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# New insights into the genetics of primary open-angle glaucoma based on meta-analyses of intraocular pressure and optic disc characteristics

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## 1 Title

- 2 New insights into the genetics of primary open-angle glaucoma based on meta-analyses of
- 3 intraocular pressure and optic disc characteristics.
- 4

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## 112 Abstract

113 Primary open-angle glaucoma (POAG), the most common optic neuropathy, is a heritable disease. 114 Siblings of POAG cases have a ten-fold increase risk of developing the disease. Intraocular pressure 115 (IOP) and optic nerve head characteristics are used clinically to predict POAG risk. We conducted a 116 genome-wide association meta-analysis of IOP and optic disc parameters and validated our findings 117 in multiple sets of POAG cases and controls. Using imputation to the 1000 genomes (1000G) 118 reference set, we identified 9 new genomic regions associated with vertical cup disc ratio (VCDR) and 119 1 new region associated with IOP. Additionally, we found 5 novel loci for optic nerve cup area and 6 120 for disc area. Previously it was assumed that genetic variation influenced POAG either through IOP or 121 via changes to the optic nerve head; here we present evidence that some genomic regions affect 122 both IOP and the disc parameters. We characterized the effect of the novel loci through pathway 123 analysis and found that pathways involved are not entirely distinct as assumed so far. Further, we 124 identified a novel association between CDKN1A and POAG. Using a zebrafish model we show that 125 six6b (associated with POAG and optic nerve head variation) alters the expression of cdkn1a. In 126 summary, we have identified several novel genes influencing the major clinical risk predictors of 127 POAG and showed that genetic variation in CDKN1A is important in POAG risk. 128

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## 134 Introduction

135 In primary open-angle glaucoma (POAG), loss of retinal ganglion cells and nerve fibers manifests itself 136 clinically as optic nerve damage, which leads to visual field loss and, eventually in 15% of these with 137 visual loss to visual impairment and blindness (1-3). The optic nerve damage is characterized by an 138 increase in cup area, the central portion of the optic nerve head (or optic disc), and/or a decrease in 139 rim area (the area of the disc occupied by the retinal nerve fiber axons). This damage can be 140 quantified by the vertical cup-disc ratio (VCDR), comparing the vertical diameter of the cup with the 141 vertical diameter of the total optic disc. The VCDR ranges from 0 to 1. In the clinical setting, optic 142 nerve heads with a VCDR above 0.7 or an asymmetry between eyes above 0.2 are considered to be 143 suspect for glaucoma. The interpretation of VCDR depends on the disc area, i.e. discs with larger area 144 have on average a higher VCDR and the cut-off point between a normal and abnormal VCDR might be 145 higher (4).

146 Elevated intraocular pressure (IOP) is a well-recognized risk factor for POAG and current therapies 147 lower IOP by various mechanisms; eye pressure is considered to be normal between 8 and 21 mmHg, 148 although many eyes with eye pressure in this range can exhibit glaucomatous optic nerve features. 149 Analyses of first-degree relatives of POAG patients, have shown that the sib relative risk is  $10.4 \pm 2.5$ 150 (5). Classic twin studies in POAG are lacking and there is no consensus on heritability for POAG, but 151 work by Cuellar-Partida et al estimated the heritability of POAG based on genome-wide array data at 152 0.42 ± 0.09(6). Several genome-wide association studies (GWAS) have identified new POAG genes by 153 examining POAG directly or studying endophenotypes like VCDR and IOP(7-16). Several genes 154 associated with VCDR and IOP - CDKN2B-AS1, SIX6 (VCDR); and CAV1/CAV2, TMCO1, ABCA1 and 155 ARHGEF12 (IOP) - are associated with POAG. Notably, no genes have been genome-wide associated 156 with both VCDR and IOP. Charlesworth et al. previously found a genetic correlation between VCDR 157 and IOP (RhoG = 0.45, P = 0.0012), however, genes underlying this relationship have not yet been 158 identified(17).

- 159 The aims of this study were to (1) identify new genes associated with the POAG endophenotypes IOP,
- 160 VCDR, cup area, and disc area, and ultimately POAG, using the 1000 Genomes imputations reference
- 161 panel, and (2) investigate the genetic overlap between the different endophenotypes. To accomplish
- these aims we performed a meta-analysis of GWAS of these four traits within the International
- 163 Glaucoma Genetics Consortium (IGGC).

## 165 **Results**

#### 166 Intraocular pressure

167 We first conducted a meta-analysis of GWAS in the European cohorts, top hit variants were 168 replicated in the Asians meta-analysis. Next, we performed a meta-analyses of the European and 169 Asians cohorts. After removal of single nucleotide polymorphisms (SNPs) with minor allele frequency 170 (MAF) < 0.01 and low imputation quality, approximately 8 million SNPs were included. Whilst the 171 meta-analysis of individuals of European descent yielded no novel associations, combined meta-172 analysis of individuals of European and Asian descent (n = 37,930,  $\lambda$  = 1.05; Supplementary Material, 173 Figs S1a, S1b and S2b), yielded nine genomic regions reaching genome-wide significance, of which 174 eight genomic regions were already known (Supplementary Material, Figs S1a, S1b, S2b, and Table 175 S3)(9, 11, 13). The peak SNP in the new genomic region was rs55796939 on chromosome 11q25 near 176 ADAMTS8 (Supplementary Material, Figs S3 and S4). The estimated heritability (h<sup>2</sup>) of IOP using the 177 LD score regression method(18) and the summary statistics of European-only meta-analysis was 0.13.

178

#### 179 Vertical cup-disc ratio

180 In the meta-analysis of individuals of European descent (n = 23,899,  $\lambda$  = 1.08), 21 genomic regions 181 were genome-wide significant (Supplementary Material, Figs S5a, S6a and Table S4). Five genomic 182 regions were novel (near to the genes *RPE65* on chr. 1p31, *F5* on chr. 1q23, *PDZD2* on chr. 5p13.3, 183 RREB1 on chr. 6p25, and DGKB on chr. 7p21.2) (Supplementary Material, Figs S7 and S8); the other 184 genomic regions have been previously associated with VCDR or cup area, two highly correlated 185 traits(19-21). Of the five novel genomic regions, *RREB1* (p-value =  $4.13 \times 10^{-3}$ ) was nominally 186 significant in the analysis of individuals of Asian descent (n= 8,373,  $\lambda$  = 1.01). In the combined analysis 187 (n = 32,272,  $\lambda$  = 1.06), another four novel genomic regions, near to the genes VCAN on chr. 5q14.3, PSCA on chr. 8q24.2, ENO4 on chr. 10q25.3, and RBM23 on chr. 14q11.2 (Supplementary Material, 188

**Figs S5b** and **S6b**), were genome-wide significant leading to a total of nine (5+4) novel genomic regions associated with VCDR. Of these novel genomic regions, *F5* has been associated with disc area previously(21). Disc area influences the VCDR(22), and therefore we corrected VCDR for disc area in a secondary analysis. After correction for disc area, the  $\beta$  (p-value) decreased from -0.007 (2.15 x 10<sup>-9</sup>) to -0.002 (5.60 x 10<sup>-2</sup>) in the subset with disc area available, suggesting that *F5* acts primarily on disc area and secondary to VCDR through its relation to disc area. The calculated h<sup>2</sup> of VCDR using the European -only meta-analysis was 0.31.

196

#### 197 Cup area

198 The meta-analysis of individuals of European descent (n = 22,489,  $\lambda$  = 1.06) yielded 17 genome-wide 199 significant regions of which 14 regions were already implicated for cup area or VCDR (Supplementary 200 Material, Figs S9a, S10a and Table S5)(20, 21). There were three novel associations on chr. 1q42.11 201 near CDC42BPA, chr. 8q21.11 near CRISPLD1, and on chr. 15q26.3 near FAM169B (Figs S11 and S12). 202 CDC42BPA has previously been associated with disc area and the fact that the association with cup 203 area adjusting for disc area is genome wide significant suggests an independent effect on cup area. In 204 the combined analysis of European and Asian individuals (n = 29,828,  $\lambda$  = 1.06, **Supplementary** 205 Material, Figs S9b and S10b) all loci except FAM169B and CRISPLD1 remained genome-wide 206 significant, and there was one additional genome-wide significant SNPs at chr. 6p21.2 (CDKN1A) and 207 one highly suggestive significant SNP at chr. 9q34.2 (ABO; previously associated to IOP). For cup area

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#### 210 Disc area

the estimated  $h^2$  was 0.27.

The meta-analysis of individuals of European descent (n = 22,504,  $\lambda$  = 1.06) resulted in 13 genome-

wide significant regions, of which two were not previously associated with disc area: *UGT8* on chr.

4q26 and *CTNNA3* on chr. 10q22.2 (**Supplementary Material, Figs S13a**, **S14a**, **S15**, **S16**, and **Table S6**). These SNPs were not significant in the meta-analysis of individuals of Asian descent (n = 7,307,  $\lambda$ = 1.02). An additional four SNPs reached genome-wide significance in the combined meta-analysis (n = 29,811,  $\lambda$  = 1.07): *PRDM16* on chr. 1p36.23-p33, *GADD45A* on chr. 1p31.2, *VGLL4* on chr. 3p25.3, and *ASB7* on chr. 15q26.3 (**Supplementary Material, Figs S13b** and **S14b**). The estimated h<sup>2</sup> for disc area was 0.27.

