domingue, B., Harrington, H., Hoggan, S., Houts, R., Ramrakha, S., Sugden, K., Williams, B.S., et al. (2016). The Genetics of Success: How Single-Nucleotide Polymorphisms Associated With Educational Attainment Relate to Life-Course Development. *Psychol. Sci.* **27**, 957–972.
- Benyamin, B., Pourcain, B.S., Davis, O.S., Davies, G., Hansell, N.K., Brion, M.-J., Kirkpatrick, R.M., Cents, R. a. M., Franić, S., Miller, M.B., et al. (2013). Childhood intelligence is heritable, highly polygenic and associated with FBNP1L. *Mol. Psychiatry* **19**, 253–258.
- Buckner, R.L. (2013). The cerebellum and cognitive function: 25 years of insight from anatomy and neuroimaging. *Neuron* **80**, 807–815.
- Bulik-Sullivan, B., Finucane, H.K., Anttila, V., Gusev, A., Day, F.R., Loh, P.R., Duncan, L., Perry, J.R., Patterson, N., Robinson, E.B., et al.; ReproGen Consortium; Psychiatric Genomics Consortium; Genetic Consortium for Anorexia Nervosa of the Wellcome Trust Case Control Consortium 3 (2015a). An atlas of genetic correlations across human diseases and traits. *Nat. Genet.* **47**, 1236–1241.
- Bulik-Sullivan, B.K., Loh, P.-R., Finucane, H.K., Ripke, S., Yang, J., Patterson, N., Daly, M.J., Price, A.L., and Neale, B.M.; Schizophrenia Working Group of the Psychiatric Genomics Consortium (2015b). LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat. Genet.* **47**, 291–295.
- Burdick, K.E., Goldberg, T.E., Cornblatt, B.A., Keefe, R.S., Gopin, C.B., Derosse, P., Braga, R.J., and Malhotra, A.K. (2011). The MATRICS consensus cognitive battery in patients with bipolar I disorder. *Neuropsychopharmacology* **36**, 1587–1592.
- Cahoy, J.D., Emery, B., Kaushal, A., Foo, L.C., Zamanian, J.L., Christopherson, K.S., Xing, Y., Lubischer, J.L., Krieg, P.A., Krupenko, S.A., et al. (2008). A transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for understanding brain development and function. *J. Neurosci.* **28**, 264–278.
- Chai, G., Goffinet, A.M., and Tissir, F. (2015). Celsr3 and Fzd3 in axon guidance. *Int. J. Biochem. Cell Biol.* **64**, 11–14.
- Chen, W., Larrabee, B.R., Ovsyannikova, I.G., Kennedy, R.B., Haralambieva, I.H., Poland, G.A., and Schaid, D.J. (2015). Fine Mapping Causal Variants with an Approximate Bayesian Method Using Marginal Test Statistics. *Genetics* **200**, 719–736.
- Cooper, G.M., Coe, B.P., Girirajan, S., Rosenfeld, J.A., Vu, T.H., Baker, C., Williams, C., Stalker, H., Hamid, R., Hannig, V., et al. (2011). A copy number variation morbidity map of developmental delay. *Nat. Genet.* **43**, 838–846.
- Davies, G., Tenesa, A., Payton, A., Yang, J., Harris, S.E., Liewald, D., Ke, X., Le Hellard, S., Christoforou, A., Luciano, M., et al. (2011a). Genome-wide association studies establish that human intelligence is highly heritable and polygenic. *Mol. Psychiatry* **16**, 996–1005.
- Davies, G., Tenesa, A., Payton, A., Yang, J., Harris, S.E., Liewald, D., Ke, X., Le Hellard, S., Christoforou, A., Luciano, M., et al. (2011b). Genome-wide association studies establish that human intelligence is highly heritable and polygenic. *Mol. Psychiatry* **16**, 996–1005.
- Davies, G., Armstrong, N., Bis, J.C., Bressler, J., Chouraki, V., Giddaluru, S., Hofer, E., Ibrahim-Verbaas, C.A., Kirin, M., Lahti, J., et al.; Generation Scotland (2015). Genetic contributions to variation in general cognitive function: a meta-analysis of genome-wide association studies in the CHARGE consortium (N=53949). *Mol. Psychiatry* **20**, 183–192.
- Davies, G., Marioni, R.E., Liewald, D.C., Hill, W.D., Hagenaars, S.P., Harris, S.E., Ritchie, S.J., Luciano, M., Fawns-Ritchie, C., and Lyall, D. (2016). Genome-wide association study of cognitive functions and educational attainment in UK Biobank (N=112 151). *Mol. Psychiatry* **21**, 758–767.