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#### 220 Characterization of the lead association signals

221 In total, 82 SNPs were associated with one or more of the above endophenotypes. Functional 222 characterization of the 82 SNPs was performed using a range of bioinformatics tools (see Methods). In total, 650 variants in linkage disequilibrium (LD) with the 82 lead SNPs (R<sup>2</sup>>0.8) were examined for 223 224 functional annotation. Overall, 61% (50/82) of the associated loci are in LD with variants located in 225 regulatory regions according to the ENCODE data (e.g. DNase I hypersensitive sites, transcription 226 factor binding sites and motifs; see Supplementary Material, Table S7). We investigated the 227 expression levels of the identified candidate genes using the UniGene database(23). Of all reviewed 228 genes, CDKN1A, PAX6 and DUSP1 showed the highest number of transcripts per million in the eye 229 (Supplementary Material, Table S8). According to the Ocular Tissue database(24), CDKN1A is highly 230 expressed in the optic nerve head, as well as DUSP1, which also shows high expression in the 231 trabecular meshwork. Both genes were associated with optic nerve head parameters. PAX6 is highly 232 expressed in the ciliary body and retina, in this study we found it associated with disc area. Other 233 highly expressed genes in the optic nerve include *EFEMP1* and *ABI3BP*, which are associated with cup 234 area and disc area, respectively (Supplementary Material, Table S9). To evaluate whether associated 235 genes are highly expressed in a particular tissue, we performed tissue enrichment analyses for each trait using suggestive SNPs (p-value <  $1.00 \times 10^{-05}$ ) in DEPICT(25). No FDR significant tissue enrichment 236 237 was found for IOP, VCDR or disc area. However, for cup area we found an enrichment for

membranes, joints, stem cells and other related connective tissues. Similar results were found when
 suggestive SNPs associated with VCDR, cup and disc area (the optic nerve parameters) were analyzed
 together (Table S10).

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## 242 Gene-based test

243 To identify new loci not found through per-SNP tests, we performed gene-based testing using VEGAS2. Reflecting the smaller number of tests, our gene-based significance threshold is P<sub>gene-based</sub> < 244  $0.05/24,769 = 2.02 \times 10^{-6}$  (24,769 genes tested). Using the gene-based test we found several novel 245 loci (Supplementary Material, Table S11). C9 was significantly associated with IOP (p-value 1.61 x 10<sup>-</sup> 246 <sup>6</sup>); *RARB* (p-value 1.86 x 10<sup>-6</sup>) and *HORMAD2-AS1* (p-value 1.04 x 10<sup>-6</sup>) were associated with VCDR. 247 248 These genes were previously associated with disc area, so the novel associations with VCDR could 249 possibly be driven by the influence of disc area on VCDR(21). In the cup area analysis, the genes *LRP10* (p-value  $1.20 \times 10^{-6}$ ) and *REM2* (p-value  $1.55 \times 10^{-6}$ ), and *THSD4* (p-value  $5.44 \times 10^{-8}$ ) were 250 significantly associated. The first two genes are located near to RBM23, which was significant in the 251 252 per-SNP test. THSD4 is located near to KPNB1, which was associated with VCDR in our previous meta-253 analysis(20). In the disc area analysis we found two genes that were significantly associated with disc area: ANKRA2 (p-value 8.42 x 10<sup>-7</sup>) and LOC149950 (p-value 3.87 x 10<sup>-7</sup>). 254

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#### 256 Characterizing the overlap in biological pathways involved in glaucoma endophenotypes

In total, 86 SNPs were associated with one or more of the above endophenotypes. The effect
estimates and p-values of these SNPs for all four endophenotypes are shown in Table 1-3. ADAMTS8
(IOP and VCDR, Table 1 and Table2b) and ABO (IOP and cup area, Table 1) were genome-wide
significantly associated with two traits. Of note is that there were different variants involved in
ADAMTS8: rs55796939 for IOP and rs4936099 for VCDR (r2=0.03 between these SNPs in 1000G

262 European samples). Figure 1 shows the overlap in associations across endophenotypes – we depict 263 annotated genes for which at least one SNP was genome-wide significant in at least one trait. 264 Overlap is defined as nominal significance or stronger for the second trait. The figure shows as 265 expected a strong overlap in variants associated to disc area, cup area and VCDR. Further, overlap is 266 noted in genes associated to IOP, cup area and VCDR. We next explored the genetic overlap between 267 optic disc parameters and IOP using our GWAS results from the European-only meta-analyses, which 268 comprised a larger sample size (n=22,489-29,578). As expected genetic correlation between optic 269 disc parameters was high, VCDR and cup area showed the highest correlation (RhoG = 0.83, p-value < 1.0 x 10<sup>-308</sup>), followed by VCDR and disc area (RhoG = 0.62, p-value =  $7.36 \times 10^{-23}$ ). Genetic correlation 270 between cup and disc area was 0.31 (p-value =  $1.00 \times 10^{-04}$ ). No significant genetic correlation was 271 272 found between IOP and optic disc parameters, the highest correlation was between IOP and VCDR 273 (RhoG = 0.06, p-value 0.3) (Table S12).

274 To further characterize the overlap in biological functions, gene set enrichment of loci associated 275 with IOP and optic disc parameters was performed using DEPICT(25). We first investigated enriched 276 pathways or gene sets using only genome-wide associated SNPs. No significant pathways were found 277 after FDR correction. However, pathways involved in metabolic processes such as "increased 278 circulating leptin level", "abnormal fat cell morphology" and "increased insulin sensitivity" were 279 suggestive when we analyzed the list of SNPs associated with VCDR, cup area and disc area (FDR<0.2, 280 see Supplementary Material, Table S13). We next searched for enriched pathways using suggestive SNPs (p-value <1.0 x 10<sup>-5</sup>). We further investigated potential overlap in pathways across the 281 282 endophenotypes, and found 57 significant pathways when using VCDR, cup area and IOP variants; 283 and 100 pathways when analysing suggestive VCDR, cup area and disc area variants. Note that in the 284 first analysis we investigated pathways enriched when IOP genes are taken into account, while in the 285 second one we analysed genes influencing the optic nerve head characteristics. Due to a high degree 286 of redundancy between pathways, we clustered the significant pathways into meta-pathways,

287 resulting in 11 meta-pathways for VCDR, cup area and IOP (Figure 2a, Supplementary Material, 288 Table S14); and 17 for VCDR, cup area, and disc area (Figure 2b, Supplementary Material, Table 289 **\$15**). Most of the gene sets found in both analyses highlighted pathways involved in cell 290 differentiation, notch signaling, regulatory DNA binding and embryonic development, which reflects 291 the pathways found when VCDR and CA variants are analyzed (Supplementary Material, Fig S17). Furthermore, we found "abnormal fat cell morphology" and "abnormal liver morphology" 292 293 significantly enriched; a key gene in these pathways is ABCA1. When IOP genes are included the 294 elongation factor, RNA Polymerase II (ELL2) protein complex" shows an enrichment. When disc area 295 genes are included, pathways such as "blood vessel development", "protein import into nucleus", 296 "Thrombospondin 1 (THBS1) and SMAD3 protein complex", and "abnormal eye morphology" were 297 significant. Key genes in the latter include: CDKN2B, FAT4, LRIG3, SIX6, COL8A1, SOX11, RND3, BOC, 298 WNT2B and CYP26A1.

299

## 300 From endophenotypes to primary open-angle glaucoma

301	To evaluate the implications of our findings in the context of glaucoma, we examined the association
302	between the genome-wide significant SNPs found in this study and 4 independent POAG studies (n=
303	6,429 cases and 41,404 controls). In total, 75 independent (i.e. $R^2 < 0.8$ ) SNPs associated with one or
304	more of the endophenotypes were assessed in the case/control studies. Of these, 32 were nominal
305	significantly associated with POAG (p-value < 0.05; the chance that 32 SNPs of 75 SNPs have a p-value
306	<0.05 is < 2.2 x $10^{-16}$ ), and 11 independent SNPs were Bonferroni significantly associated with POAG
307	(p-value 0.05/75 = 6.67 x $10^{-4}$ ) ( <b>Table 4</b> ). Two out of the 11 Bonferroni significant SNPs, the
308	rs2487048 in the ABCA1 gene and the 11:120357425 in the ARHGEF12 showed high heterogeneity
309	(I <sup>2</sup> ). To estimate the common effect size we performed a random effect meta-analysis. The odds ratio
310	(OR) remained almost the same for both variants, although p-values were not significant after
311	adjusting for multiple testing, which is in line with the heterogeneity observed. All other nine SNPs

312 surpassed the Bonferroni threshold for significance in both fixed and random-effect models. The association between CDKN1A and POAG is novel (OR = 1.14, p-value =  $7.4 \times 10^{-7}$ ). In our previous 313 paper, the SNP rs6054374 near to BMP2 was already associated with POAG (OR = 0.92, p-value 3.74 x 314 10<sup>-3</sup>), but the most significantly associated SNP in the current meta-analysis rs6107845 near to BMP2 315 shows a slightly larger effect on POAG (OR = 0.89, p-value =  $8.52 \times 10^{-6}$ ). CDKN1A, the novel 316 317 associated POAG candidate gene belongs to the cyclin dependent kinase inhibitor (CDKN) gene family 318 as well as CDKN2A, CDKN2B/CDKN2B-AS1, which all lie in a well-known glaucoma associated locus on 319 chr.9p21.

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#### 321 Expression of cdkn1a after knockdown of six6b in zebrafish

322 Previous studies have shown that the transcription factor SIX6 alters the expression of CDKN2A and 323 CDKN2B (26, 27). All three genes (SIX6, CDKN2A and CDKN2B) are in loci associated with POAG, 324 suggesting a possible functional link between these POAG-loci. Given that CDKN1A is part of the 325 same gene family as CDKN2A and CDKN2B we tested whether SIX6 also regulates CDKN1A. To assess 326 the potential for functional regulation of CDKN1A through SIX6, we first performed in silico analyses 327 and observed that SIX6 binds to CDKN1A (core score = 0.812). Then we used a previously studied 328 zebrafish model(27), in which we tested whether knockdown of *six6b* alters the expression of *cdkn1a* 329 in vivo. Knockdown of six6b was achieved using morpholino technology(27). 85% of the knockdown 330 embryos showed a small eye phenotype, reduced optic nerve thickness and an up-regulation of the 331 expression levels of cdkn2a/cdkn2b, as observed in previous studies (n=220)(27, 28). In zebrafish, there is only one gene which is analogous to the human CDKN2A and CDKN2B and it is referred to in 332 333 this paper as *cdkn2a/cdkn2b*. We evaluated the expression levels of *cdkn1a* in eyes of *six6b* deficient 334 embryos by RT-qPCR. A 41-fold overexpression of *cdkn1a* in the eye of *six6b* knockdown embryos 335 was found (p-value = 0.001) (Figure 3), showing that in vivo downregulation of six6b affects the

expression levels not only of *cdkn2a/cdkn2b* but also of *cdkn1a*, likely by binding to their sequence,
repressing their expression.

338

## 339 Discussion

This meta-analysis within the IGGC identified a novel genomic region associated with IOP, nine genomic regions associated with VCDR, five with cup area, and six with disc area. Eleven genomic regions were associated with POAG. Of these regions, the association between *CDKN1A* and POAG is novel.

We identify some specific loci that underlie the genetic correlation between IOP and VCDR described 344 345 earlier(17). ADAMTS8 and ABO were genome-wide significant for both IOP and VCDR or cup area. 346 Variants found close to ABO (rs8176672 for cup area and rs8176741 for IOP) are in LD (r2 >0.85) with 347 rs12216891, which lies in an enhancer and promoter histone mark, suggesting a potential regulatory 348 mechanism in that region. Furthermore, TRIOBP is genome-wide significant for cup area, and reached a p-value of 3.42 x 10<sup>-6</sup> for IOP. Interestingly, *TRIOBP* is approximately 180 kb away from *CARD10* 349 350 which is associated with disc area. There is a large overlap between VCDR/cup area and disc area. 351 Since VCDR is related to disc area, it might be that the effect found for VCDR is due to the effect of 352 disc area. Most of these overlapping genes are still Bonferroni significant in the cup area analysis in 353 which we corrected for disc area. Only CDC7/TGFBR3 and F5 are genome-wide significant for VCDR as 354 well as for disc area, but the effect is negligible after correction for disc area, suggesting that these 355 two genes play primarily a role in disc area.

356 When suggestive SNPs (p-value  $<1.0 \times 10^{-5}$ ) for VCDR and cup area are analyzed together using

357 DEPICT, we found an enrichment of pathways involved in cell differentiation, development,

358 regulatory DNA binding and Notch signaling. Including disc area SNPs to the VCDR and cup area

analysis reveals additional joint pathways: 1) eye and blood vessel development, 2) cancer, 3) protein

360 import into nucleus, and 4) thrombospondin 1 and SMAD3 complexes, related to the extracellular 361 matrix. The extracellular matrix pathway has been previously implicated in optic nerve 362 degeneration(20). ADAMTS8 , COL8A1 and the novel identified gene VCAN (versican) are involved in 363 the metabolism and composition of the extracellular matrix. Interestingly, mutations in VCAN have 364 been implicated in several ophthalmologic disorders(29).Of interest, known POAG genes also fit in 365 the pathways identified in this study, for example GAS7 and SIX6 play a role during development(27, 366 30), TGFBR3 has been implicated in extracellular matrix regulation(31) and in cancer as well as 367 GMDS(32).

368 Common variants in CDKN2B and its antisense CDKN2B-AS1 have been robustly found associated 369 with POAG. The gene CDKN1A, also known as p21, CIP-1 or WAF-1, belongs to the same family as 370 CDKN2B and also encodes a cyclin-dependent kinase inhibitor. Upregulation of CDKN1A causes G1 371 arrest and inhibits proliferation of the cell. Herein, for the first time, we provide genome-wide 372 significant evidence for association of CDKN1A variants with cup area. Two prior small cohort studies 373 suggested a possible role of CDKN1A in POAG. Tsai et al.(33) found an association between a codon 374 31 polymorphism in CDKN1A and POAG in 58 patients and 59 controls from China (OR = 2.39 [1.14-375 5.01]). Saglar et al. found no statistically significant association between the codon 31 polymorphism 376 and POAG in 75 patients and 119 controls from Turkey (OR = 1.70, p-value = 0.25)(34). Our study 377 provides strong evidence for the role of CDKN1A in POAG risk in a large sample consisting of 6,429 378 cases and 41,404 controls and shows the first convincing evidence for association of CDKN1A and 379 POAG in individuals of European descent. A potential functional interaction between POAG-loci (i.e. 380 SIX6 and CDKN genes) has been shown previously (26, 27). We performed in vivo studies in 381 embryonic zebrafish eye and found that knockdown of *six6b* upregulates both *cdkn2a/cdkn2b* and 382 cdkn1a. The mechanism behind the genetic variation in SIX6, and the CDKN genes remains obscure. 383 However, in a recent study, Skowronska-Krawczyk et al. showed that SIX6 regulates the expression of 384 CDKN2A in the context of IOP elevation (26). More comprehensive studies at the individual tissue

level e.g. retinal ganglion cell layer or optic nerve should be performed to evaluate the consequences
 of the genetic variation associated with POAG; for example, if associated variants in *SIX6* lead to up regulation of CDKN genes.

388 Genetic variation in CDK inhibitor genes has been found associated with various diseases and traits. 389 SNPs close to CDKN1A, for example, have been associated with electrocardiographic measures (35) 390 and colorectal cancer(36). While the CDKN2A/CDKN2B and CDKN2B-AS1 locus has been found in 391 GWAS of Type 2 Diabetes(37), fasting glucose(38), intracranial aneurysm(39), myocardia 392 infraction(40), coronary heart disease(41) and various cancer, including basal cell carcinoma(42), 393 breast(43) and prostate(44) cancer, and glioma(45). Interestingly, the same allele in CDKN2B that 394 decrease risk of POAG, was found associated with increased risk of glioma. The functional 395 significance of the genetic variation in the CDK genes remains to be determined, possible mechanism 396 in POAG might be related with the disruption of a delicate balance between proliferation and 397 apoptosis.

398 It has been suggested that p53 plays a role in POAG, especially in POAG with paracentral visual field 399 loss(46). Intriguingly, p53 has been also related to the CDK-inhibitors and to four of the new genes 400 pointed out by this study (GADD45A, PDZD2, RREB1 and PSCA). GADD45A is involved in growth arrest 401 through p53-dependent and independent mechanisms(47, 48) and can interact via CDKN1A(49). 402 PDZD2 is a tumor suppressor gene, reported to activate p53 by transcriptional regulation (50). RREB1 403 has an effect on p53 by binding to its promotor and transactivates its expression(51). While PSCA 404 expression correlates with the expression of p53 in choriocarcinoma, suggesting a role of PSCA in cell 405 growth through p53-related pathways(52). Other genes play a role in apoptosis or cell growth via 406 other mechanisms than p53: VGLL4 inhibits Bax- and TNFa-induced apoptosis(53) and DGKB is a regulator of diacylglycerol, which is important for cell growth and differentiation. UGT8 plays a role in 407 408 the biosynthesis of the sphingolipids of myelin membranes of the central and peripheral nervous 409 system; sphingolipids are also implicated in apoptosis(54).

410 Another interesting novel gene is *RPE65* (retinal pigment epithelium -specific protein 65kDa). This 411 gene has been associated with retinitis pigmentosa (RP) (55, 56) and Leber congenital amaurosis type 412 2 (LCA2)(57). As the name implies, the encoded protein is located in the retinal pigment 413 epithelium(58). Both diseases (RP and LCA2) are not characterized by an excavation of the optic 414 nerve head. However, we have checked several online databases for expression in different tissues. 415 In the eye, it is also highly expressed in the optic nerve head (S8 and S9 Tables) suggesting that this 416 gene could be involved in other ocular processes. Little expression is found in the brain, with no 417 expression in other tissues or organs in the body. Future studies are necessary to confirm our finding.

418 Of the genes identified by gene-based testing, C9 (complement component 9) is especially

419 interesting. Its protein is part of the membrane attack complex (MAC), together with the proteins

420 C5b, C6, C7, and C8. This complex activates several steps that lead to cell death, and cells protect

422 endocytosis and the CAV1/CAV2 genes are associated with IOP and POAG. It has been shown that

themselves by removing the complex through endocytosis. Caveolin is one of the proteins involved in

423 inhibition of caveolin-1 inhibits the endocytosis of MAC(59).

421

424 To our best knowledge, this meta-analysis is the largest study of IOP and optic nerve head 425 parameters to date, using well-characterized datasets from populations world-wide. A limitation of 426 our study is the lack of an available dataset for replication of the novel associations detected by 427 combined European and Asian ancestry samples. However, the heterogeneity of these novel genomic 428 regions is generally low in the meta-analysis. For VCDR, cup area, and disc area we have identified 429 novel SNPs in the analysis of individuals with European ancestry. Of the nine novel associations found 430 in these populations (RPE65, PDZD2, RREB1, DGKB for VCDR; CDC42BPA, CRISPLD1 and FAM169B for 431 cup area; and CTNNA3 and UGT8 for disc area), only RREB1 was nominally significant in the 432 individuals with Asian ancestry. Five of the seven non-significant SNPs in the individuals with Asian 433 ancestry had an effect estimate in the same direction. As the analysis in individuals with Asian 434 ancestry contains a smaller number of individuals, this could be due to lack of power.