- de Leeuw, C.A., Mooij, J.M., Heskes, T., and Posthuma, D. (2015). MAGMA: generalized gene-set analysis of GWAS data. *PLoS Comput. Biol.* **11**, e1004219.
- Deary, I.J., Johnson, W., and Houlihan, L.M. (2009). Genetic foundations of human intelligence. *Hum. Genet.* **126**, 215–232.
- Demontis, D., Walters, R.K., Martin, J., Mattheisen, M., Als, T.D., Agerbo, E., Belliveau, R., Bybjerg-Grauholm, J., Bækved-Hansen, M., Cerrato, F., et al. (2017). Discovery Of The First Genome-Wide Significant Risk Loci For ADHD. *bioRxiv*. <https://doi.org/10.1101/145581>.
- Downing, A.M., Kinon, B.J., Millen, B.A., Zhang, L., Liu, L., Morozova, M.A., Brenner, R., Rayle, T.J., Nisenbaum, L., Zhao, F., and Gomez, J.C. (2014). A Double-Blind, Placebo-Controlled Comparator Study of LY2140023 monohydrate in patients with schizophrenia. *BMC Psychiatry* **14**, 351.
- Euesden, J., Lewis, C.M., and O'Reilly, P.F. (2015). PRSice: Polygenic Risk Score software. *Bioinformatics* **31**, 1466–1468.
- Faust, P.L., Kaye, E.M., and Powers, J.M. (2010). Myelin lesions associated with lysosomal and peroxisomal disorders. *Expert Rev. Neurother.* **10**, 1449–1466.
- Fernández-Fernández, D., Rosenbrock, H., and Kroker, K.S. (2015). Inhibition of PDE2A, but not PDE9A, modulates presynaptic short-term plasticity measured by paired-pulse facilitation in the CA1 region of the hippocampus. *Synapse* **69**, 484–496.
- Ferreri, F., Lapp, L.K., and Peretti, C.-S. (2011). Current research on cognitive aspects of anxiety disorders. *Curr. Opin. Psychiatry* **24**, 49–54.
- Filippini, N., Rao, A., Wetten, S., Gibson, R.A., Borrie, M., Guzman, D., Kertesz, A., Loy-English, I., Williams, J., Nichols, T., et al. (2009). Anatomically-distinct genetic associations of APOE epsilon4 allele load with regional cortical atrophy in Alzheimer's disease. *Neuroimage* **44**, 724–728.
- Finucane, H., Reshef, Y., Anttila, V., Slowikowski, K., Gusev, A., Byrnes, A., Gazal, S., Loh, P.-R., Genovese, G., Saunders, A., et al. (2017). Heritability

- enrichment of specifically expressed genes identifies disease-relevant tissues and cell types. *bioRxiv*. <https://doi.org/10.1101/103069>.
- Früh, S., Romanos, J., Panzanelli, P., Bürgisser, D., Tyagarajan, S.K., Campbell, K.P., Santello, M., and Fritschy, J.-M. (2016). Neuronal Dystroglycan Is Necessary for Formation and Maintenance of Functional CCK-Positive Basket Cell Terminals on Pyramidal Cells. *J. Neurosci.* **36**, 10296–10313.
- Gaspar, H.A., and Breen, G. (2017). Drug enrichment and discovery from schizophrenia genome-wide association results: an analysis and visualisation approach. *Sci. Rep.* **7**, 12460.
- Gonzalez-Sandoval, A., and Gasser, S.M. (2016). On TADs and LADs: Spatial Control Over Gene Expression. *Trends Genet.* **32**, 485–495.
- Gudbjartsson, D.F., Walters, G.B., Thorleifsson, G., Stefansson, H., Halldorsson, B.V., Zusmanovich, P., Sulem, P., Thorlacius, S., Gylfason, A., Steinberg, S., et al. (2008). Many sequence variants affecting diversity of adult human height. *Nat. Genet.* **40**, 609–615.
- Guha, S., Rees, E., Darvasi, A., Ivanov, D., Ikeda, M., Bergen, S.E., Magnusson, P.K., Cormican, P., Morris, D., Gill, M., et al.; Molecular Genetics of Schizophrenia Consortium; Wellcome Trust Case Control Consortium 2 (2013). Implication of a rare deletion at distal 16p11.2 in schizophrenia. *JAMA Psychiatry* **70**, 253–260.