435 We have identified 21 genetic variants associated with POAG endophenotypes: IOP, VCDR, cup area, 436 and disc area. The effect estimates of single SNP associations are small. We expected small effects of 437 many SNPs, since glaucoma endophenotypes are highly polygenic traits(60). Although individual SNP 438 effects are small, the joint effect of the various genetic variants could yield a clinical relevant 439 parameter. These association results do not imply that the variants described here have a causal 440 effect. Fine-mapping and functional studies are required to identify the causal variants tagged by our 441 findings and the exact molecular mechanisms involved in POAG. Although the overlap between IOP-442 loci and the optic disc parameters-loci is not large, this is the first study showing a genome-wide 443 significant evidence of the genetic correlation between IOP and VCDR; we expect that larger sample 444 sizes and improved imputation accuracy may help to find more of the loci underlying the genetic 445 correlation between these two endophenotypes. Of the novel associations, CDKN1A is strongly 446 associated with POAG, This finding is in line with other studies(26), pointing to the CDK-inhibitor 447 genes as key players in the development of POAG. The p53 pathway has been implicated in POAG 448 and interact in different cellular contexts with four of the new genes pointed out by this study 449 (GADD45A, PDZD2, RREB1 and PSCA). Functional studies need to be performed to assess the role of 450 p53 and CDK-inhibitors in the pathophysiology of POAG. A more comprehensive study of these 451 mechanisms may inform the development of new therapies for POAG.

## 453 Materials and methods

#### 454 Study design

455 We performed a meta-analysis on directly genotyped and imputed SNPs to the 1000 Genomes 456 reference panel. We analyzed four outcomes: IOP, VCDR, cup area, and disc area; the same approach 457 was used for all four outcomes. In the first stage, we conducted a meta-analysis of GWAS including 458 22,489-29,578 individuals of European ancestry. Subsequently, we evaluated the genome-wide 459 significant SNPs from the first stage in a meta-analysis of GWAS including 7,307-8,373 individuals of 460 Asian ancestry. Finally, we performed a mega/meta-analysis of GWAS from all individual studies 461 including individuals of European and Asian ancestry. We subsequently tested the effect of all 462 genome-wide significant SNPs on POAG in four independent case-control studies (n = 6,429/41,404). 463 These cases are not part of our primary meta-analyses which focused on quantitative traits in the 464 general population (IOP, VCDR, cup area and disc area). Three of the case-control studies were of 465 European ancestry(7, 61, 62) and one of Asian ancestry(63).

466

#### 467 Subjects, phenotyping and genotyping

468 All 19 studies included in this meta-analysis are part of the IGGC (S1a Table). Details for each 469 individual study can be found in Supplementary Material and Tables S1b, S1c and S2. The 470 ophthalmological examinations included measurements of IOP and optic nerve head assessment. All 471 19 studies contributed to the IOP mega/meta-analysis, 18 to the VCDR and 16 to the cup area and 472 disc area mega/meta-analysis .Validation of mega/meta-analysis results was performed in four 473 independent POAG case-control studies. Studies performed genomic imputation using 1000 474 Genomes phase 1 reference samples . Study-specific quality control can be found in the 475 **Supplementary Material.** All studies were performed with the approval of their local medical ethics

476 committee, and written informed consent was obtained from all participants in accordance with the477 Declaration of Helsinki.

478

## 479 Statistical analysis

480 In the IOP analysis, individuals who underwent IOP-lowering laser or surgery were removed from the 481 analysis; in individuals receiving IOP-lowering medication, the IOP value was multiplied by 1.3 to 482 estimate a pre-medication IOP value(64). The mean IOP, VCDR, cup area, and disc area of both eyes 483 was used for the analyses. SNPs with MAF < 0.01 and imputation quality scores <0.3 (proper-info of 484 IMPUTE) or R2<0.3 (MACH) were removed from the analyses. Each individual study performed a 485 linear regression between each endophenotype (IOP, VCDR, cup area, and disc area) and the SNPs, 486 under the assumption of an additive model for the effect of the risk allele. Analyses were adjusted 487 for age, sex and the first five principal components (for population-based studies) or family structure 488 (for family-based studies).

489 We performed an inverse variance weighted fixed-effect meta-analysis with METAL software(65). P 490 values for heterogeneity were calculated by using the Cochran's Q-test for heterogeneity. SNPs with 491 a p-value for heterogeneity <0.001 were removed from the results, as well as SNPs only present in 492 three studies. We used the 'genomic control' option in METAL to correct the standard error of each 493 individual study for estimated genomic inflation. To evaluate if our results exhibit signs of population 494 stratification we estimated the intercept using the LD score regression method(18) in the European-495 only meta-analyses. The intercept, which estimates inflation after removing polygenic signals, was: 496 1.0142 for IOP, 1.0199 for VCDR, 1.0074 for cup area and 1.0167 for disc area (Table S16).

In the meta-analyses of individuals with European ancestry, a p-value <5.0 x 10<sup>-8</sup> (the genome-wide
threshold of association) was considered significant. In the second stage, these genome-wide
significant SNPs were validated in individuals with Asian ancestry, and in this look-up a p value <0.05</li>

500 was considered significant. Finally, in the mega/meta-analysis of individuals with European and Asian ancestry a p-value of  $<5.0 \times 10^{-8}$  was considered significant. In total, we identified 75 independent 501 502 SNPs across different genomic regions for all the traits together. Therefore, the significance level 503 after Bonferroni correction in the meta-analysis of POAG cohorts was =  $6.67 \times 10^{-4} (0.05 / 75)$ 504 independent SNPs). To estimate the common effect size of the top SNPs associated with IOP, optic 505 disc parameters and their effect in the look-up in the POAG cohorts a random-effect meta-analysis 506 was performed using plink(66) http://pngu.mgh.harvard.edu/purcell/plink/ parameter --meta-507 analysis. Manhattan, regional and forest plots were made using R(67) and LocusZoom(68). 508 Genetic correlation between optic disc parameters (VCDR, cup and disc area) and IOP was estimated 509 using the LD score regression method(18, 69). GWAS summary statistics from the Europeans-only 510 meta-analyses were used to calculate the genetic overlap. To restrict the analyses to well imputed 511 SNPs, we included only SNPs with MAF > 0.01 that were present in HapMap3. GWAS data were harmonized using the "munge\_sumstats.py" function. For analyses we used the pre-computed LD 512 513 scores for Europeans "eur\_w\_ld\_ch" available at https://github.com/bulik/ldsc/wiki. The ldsc.py 514 function (with all default settings) was used to calculate the genetic correlation between traits. In addition, we calculated the heritability (h<sup>2</sup>) estimates from the GWAS meta-analysis using the same 515 516 function.

#### 517 Gene-based test using VEGAS

A gene-based test was performed using the VEGAS2 software(70), with a 50kb gene boundary. We used the parameter '-top 100' (default) to perform gene-based tests. This parameter considers association test statistics of all variants mapped to a gene to compute gene-based test statistics. The 1000 Genomes European and Asian populations were used as a reference to calculate LD for European and Asian ancestry data respectively. After calculation of gene-based test statistics for Asian and European ancestry populations separately, meta-analyses were conducted using Fisher's method for combining p-values.

525	Functional characterization, expression data, zebrafish and gene-set enrichment
526	We investigated for evidence of regulatory functions of associated loci HaploReg version 2(71) and
527	Regulomedb version 1.1(72). We investigated the expression of the associated genes using NCBI's
528	UniGene(23) and The Ocular Tissue Database(24). We also investigated the expression of <i>cdkn1a</i> in a
529	six6b knockdown zebrafish and used DEPICT to investigate gene-set enrichment. More information
530	about these analyses can be found in the Supplementary Material.
531	Data access
532	Access to the summary association statistics for IOP and optic disc parameters is available at
533	http://getfiles.qimr.edu.au/puyaGharahkhani/1000G_imputed_IGGC_eye_quantitative_traits/
534	

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541	
542	Conflict of interest
543	Dr. Pasquale has been a paid speaker for Allergan. He also served as a nonpaid consultant to Novartis
544	and a paid consultant to Bausch + Lomb. He has received support to travel to the Exfoliation
545	Glaucoma Think Tank Meeting in NYC by the Glaucoma Foundation.
546	Dr. Jonas: Consultant for MundiPharma Co.; Allergan Inc.; Merck Sharp & Dohme Co., Inc.; Alimera
547	Co.; Boehringer Ingelheim Co., Sanofi Co., Pfizer Co.; Patent holder with CellMed AG, Alzenau,
548	Germany and with University of Heidelberg / Germany
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## 555 References

556 Weinreb, R.N., Aung, T. and Medeiros, F.A. (2014) The pathophysiology and treatment of 1 557 glaucoma: a review. JAMA, **311**, 1901-1911. 558 Ernest, P.J., Busch, M.J., Webers, C.A., Beckers, H.J., Hendrikse, F., Prins, M.H. and Schouten, 2 559 J.S. (2013) Prevalence of end-of-life visual impairment in patients followed for glaucoma. Acta 560 *Ophthalmol*, **91**, 738-743. 561 3 Peters, D., Bengtsson, B. and Heijl, A. (2014) Factors associated with lifetime risk of openangle glaucoma blindness. Acta Ophthalmol, 92, 421-425. 562 Ramdas, W.D., Rizopoulos, D., Wolfs, R.C., Hofman, A., de Jong, P.T., Vingerling, J.R. and 563 4 564 Jansonius, N.M. (2011) Defining glaucomatous optic neuropathy from a continuous measure of optic 565 nerve damage - the optimal cut-off point for risk-factor analysis in population-based epidemiology. 566 Ophthalmic Epidemiol, 18, 211-216. 567 5 Wolfs, R.C., Klaver, C.C., Ramrattan, R.S., van Duijn, C.M., Hofman, A. and de Jong, P.T. (1998) 568 Genetic risk of primary open-angle glaucoma. Population-based familial aggregation study. Arch 569 Ophthalmol, 116, 1640-1645. Cuellar-Partida, G., Craig, J.E., Burdon, K.P., Wang, J.J., Vote, B.J., Souzeau, E., McAllister, I.L., 570 6 571 Isaacs, T., Lake, S., Mackey, D.A. et al. (2016) Assessment of polygenic effects links primary open-572 angle glaucoma and age-related macular degeneration. Sci Rep, 6, 26885. 573 7 Burdon, K.P., Macgregor, S., Hewitt, A.W., Sharma, S., Chidlow, G., Mills, R.A., Danoy, P., 574 Casson, R., Viswanathan, A.C., Liu, J.Z. et al. (2011) Genome-wide association study identifies 575 susceptibility loci for open angle glaucoma at TMCO1 and CDKN2B-AS1. Nat Genet, 43, 574-578. 576 Gharahkhani, P., Burdon, K.P., Fogarty, R., Sharma, S., Hewitt, A.W., Martin, S., Law, M.H., 8 577 Cremin, K., Bailey, J.N., Loomis, S.J. et al. (2014) Common variants near ABCA1, AFAP1 and GMDS 578 confer risk of primary open-angle glaucoma. Nat Genet, 46, 1120-1125. 579 9 Hysi, P.G., Cheng, C.Y., Springelkamp, H., Macgregor, S., Bailey, J.N., Wojciechowski, R., Vitart, V., Nag, A., Hewitt, A.W., Hohn, R. et al. (2014) Genome-wide analysis of multi-ancestry cohorts 580 581 identifies new loci influencing intraocular pressure and susceptibility to glaucoma. Nat Genet, 46, 582 1126-1130. 583 Thorleifsson, G., Walters, G.B., Hewitt, A.W., Masson, G., Helgason, A., DeWan, A., 10 584 Sigurdsson, A., Jonasdottir, A., Gudjonsson, S.A., Magnusson, K.P. et al. (2010) Common variants near 585 CAV1 and CAV2 are associated with primary open-angle glaucoma. Nat Genet, 42, 906-909. 11 586 van Koolwijk, L.M., Ramdas, W.D., Ikram, M.K., Jansonius, N.M., Pasutto, F., Hysi, P.G., 587 Macgregor, S., Janssen, S.F., Hewitt, A.W., Viswanathan, A.C. et al. (2012) Common genetic 588 determinants of intraocular pressure and primary open-angle glaucoma. PLoS Genet, 8, e1002611. 589 12 Ramdas, W.D., van Koolwijk, L.M., Lemij, H.G., Pasutto, F., Cree, A.J., Thorleifsson, G., 590 Janssen, S.F., Jacoline, T.B., Amin, N., Rivadeneira, F. et al. (2011) Common genetic variants 591 associated with open-angle glaucoma. *Hum Mol Genet*, **20**, 2464-2471. 592 13 Springelkamp, H., Iglesias, A.I., Cuellar-Partida, G., Amin, N., Burdon, K.P., van Leeuwen, E.M., 593 Gharahkhani, P., Mishra, A., van der Lee, S.J., Hewitt, A.W. et al. (2015) ARHGEF12 influences the risk 594 of glaucoma by increasing intraocular pressure. Hum Mol Genet, 24, 2689-2699. 595 14 Wiggs, J.L., Kang, J.H., Yaspan, B.L., Mirel, D.B., Laurie, C., Crenshaw, A., Brodeur, W., 596 Gogarten, S., Olson, L.M., Abdrabou, W. et al. (2011) Common variants near CAV1 and CAV2 are 597 associated with primary open-angle glaucoma in Caucasians from the USA. Hum Mol Genet, 20, 4707-598 4713. 599 Wiggs, J.L., Yaspan, B.L., Hauser, M.A., Kang, J.H., Allingham, R.R., Olson, L.M., Abdrabou, W., 15 600 Fan, B.J., Wang, D.Y., Brodeur, W. et al. (2012) Common variants at 9p21 and 8q22 are associated 601 with increased susceptibility to optic nerve degeneration in glaucoma. PLoS Genet, 8, e1002654. 602 16 Chen, Y., Lin, Y., Vithana, E.N., Jia, L., Zuo, X., Wong, T.Y., Chen, L.J., Zhu, X., Tam, P.O., Gong, 603 B. et al. (2014) Common variants near ABCA1 and in PMM2 are associated with primary open-angle 604 glaucoma. Nat Genet, 46, 1115-1119.

Charlesworth, J., Kramer, P.L., Dyer, T., Diego, V., Samples, J.R., Craig, J.E., Mackey, D.A.,
Hewitt, A.W., Blangero, J. and Wirtz, M.K. (2010) The path to open-angle glaucoma gene discovery:
endophenotypic status of intraocular pressure, cup-to-disc ratio, and central corneal thickness. *Invest Ophthalmol Vis Sci*, **51**, 3509-3514.

Bulik-Sullivan, B.K., Loh, P.R., Finucane, H.K., Ripke, S., Yang, J., Schizophrenia Working Group
of the Psychiatric Genomics, C., Patterson, N., Daly, M.J., Price, A.L. and Neale, B.M. (2015) LD Score
regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*, 47, 291-295.

Ramdas, W.D., van Koolwijk, L.M., Ikram, M.K., Jansonius, N.M., de Jong, P.T., Bergen, A.A.,
Isaacs, A., Amin, N., Aulchenko, Y.S., Wolfs, R.C. *et al.* (2010) A genome-wide association study of
optic disc parameters. *PLoS Genet*, **6**, e1000978.

Springelkamp, H., Hohn, R., Mishra, A., Hysi, P.G., Khor, C.C., Loomis, S.J., Bailey, J.N., Gibson,
J., Thorleifsson, G., Janssen, S.F. *et al.* (2014) Meta-analysis of genome-wide association studies

identifies novel loci that influence cupping and the glaucomatous process. *Nat Commun*, 5, 4883.
Springelkamp, H., Mishra, A., Hysi, P.G., Gharahkhani, P., Hohn, R., Khor, C.C., Cooke Bailey,
J.N., Luo, X., Ramdas, W.D., Vithana, E. *et al.* (2015) Meta-analysis of Genome-Wide Association

621 Studies Identifies Novel Loci Associated With Optic Disc Morphology. *Genet Epidemiol*, **39**, 207-216.

622 22 Ramdas, W.D., Wolfs, R.C., Hofman, A., de Jong, P.T., Vingerling, J.R. and Jansonius, N.M.

(2011) Heidelberg Retina Tomograph (HRT3) in population-based epidemiology: normative values
and criteria for glaucomatous optic neuropathy. *Ophthalmic Epidemiol*, **18**, 198-210.

625 23 Coordinators, N.R. (2015) Database resources of the National Center for Biotechnology 626 Information. *Nucleic Acids Res*, **43**, D6-17.

Wagner, A.H., Anand, V.N., Wang, W.H., Chatterton, J.E., Sun, D., Shepard, A.R., Jacobson, N.,
Pang, I.H., Deluca, A.P., Casavant, T.L. *et al.* (2013) Exon-level expression profiling of ocular tissues. *Exp Eye Res*, **111**, 105-111.

Pers, T.H., Karjalainen, J.M., Chan, Y., Westra, H.J., Wood, A.R., Yang, J., Lui, J.C., Vedantam,
S., Gustafsson, S., Esko, T. *et al.* (2015) Biological interpretation of genome-wide association studies
using predicted gene functions. *Nat Commun*, **6**, 5890.

Skowronska-Krawczyk, D., Zhao, L., Zhu, J., Weinreb, R.N., Cao, G., Luo, J., Flagg, K., Patel, S.,
Wen, C., Krupa, M. *et al.* (2015) P16INK4a Upregulation Mediated by SIX6 Defines Retinal Ganglion
Cell Pathogenesis in Glaucoma. *Mol Cell*, **59**, 931-940.

Iglesias, A.I., Springelkamp, H., van der Linde, H., Severijnen, L.A., Amin, N., Oostra, B., Kockx,
C.E., van den Hout, M.C., van Ijcken, W.F., Hofman, A. *et al.* (2014) Exome sequencing and functional
analyses suggest that SIX6 is a gene involved in an altered proliferation-differentiation balance early
in life and optic nerve degeneration at old age. *Hum Mol Genet*, **23**, 1320-1332.

Carnes, M.U., Liu, Y.P., Allingham, R.R., Whigham, B.T., Havens, S., Garrett, M.E., Qiao, C.,
Investigators, N.C., Katsanis, N., Wiggs, J.L. *et al.* (2014) Discovery and functional annotation of SIX6
variants in primary open-angle glaucoma. *PLoS Genet*, **10**, e1004372.

Black, G.C., Perveen, R., Wiszniewski, W., Dodd, C.L., Donnai, D. and McLeod, D. (1999) A
novel hereditary developmental vitreoretinopathy with multiple ocular abnormalities localizing to a
5-cM region of chromosome 5q13-q14. *Ophthalmology*, **106**, 2074-2081.

646 30 Hung, F.C., Cheng, Y.C., Sun, N.K. and Chao, C.C. (2013) Identification and functional

characterization of zebrafish Gas7 gene in early development. *J Neurosci Res*, **91**, 51-61.

64831Coulson-Thomas, V.J., Gesteira, T.F., Coulson-Thomas, Y.M., Vicente, C.M., Tersariol, I.L.,649Nader, H.B. and Toma, L. (2010) Fibroblast and prostate tumor cell cross-talk: fibroblast

differentiation, TGF-beta, and extracellular matrix down-regulation. *Exp Cell Res*, **316**, 3207-3226.