- Hagenaars, S.P., Harris, S.E., Davies, G., Hill, W.D., Liewald, D.C.M., Ritchie, S.J., Marioni, R.E., Fawns-Ritchie, C., Cullen, B., Malik, R., et al.; METASTROKE Consortium, International Consortium for Blood Pressure GWAS; SpiroMeta Consortium; CHARGE Consortium Pulmonary Group, CHARGE Consortium Aging and Longevity Group (2016). Shared genetic aetiology between cognitive functions and physical and mental health in UK Biobank (N=112 151) and 24 GWAS consortia. *Mol. Psychiatry* **27**, 1624–1632.
- Harripaul, R., Vasli, N., Mikhailov, A., Rafiq, M.A., Mittal, K., Windpassinger, C., Sheikh, T.I., Noor, A., Mahmood, H., Downey, S., et al. (2017). Mapping autosomal recessive intellectual disability: combined microarray and exome sequencing identifies 26 novel candidate genes in 192 consanguineous families. *Mol. Psychiatry*, Published online April 11, 2017. <https://doi.org/10.1038/mp.2017.60>.
- Hauberg, M.E., Zhang, W., Giambartolomei, C., Franzén, O., Morris, D.L., Vyse, T.J., Ruusalepp, A., Sklar, P., Schadt, E.E., Björkegren, J.L.M., and Roussos, P.; CommonMind Consortium (2017). Large-Scale Identification of Common Trait and Disease Variants Affecting Gene Expression. *Am. J. Hum. Genet.* **100**, 885–894.
- Johnson, W., te Nijenhuis, J., and Bouchard, T.J., Jr. (2008). Still just 1 g: Consistent results from five test batteries. *Intelligence* **36**, 81–95.
- Johnson, W., Deary, I.J., Silventoinen, K., Tynelius, P., and Rasmussen, F. (2010). Family background buys an education in Minnesota but not in Sweden. *Psychol. Sci.* **21**, 1266–1273.
- Keefe, R.S., and Harvey, P.D. (2012). Cognitive impairment in schizophrenia. In *Handbook of Experimental Pharmacology*, M.A. Geyer and G. Gross, eds. (Berlin, Heidelberg: Springer), pp. 11–37.
- Lencz, T., and Malhotra, A.K. (2015). Targeting the schizophrenia genome: a fast track strategy from GWAS to clinic. *Mol. Psychiatry* **20**, 820–826.
- Lencz, T., Knowles, E., Davies, G., Guha, S., Liewald, D.C., Starr, J.M., Djurovic, S., Melle, I., Sundet, K., Christoforou, A., et al. (2014). Molecular genetic evidence for overlap between general cognitive ability and risk for schizophrenia: a report from the Cognitive Genomics consortium (COGENT). *Mol. Psychiatry* **19**, 168–174.
- Li, H., Wetten, S., Li, L., St. Jean, P.L., Upmanyu, R., Surh, L., Hosford, D., Barnes, M.R., Briley, J.D., Borrie, M., et al. (2008). Candidate single-nucleotide polymorphisms from a genomewide association study of Alzheimer disease. *Arch. Neurol.* **65**, 45–53.
- Locke, A.E., Kahali, B., Berndt, S.I., Justice, A.E., Pers, T.H., Day, F.R., Powell, C., Vedantam, S., Buchkovich, M.L., Yang, J., et al.; LifeLines Cohort Study; ADIPOGen Consortium; AGEN-BMI Working Group; CARDIOGRAMplusC4D Consortium; CKDGen Consortium; GLGC; ICBP; MAGIC Investigators; MUTHER Consortium; MIGen Consortium; PAGE Consortium; ReproGen Consortium; GENIE Consortium; International Endogene Consortium (2015). Genetic studies of body mass index yield new insights for obesity biology. *Nature* **518**, 197–206.
- McCarthy, S., Das, S., Kretschmar, W., Delaneau, O., Wood, A.R., Teumer, A., Kang, H.M., Fuchsberger, C., Danecek, P., Sharp, K., et al.; Haplotype Reference Consortium (2016). A reference panel of 64,976 haplotypes for genotype imputation. *Nat. Genet.* **48**, 1279–1283.
- McLaren, W., Gil, L., Hunt, S.E., Riat, H.S., Ritchie, G.R.S., Thormann, A., Flicek, P., and Cunningham, F. (2016). The Ensembl Variant Effect Predictor. *Genome Biol.* **17**, 122.
- Moriguchi, S., Shioda, N., Yamamoto, Y., Tagashira, H., and Fukunaga, K. (2012). The T-type voltage-gated calcium channel as a molecular target of the novel cognitive enhancer ST101: enhancement of long-term potentiation and CaMKII autophosphorylation in rat cortical slices. *J. Neurochem.* **121**, 44–53.