Alama Shakayama, K., Moriwaki, K., Imai, T., Shinzaki, S., Kamada, Y., Murata, K. and Miyoshi, E.

652 (2013) Mutation of GDP-mannose-4,6-dehydratase in colorectal cancer metastasis. *PLoS One*, **8**,

653 e70298.

- 654 33 Tsai, F.J., Lin, H.J., Chen, W.C., Tsai, C.H. and Tsai, S.W. (2004) A codon 31ser-arg 655 polymorphism of the WAF-1/CIP-1/p21/tumour suppressor gene in Chinese primary open-angle 656 glaucoma. Acta Ophthalmol Scand, 82, 76-80. 657 Saglar, E., Yucel, D., Bozkurt, B., Ozgul, R.K., Irkec, M. and Ogus, A. (2009) Association of 34 658 polymorphisms in APOE, p53, and p21 with primary open-angle glaucoma in Turkish patients. Mol 659 Vis, 15, 1270-1276. 660 Sotoodehnia, N., Isaacs, A., de Bakker, P.I., Dorr, M., Newton-Cheh, C., Nolte, I.M., van der 35 661 Harst, P., Muller, M., Eijgelsheim, M., Alonso, A. et al. (2010) Common variants in 22 loci are 662 associated with QRS duration and cardiac ventricular conduction. Nat Genet, 42, 1068-1076. 663 Dunlop, M.G., Dobbins, S.E., Farrington, S.M., Jones, A.M., Palles, C., Whiffin, N., Tenesa, A., 36 664 Spain, S., Broderick, P., Ooi, L.Y. et al. (2012) Common variation near CDKN1A, POLD3 and SHROOM2 665 influences colorectal cancer risk. Nat Genet, 44, 770-776. 666 Scott, L.J., Mohlke, K.L., Bonnycastle, L.L., Willer, C.J., Li, Y., Duren, W.L., Erdos, M.R., 37 667 Stringham, H.M., Chines, P.S., Jackson, A.U. et al. (2007) A genome-wide association study of type 2 668 diabetes in Finns detects multiple susceptibility variants. Science, **316**, 1341-1345. 669 38 Manning, A.K., Hivert, M.F., Scott, R.A., Grimsby, J.L., Bouatia-Naji, N., Chen, H., Rybin, D., Liu, 670 C.T., Bielak, L.F., Prokopenko, I. et al. (2012) A genome-wide approach accounting for body mass 671 index identifies genetic variants influencing fasting glycemic traits and insulin resistance. Nat Genet, 672 44, 659-669. 39 673 Foroud, T., Koller, D.L., Lai, D., Sauerbeck, L., Anderson, C., Ko, N., Deka, R., Mosley, T.H., 674 Fornage, M., Woo, D. et al. (2012) Genome-wide association study of intracranial aneurysms 675 confirms role of Anril and SOX17 in disease risk. *Stroke*, **43**, 2846-2852. 676 40 Myocardial Infarction Genetics, C., Kathiresan, S., Voight, B.F., Purcell, S., Musunuru, K., 677 Ardissino, D., Mannucci, P.M., Anand, S., Engert, J.C., Samani, N.J. et al. (2009) Genome-wide 678 association of early-onset myocardial infarction with single nucleotide polymorphisms and copy 679 number variants. Nat Genet, 41, 334-341. 680 41 Lu, X., Wang, L., Chen, S., He, L., Yang, X., Shi, Y., Cheng, J., Zhang, L., Gu, C.C., Huang, J. et al. 681 (2012) Genome-wide association study in Han Chinese identifies four new susceptibility loci for 682 coronary artery disease. Nat Genet, 44, 890-894. 683 42 Stacey, S.N., Helgason, H., Gudjonsson, S.A., Thorleifsson, G., Zink, F., Sigurdsson, A., Kehr, B., 684 Gudmundsson, J., Sulem, P., Sigurgeirsson, B. et al. (2015) New basal cell carcinoma susceptibility 685 loci. Nat Commun, 6, 6825. 686 43 Turnbull, C., Ahmed, S., Morrison, J., Pernet, D., Renwick, A., Maranian, M., Seal, S., 687 Ghoussaini, M., Hines, S., Healey, C.S. et al. (2010) Genome-wide association study identifies five new 688 breast cancer susceptibility loci. Nat Genet, 42, 504-507. 689 Al Olama, A.A., Kote-Jarai, Z., Berndt, S.I., Conti, D.V., Schumacher, F., Han, Y., Benlloch, S., 44 690 Hazelett, D.J., Wang, Z., Saunders, E. et al. (2014) A meta-analysis of 87,040 individuals identifies 23 691 new susceptibility loci for prostate cancer. Nat Genet, 46, 1103-1109. 692 45 Shete, S., Hosking, F.J., Robertson, L.B., Dobbins, S.E., Sanson, M., Malmer, B., Simon, M., 693 Marie, Y., Boisselier, B., Delattre, J.Y. et al. (2009) Genome-wide association study identifies five 694 susceptibility loci for glioma. Nat Genet, 41, 899-904. 695 Wiggs, J.L., Hewitt, A.W., Fan, B.J., Wang, D.Y., Figueiredo Sena, D.R., O'Brien, C., Realini, A., 46 696 Craig, J.E., Dimasi, D.P., Mackey, D.A. et al. (2012) The p53 codon 72 PRO/PRO genotype may be 697 associated with initial central visual field defects in caucasians with primary open angle glaucoma. 698 *PLoS One*, **7**, e45613. 699 47 Smith, M.L., Chen, I.T., Zhan, Q., Bae, I., Chen, C.Y., Gilmer, T.M., Kastan, M.B., O'Connor, 700 P.M. and Fornace, A.J., Jr. (1994) Interaction of the p53-regulated protein Gadd45 with proliferating 701 cell nuclear antigen. Science, 266, 1376-1380. 702 48 Kastan, M.B., Zhan, Q., el-Deiry, W.S., Carrier, F., Jacks, T., Walsh, W.V., Plunkett, B.S., 703 Vogelstein, B. and Fornace, A.J., Jr. (1992) A mammalian cell cycle checkpoint pathway utilizing p53
- and GADD45 is defective in ataxia-telangiectasia. *Cell*, **71**, 587-597.

705 49 Kearsey, J.M., Coates, P.J., Prescott, A.R., Warbrick, E. and Hall, P.A. (1995) Gadd45 is a 706 nuclear cell cycle regulated protein which interacts with p21Cip1. Oncogene, **11**, 1675-1683. 707 Tam, C.W., Liu, V.W., Leung, W.Y., Yao, K.M. and Shiu, S.Y. (2008) The autocrine human 50 708 secreted PDZ domain-containing protein 2 (sPDZD2) induces senescence or quiescence of prostate, 709 breast and liver cancer cells via transcriptional activation of p53. Cancer Lett, 271, 64-80. 710 51 Liu, H., Hew, H.C., Lu, Z.G., Yamaguchi, T., Miki, Y. and Yoshida, K. (2009) DNA damage 711 signalling recruits RREB-1 to the p53 tumour suppressor promoter. *Biochem J*, **422**, 543-551. 712 Feng, H.C., Tsao, S.W., Ngan, H.Y., Xue, W.C., Kwan, H.S., Siu, M.K., Liao, X.Y., Wong, E. and 52 713 Cheung, A.N. (2008) Overexpression of prostate stem cell antigen is associated with gestational 714 trophoblastic neoplasia. Histopathology, 52, 167-174. 715 53 Jin, H.S., Park, H.S., Shin, J.H., Kim, D.H., Jun, S.H., Lee, C.J. and Lee, T.H. (2011) A novel 716 inhibitor of apoptosis protein (IAP)-interacting protein, Vestigial-like (VgI)-4, counteracts apoptosis-717 inhibitory function of IAPs by nuclear sequestration. Biochem Biophys Res Commun, 412, 454-459. 718 54 Gomez-Munoz, A., Kong, J.Y., Salh, B. and Steinbrecher, U.P. (2004) Ceramide-1-phosphate 719 blocks apoptosis through inhibition of acid sphingomyelinase in macrophages. J Lipid Res, 45, 99-105. 720 55 Flicek, P., Amode, M.R., Barrell, D., Beal, K., Billis, K., Brent, S., Carvalho-Silva, D., Clapham, P., 721 Coates, G., Fitzgerald, S. et al. (2014) Ensembl 2014. Nucleic Acids Res, 42, D749-755. 722 56 Gu, S.M., Thompson, D.A., Srikumari, C.R., Lorenz, B., Finckh, U., Nicoletti, A., Murthy, K.R., 723 Rathmann, M., Kumaramanickavel, G., Denton, M.J. et al. (1997) Mutations in RPE65 cause 724 autosomal recessive childhood-onset severe retinal dystrophy. Nat Genet, 17, 194-197. 725 Marlhens, F., Bareil, C., Griffoin, J.M., Zrenner, E., Amalric, P., Eliaou, C., Liu, S.Y., Harris, E., 57 726 Redmond, T.M., Arnaud, B. et al. (1997) Mutations in RPE65 cause Leber's congenital amaurosis. Nat 727 Genet, 17, 139-141. 728 58 Hamel, C.P., Tsilou, E., Pfeffer, B.A., Hooks, J.J., Detrick, B. and Redmond, T.M. (1993) 729 Molecular cloning and expression of RPE65, a novel retinal pigment epithelium-specific microsomal 730 protein that is post-transcriptionally regulated in vitro. J Biol Chem, 268, 15751-15757. 731 59 Moskovich, O., Herzog, L.O., Ehrlich, M. and Fishelson, Z. (2012) Caveolin-1 and dynamin-2 732 are essential for removal of the complement C5b-9 complex via endocytosis. J Biol Chem, 287, 19904-733 19915. 734 60 Ramdas, W.D., Amin, N., van Koolwijk, L.M., Janssens, A.C., Demirkan, A., de Jong, P.T., 735 Aulchenko, Y.S., Wolfs, R.C., Hofman, A., Rivadeneira, F. et al. (2011) Genetic architecture of open 736 angle glaucoma and related determinants. J Med Genet, 48, 190-196. 737 Wiggs, J.L., Hauser, M.A., Abdrabou, W., Allingham, R.R., Budenz, D.L., Delbono, E., Friedman, 61 738 D.S., Kang, J.H., Gaasterland, D., Gaasterland, T. et al. (2013) The NEIGHBOR consortium primary 739 open-angle glaucoma genome-wide association study: rationale, study design, and clinical variables. J 740 Glaucoma, 22, 517-525. 741 62 Gibson, J., Griffiths, H., De Salvo, G., Cole, M., Jacob, A., Macleod, A., Yang, Y., Menon, G., 742 Cree, A., Ennis, S. et al. (2012) Genome-wide association study of primary open angle glaucoma risk 743 and quantitative traits. Mol Vis, 18, 1083-1092. 744 Lavanya, R., Jeganathan, V.S., Zheng, Y., Raju, P., Cheung, N., Tai, E.S., Wang, J.J., Lamoureux, 63 745 E., Mitchell, P., Young, T.L. et al. (2009) Methodology of the Singapore Indian Chinese Cohort (SICC) 746 eye study: quantifying ethnic variations in the epidemiology of eye diseases in Asians. Ophthalmic 747 *Epidemiol*, **16**, 325-336. 748 van der Valk, R., Webers, C.A., Schouten, J.S., Zeegers, M.P., Hendrikse, F. and Prins, M.H. 64 749 (2005) Intraocular pressure-lowering effects of all commonly used glaucoma drugs: a meta-analysis 750 of randomized clinical trials. *Ophthalmology*, **112**, 1177-1185. 751 Willer, C.J., Li, Y. and Abecasis, G.R. (2010) METAL: fast and efficient meta-analysis of 65 752 genomewide association scans. *Bioinformatics*, 26, 2190-2191. 753 Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M.A., Bender, D., Maller, J., Sklar, 66 754 P., de Bakker, P.I., Daly, M.J. et al. (2007) PLINK: a tool set for whole-genome association and

population-based linkage analyses. *American journal of human genetics*, **81**, 559-575.

- 756 67 Team., R.C. (2014) R: a language and environment for statistical computing,
- 757 <u>http://www.R-project.org</u>., in press.
- 758 68 Pruim, R.J., Welch, R.P., Sanna, S., Teslovich, T.M., Chines, P.S., Gliedt, T.P., Boehnke, M.,
- Abecasis, G.R. and Willer, C.J. (2010) LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics*, **26**, 2336-2337.
- 761 69 Bulik-Sullivan, B., Finucane, H.K., Anttila, V., Gusev, A., Day, F.R., Loh, P.R., ReproGen, C.,
- 762 Psychiatric Genomics, C., Genetic Consortium for Anorexia Nervosa of the Wellcome Trust Case
- 763 Control, C., Duncan, L. *et al.* (2015) An atlas of genetic correlations across human diseases and traits.
- 764 Nat Genet, **47**, 1236-1241.
- 765 70 Mishra, A. and Macgregor, S. (2015) VEGAS2: Software for More Flexible Gene-Based Testing.
   766 *Twin Res Hum Genet*, **18**, 86-91.
- 767 71 Ward, L.D. and Kellis, M. (2012) HaploReg: a resource for exploring chromatin states,
- conservation, and regulatory motif alterations within sets of genetically linked variants. *Nucleic Acids Res*, 40, D930-934.
- 770 72 Boyle, A.P., Hong, E.L., Hariharan, M., Cheng, Y., Schaub, M.A., Kasowski, M., Karczewski, K.J.,
- Park, J., Hitz, B.C., Weng, S. *et al.* (2012) Annotation of functional variation in personal genomes using
- 772 RegulomeDB. *Genome Res*, **22**, 1790-1797.

#### **Legends to Figures**

**Figure 1**. Overlap between the genes associated with one or more endophenotypes. Genes with a genome-wide significant association for at least one trait are shown. These genes are counted as overlapping genes if they are Bonferroni significantly associated with the other trait(s). Chr 11p11.2 (see intraocular pressure circle) means a region on chromosome 11p11.2 that is associated with IOP and has many genes in it; the likely causative gene in this region is not identified yet. Genes in bold are genes associated with primary open-angle glaucoma (POAG) in our meta-analysis of four case-control studies.\*Genes associated with familial forms of POAG (e.g. *MYOC* and *OPTN*) or found in case-control association studies which did not show an association with the endophenotypes explored in this study.

Figure 2. Pathways significantly enriched for: A) Loci associated with the vertical cup-disc ratio, cup area and intraocular pressure (p-value <7.0 x 10-6 in the GWAS). In total 11 meta-pathways were identified after clustering the 57 pathways identified by DEPICT. B) Loci associated with vertical cup-disc ratio, cup area and disc area (p-value <1.0 x 10-5). In total 17 meta-pathways were identified after clustering the 100 pathways identified by DEPICT. In both figures, meta-pathways are represented by nodes coloured according to statistical significance, and edges are scaled according to the correlation between meta-pathways. \*The pathway "Abnormal eye morphology" clustered with the meta-pathway "Chordate embryonic development". ELL2=Elongation Factor, RNA Polymerase II, DVL3= Dishevelled Segment Polarity Protein 3, THBS1=Thrombospondin 1, RFX2= Regulatory Factor X, 2.

## Figure 3. cdkn1a mRNA expression change

Overexpression of *cdkn1a* and *cdkn2a/cdkn2b* in response to six6b depletion is shown. All samples expression were normalized to the control gene sdha. Relative expression was calculated by setting the wild-type expression level at 1. Values represent mean ± standard error of the mean. \*P<0.05; \*\*P<0.005.

				IOP			VCDR		Cup area				Disc area	i
SNP	Nearest gene	A1/A2	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs10918274	TMCO1	t/c	0.26	0.04	5.64E-12	0.005	0.002	8.38E-03	0.010	0.003	2.47E-03	0.000	0.006	9.49E-01
rs7635832	FNDC3B	g/t	-0.22	0.03	6.61E-13	-0.001	0.001	3.35E-01	-0.004	0.003	1.27E-01	0.002	0.005	7.08E-01
rs10281637	CAV1/CAV2	c/t	0.20	0.03	3.96E-13	0.004	0.001	5.28E-03	0.006	0.003	1.23E-02	-0.002	0.005	6.01E-01
8:78380944*	ΡΚΙΑ	i/r	1.00	0.17	7.54E-09	0.000	0.010	9.74E-01	-0.018	0.017	3.00E-01	0.018	0.031	5.61E-01
rs7815043*	ΡΚΙΑ	c/t	-0.10	0.03	4.41E-05	-0.001	0.001	3.13E-01	-0.001	0.002	8.32E-01	-0.002	0.004	5.66E-01
rs7944735	Many genes	c/g	0.19	0.03	6.00E-11	0.001	0.001	4.37E-01	0.006	0.003	3.33E-02	0.000	0.005	9.68E-01
11:120357425	ARHGEF12	d/r	0.18	0.03	2.02E-09	0.001	0.001	6.12E-01	0.001	0.003	6.45E-01	0.001	0.005	8.38E-01
rs12794618	ARHGEF12	c/t	0.17	0.03	7.86E-09	0.001	0.001	4.14E-01	0.002	0.003	4.84E-01	0.004	0.005	4.53E-01
rs55796939	ADAMTS8	t/c	0.36	0.06	2.31E-08	0.003	0.003	3.61E-01	0.006	0.006	3.19E-01	-0.003	0.010	7.95E-01
rs2472496	ABCA1	g/a	0.17	0.02	1.93E-13	0.004	0.001	6.83E-05	0.010	0.002	9.63E-07	0.003	0.004	4.75E-01
rs8176741	ABO	a/g	0.24	0.04	3.47E-10	0.007	0.002	4.51E-05	0.019	0.003	7.12E-08	0.004	0.006	5.42E-01
rs9913911	GAS7	g/a	-0.17	0.02	7.01E-12	-0.006	0.001	1.84E-07	-0.008	0.002	2.48E-04	-0.001	0.004	8.41E-01

Table 1. Single nucleotide polymorphisms (SNPs) that are genome-wide significantly associated with IOP and show and association with vertical cup-disc ratio.