- Moughamian, A.J., Osborn, G.E., Lazarus, J.E., Maday, S., and Holzbaur, E.L.F. (2013). Ordered recruitment of dynactin to the microtubule plus-end is required for efficient initiation of retrograde axonal transport. *J. Neurosci.* **33**, 13190–13203.
- Nelson, M.R., Tipney, H., Painter, J.L., Shen, J., Nicoletti, P., Shen, Y., Floratos, A., Sham, P.C., Li, M.J., Wang, J., et al. (2015). The support of human genetic evidence for approved drug indications. *Nat. Genet.* **47**, 856–860.
- Okbay, A., Beauchamp, J.P., Fontana, M.A., Lee, J.J., Pers, T.H., Rietveld, C.A., Turley, P., Chen, G.-B., Emilsson, V., Meddens, S.F.W., et al.; LifeLines Cohort Study (2016). Genome-wide association study identifies 74 loci associated with educational attainment. *Nature* **533**, 539–542.
- Pascual, M., Pozas, E., and Soriano, E. (2005). Role of class 3 semaphorins in the development and maturation of the septohippocampal pathway. *Hippocampus* **15**, 184–202.
- Plenge, R.M., Scolnick, E.M., and Altshuler, D. (2013). Validating therapeutic targets through human genetics. *Nat. Rev. Drug Discov.* **12**, 581–594.
- Plomin, R., and Deary, I.J. (2015). Genetics and intelligence differences: five special findings. *Mol. Psychiatry* **20**, 98–108.
- Ramasamy, A., Trabzuni, D., Guelfi, S., Varghese, V., Smith, C., Walker, R., De, T., Coin, L., de Silva, R., Cookson, M.R., et al.; UK Brain Expression Consortium; North American Brain Expression Consortium (2014). Genetic variability in the regulation of gene expression in ten regions of the human brain. *Nat. Neurosci.* **17**, 1418–1428.
- Ricciarelli, R., Brullo, C., Prickaerts, J., Arancio, O., Villa, C., Rebosio, C., Calcagno, E., Balbi, M., van Hagen, B.T., Argyrousi, E.K., et al. (2017). Memory-enhancing effects of GEBR-32a, a new PDE4D inhibitor holding promise for the treatment of Alzheimer's disease. *Sci. Rep.* **7**, 46320.
- Rietveld, C.A., Medland, S.E., Derringer, J., Yang, J., Esko, T., Martin, N.W., Westra, H.-J., Shakhbazov, K., Abdellaoui, A., Agrawal, A., et al.; LifeLines Cohort Study (2013). GWAS of 126,559 individuals identifies genetic variants associated with educational attainment. *Science* **340**, 1467–1471.
- Rietveld, C.A., Esko, T., Davies, G., Pers, T.H., Turley, P., Benyamin, B., Chabris, C.F., Emilsson, V., Johnson, A.D., Lee, J.J., et al. (2014). Common genetic variants associated with cognitive performance identified using the proxy-phenotype method. *Proc. Natl. Acad. Sci. USA* **111**, 13790–13794.
- Schizophrenia Working Group of the Psychiatric Genomics Consortium (2014). Biological insights from 108 schizophrenia-associated genetic loci. *Nature* **511**, 421–427.
- Smeland, O.B., Frei, O., Kauppi, K., Hill, W.D., Li, W., Wang, Y., Krull, F., Bettella, F., Eriksen, J.A., Witoelar, A., et al. (2017). Identification of Genetic Loci Jointly Influencing Schizophrenia Risk and the Cognitive Traits of Verbal-Numerical Reasoning, Reaction Time, and General Cognitive Function. *JAMA Psychiatry* **74**, 1065–1075.
- Sniekers, S., Stringer, S., Watanabe, K., Jansen, P.R., Coleman, J.R.I., Krapohl, E., Taskesen, E., Hammerslag, A.R., Okbay, A., Zabaneh, D., et al. (2017). Genome-wide association meta-analysis of 78,308 individuals identifies new loci and genes influencing human intelligence. *Nat. Genet.* **49**, 1107–1112.

- Snyder, H.R. (2013). Major depressive disorder is associated with broad impairments on neuropsychological measures of executive function: a meta-analysis and review. *Psychol. Bull.* 139, 81–132.
- Sokolov, A.A., Miall, R.C., and Ivry, R.B. (2017). The Cerebellum: Adaptive Prediction for Movement and Cognition. *Trends Cogn. Sci.* 21, 313–332.
- Stergiakouli, E., Martin, J., Hamshere, M.L., Heron, J., St Pourcain, B., Timpson, N.J., Thapar, A., and Davey Smith, G. (2017). Association between polygenic risk scores for attention-deficit hyperactivity disorder and educational and cognitive outcomes in the general population. *Int. J. Epidemiol.* 46, 421–428.
- Thakar, S., Wang, L., Yu, T., Ye, M., Onishi, K., Scott, J., Qi, J., Fernandes, C., Han, X., Yates, J.R., 3rd., et al. (2017). Evidence for opposing roles of *Celsr3* and *Vangl2* in glutamatergic synapse formation. *Proc. Natl. Acad. Sci. USA* 114, E610–E618.
- 1000 Genomes Project Consortium; Auton, A., Brooks, L.D., Durbin, R.M., Garrison, E.P., Kang, H.M., Korbel, J.O., Marchini, J.L., McCarthy, S., McVean, G.A., and Abecasis, G.R. (2015). A global reference for human genetic variation. *Nature* 526, 68–74.
- Trabanco, A.A., Buijsters, P., and Rombouts, F.J. (2016). Towards selective phosphodiesterase 2A (PDE2A) inhibitors: a patent review (2010 - present). *Expert Opin. Ther. Pat.* 26, 933–946.
- Trampush, J.W., Yang, M.L.Z., Yu, J., Knowles, E., Davies, G., Liewald, D.C., Starr, J.M., Djurovic, S., Melle, I., Sundet, K., et al. (2017). GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: a report from the COGENT consortium. *Mol. Psychiatry* 22, 336–345.
- Tran, T.S., Rubio, M.E., Clem, R.L., Johnson, D., Case, L., Tessier-Lavigne, M., Hagan, R.L., Ginty, D.D., and Kolodkin, A.L. (2009). Secreted semaphorins control spine distribution and morphogenesis in the postnatal CNS. *Nature* 462, 1065–1069.
- Turley, P., Walters, R.K., Maghzi, O., Okbay, A., Lee, J.J., Fontana, M.A., Nguyen-Viet, T.A., Furlotte, N.A., 23andMe Research Team; and Magnusson, P., et al.; Social Science Genetic Association Consortium (2017). MTAG: Multi-Trait Analysis of GWAS. *bioRxiv*. <https://doi.org/10.1101/118810>.
- Vissers, L.E.L.M., Gilissen, C., and Veltman, J.A. (2016). Genetic studies in intellectual disability and related disorders. *Nat. Rev. Genet.* 17, 9–18.
- Watanabe, K., Taskesen, E., van Bochoven, A., and Posthuma, D. (2017). FUMA: Functional mapping and annotation of genetic associations. *bioRxiv*. <https://doi.org/10.1101/110023>.
- Weedon, M.N., Lango, H., Lindgren, C.M., Wallace, C., Evans, D.M., Mangino, M., Freathy, R.M., Perry, J.R.B., Stevens, S., Hall, A.S., et al.; Diabetes Genetics Initiative; Wellcome Trust Case Control Consortium; Cambridge GEM Consortium (2008). Genome-wide association analysis identifies 20 loci that influence adult height. *Nat. Genet.* 40, 575–583.
- Willer, C.J., Li, Y., and Abecasis, G.R. (2010). METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 26, 2190–2191.
- Winkler, T.W., Day, F.R., Croteau-Chonka, D.C., Wood, A.R., Locke, A.E., Mägi, R., Ferreira, T., Fall, T., Graff, M., Justice, A.E., et al.; Genetic Investigation of Anthropometric Traits (GIANT) Consortium (2014). Quality control and conduct of genome-wide association meta-analyses. *Nat. Protoc.* 9, 1192–1212.
- Wood, A.R., Esko, T., Yang, J., Vedantam, S., Pers, T.H., Gustafsson, S., Chu, A.Y., Estrada, K., Luan, J., Kutalik, Z., et al.; Electronic Medical Records and Genomics (eMEMERGE) Consortium; MIGen Consortium; PAGEGE Consortium; LifeLines Cohort Study (2014). Defining the role of common variation in the genomic and biological architecture of adult human height. *Nat. Genet.* 46, 1173–1186.
- Zheng, J., Erzurumluoglu, A.M., Elsworth, B.L., Kemp, J.P., Howe, L., Haycock, P.C., Hemani, G., Tansey, K., Laurin, C., Pourcain, B.S., et al.; Early Genetics and Lifecourse Epidemiology (EAGLE) Eczema Consortium (2017). LD Hub: a centralized database and web interface to perform LD score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. *Bioinformatics* 33, 272–279.