For these SNPs, the associations with the other traits are also included. SNPs that are Bonferroni significantly associated with other traits are shown in bold (p-value <  $5.37 \times 10^{-4}$ ; 0.05/93). In the first rows, the SNPs genome-wide significantly associated with intraocular pressure (IOP) are shown. Next, the SNPs associated with IOP, vertical cup-disc ratio (VCDR), and cup area are shown. Nearest gene, reference NCBI build37; A1, reference allele; A2, other allele;  $\beta$ , effect size on the endophenotype (IOP, VCDR, cup area or disc area) based on allele A1; SE, standard error of the effect size; i, insertion; d, deletion; r, reference. \*This locus was not considered as new because the signal came from only one INDEL (8:78380944). \*Genome-wide associated in the European-only meta-analysis

				IOP			VCDR			Cup area			Disc area	
SNP	Nearest	A1/A2	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs6804624	COL8A1	c/t	-0.01	0.03	6.54E-01	0.008	0.001	8.63E-12	0.013	0.002	1.99E-08	0.020	0.004	9.67E-07
rs7916697	ATOH7	a/g	0.01	0.03	7.43E-01	-0.018	0.001	2.46E-45	-0.017	0.002	1.32E-12	-0.094	0.004	1.34E-102
10:96008348	PLCE1	d/r	0.01	0.03	5.73E-01	0.007	0.001	4.57E-08	0.013	0.002	1.72E-08	0.015	0.004	2.22E-04
rs324780	TMTC2	g/a	0.03	0.02	2.79E-01	-0.011	0.001	7.16E-23	-0.016	0.002	1.57E-13	-0.029	0.004	8.58E-13
rs4299136	ASB7	c/g	-0.03	0.03	4.22E-01	0.010	0.002	2.68E-12	0.018	0.003	4.09E-10	0.024	0.005	4.02E-06
16:51461915	SALL1	r/i	0.02	0.03	4.34E-01	0.010	0.001	2.62E-13	0.013	0.003	6.78E-07	0.032	0.005	2.38E-12
rs4784295	SALL1	c/g	0.02	0.03	5.63E-01	0.009	0.001	3.93E-13	0.013	0.003	1.63E-07	0.031	0.005	1.12E-11
rs5752773	CHEK2	g/c	0.01	0.03	6.91E-01	-0.012	0.001	1.49E-20	-0.024	0.003	4.12E-21	-0.024	0.005	1.48E-07
rs2092172	CARD10	a/g	0.00	0.03	8.86E-01	0.009	0.001	3.08E-12	0.011	0.003	3.34E-05	0.032	0.005	1.44E-11
rs7717697	VCAN	c/t	0.01	0.02	7.21E-01	-0.007	0.001	6.66E-09	-0.009	0.002	1.19E-05	-0.018	0.004	4.84E-06
rs1681739	ENO4	t/c	0.03	0.02	2.23E-01	0.006	0.001	2.44E-08	0.011	0.002	3.70E-07	0.019	0.004	1.85E-06
rs60779155	ASB7	a/g	-0.02	0.04	6.61E-01	0.010	0.002	3.76E-10	0.019	0.003	3.75E-09	0.030	0.006	8.26E-08
rs1830890	PLCE1	g/a	0.01	0.02	8.14E-01	0.006	0.001	3.02E-08	0.012	0.002	1.06E-07	0.013	0.004	5.51E-04
rs482507	TMTC2	c/t	0.02	0.02	3.48E-01	-0.011	0.001	2.19E-19	-0.017	0.002	2.56E-14	-0.030	0.004	4.49E-13
rs4436712	SIX6	t/g	-0.04	0.02	1.47E-01	0.009	0.001	5.48E-14	0.025	0.002	1.50E-29	-0.018	0.004	6.59E-06
rs738722	CHEK2	t/c	0.02	0.03	3.57E-01	-0.012	0.001	4.94E-20	-0.024	0.003	7.81E-22	-0.021	0.005	2.63E-06
rs2684249	HSF2	c/t	0.03	0.02	2.08E-01	-0.006	0.001	1.64E-07	-0.012	0.002	3.04E-08	-0.015	0.004	1.49E-04
rs34222435	ASB7	t/c	-0.03	0.03	3.86E-01	0.010	0.002	3.07E-12	0.019	0.003	1.07E-10	0.025	0.005	2.98E-06

Table 2a. Single nucleotide polymorphisms (SNPs) that are genome-wide significantly associated with at least one of the optic disc parameters.

				IOP			VCDR			Cup area			Disc area	
SNP	Nearest	A1/A2	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs7916410	ATOH7	t/c	0.00	0.03	9.76E-01	-0.018	0.001	1.14E-45	-0.017	0.002	6.11E-12	-0.097	0.004	7.06E-109
rs442376	TMTC2	c/t	-0.03	0.03	3.09E-01	0.011	0.001	1.50E-17	0.017	0.002	3.18E-12	0.032	0.004	4.92E-14
rs1345467	SALL1	g/a	0.01	0.03	6.53E-01	0.009	0.001	4.96E-12	0.012	0.003	1.07E-06	0.032	0.005	6.41E-13
rs5762752	CHEK2	c/g	0.01	0.03	6.61E-01	-0.011	0.001	4.83E-18	-0.021	0.002	6.72E-19	-0.023	0.004	2.26E-08
rs11129176	RARB	a/g	0.02	0.03	4.17E-01	0.005	0.001	3.17E-05	0.010	0.002	1.01E-05	0.023	0.004	3.40E-08
rs1997404	COL8A1	g/t	-0.03	0.03	3.24E-01	0.008	0.001	2.39E-11	0.013	0.002	7.71E-08	0.024	0.004	1.90E-08
rs34935520	SIX6	g/a	-0.04	0.02	1.13E-01	0.009	0.001	7.95E-14	0.025	0.002	6.96E-29	-0.023	0.004	7.61E-08

For these SNPs, the associations with the other traits are also included. Here the SNPs genome-wide significantly associated with at least one of the optic disc parameters and that are Bonferroni significantly associated with the other disc parameters are shown in bold (p-value <  $5.37 \times 10^{-4}$ ; 0.05/93).Nearest gene, reference NCBI build37; A1, reference allele; A2, other allele;  $\beta$ , effect size on the effect size on the endophenotype (IOP, VCDR, cup area or disc area) based on allele A1; SE, standard error of the effect size; i, insertion; d, deletion; r, reference.

						VCDR			Cup area		Disc area			
SNP	Nearest gene	A1/A	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs1925953	RPE65*	t/a	-0.02	0.02	3.26E-01	0.006	0.001	1.55E-07	0.010	0.002	1.50E-05	0.006	0.004	1.08E-01
rs72759609	PDZD2	c/t	-0.04	0.05	3.50E-01	-0.012	0.002	7.10E-09	-0.020	0.004	1.98E-06	-0.021	0.008	5.62E-03
rs11450334	DUSP1	t/c	-0.12	0.08	1.27E-01	-0.021	0.004	1.31E-08	-0.035	0.007	2.90E-07	-0.035	0.013	5.83E-03
rs4960295	RREB1	a/g	0.02	0.02	4.75E-01	0.007	0.001	2.49E-10	0.009	0.002	3.73E-05	0.012	0.004	3.29E-03
rs10274998	DGKB	t/c	0.02	0.03	4.38E-01	0.008	0.001	4.68E-08	0.012	0.003	8.08E-06	0.011	0.005	2.65E-02
rs2157719	CDKN2B-AS1	c/t	-0.04	0.02	9.81E-02	-0.013	0.001	3.75E-35	-0.024	0.002	3.31E-28	-0.008	0.004	3.03E-02
rs3891783	PLCE1	g/c	0.04	0.02	1.01E-01	0.007	0.001	1.06E-10	0.011	0.002	3.28E-07	0.012	0.004	1.52E-03
rs1346	SSSCA1	t/a	-0.05	0.03	1.20E-01	-0.013	0.002	7.51E-18	-0.019	0.003	9.31E-11	-0.016	0.005	2.10E-03
rs4936099	ADAMTS8	c/a	-0.03	0.03	2.38E-01	-0.007	0.001	6.70E-09	-0.013	0.002	4.96E-08	-0.006	0.004	1.72E-01
13:3662990	DCLK1	d/r	-0.02	0.03	5.70E-01	0.007	0.001	2.98E-08	0.018	0.002	2.20E-14	-0.005	0.004	2.36E-01
rs7323428	DCLK1	t/g	-0.02	0.03	4.13E-01	0.007	0.001	1.86E-08	0.019	0.002	1.67E-15	-0.005	0.004	2.23E-01
rs8015152	SIX6	t/c	-0.06	0.02	2.27E-02	0.010	0.001	2.86E-18	0.024	0.002	8.15E-26	-0.011	0.004	6.18E-03
rs6107845	BMP2	a/g	0.03	0.02	2.80E-01	-0.009	0.001	3.44E-17	-0.017	0.002	2.90E-15	-0.004	0.004	3.27E-01
rs6764184	FLNB	t/g	0.05	0.03	5.03E-02	0.007	0.001	1.89E-08	0.015	0.002	1.30E-10	0.010	0.004	1.92E-02
rs7311936	FAM101A	c/g	-0.03	0.02	1.69E-01	-0.006	0.001	2.48E-09	-0.013	0.002	4.52E-09	0.003	0.004	5.14E-01
14:2338879	RBM23	r/d	0.02	0.03	3.99E-01	0.007	0.001	2.56E-08	0.013	0.003	2.01E-07	0.009	0.005	4.29E-02
rs3794453	RBM23	a/t	0.01	0.02	7.22E-01	0.007	0.001	7.25E-08	0.011	0.002	2.88E-07	0.009	0.004	3.11E-02
rs2252865	RERE	t/c	0.05	0.03	4.11E-02	0.005	0.001	2.66E-05	0.014	0.002	1.33E-09	0.003	0.004	5.08E-01

## Table2b. Single nucleotide polymorphisms (SNPs) that are genome-wide significantly associated with vertical cup-disc ratio or with cup area

				IOP			VCDR			Cup area			Disc area	
SNP	Nearest gene	A1/A	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs13016883	TRIB2	c/g	0.01	0.03	5.64E-01	0.006	0.001	3.44E-06	0.016	0.002	1.83E-11	0.001	0.004	8.30E-01
rs35084382	DUSP1	c/t	-0.10	0.07	1.32E-01	-0.018	0.003	2.05E-08	-0.033	0.006	2.17E-08	-0.031	0.011	5.51E-03
rs11759831	CRISPLD1*	t/g	-0.05	0.05	3.10E-01	0.009	0.002	1.07E-04	0.021	0.004	1.66E-06	0.022	0.008	5.47E-03
rs1360589	CDKN2B-AS1	c/t	-0.04	0.02	8.42E-02	-0.013	0.001	1.43E-34	-0.024	0.002	2.90E-28	-0.008	0.004	4.45E-02
rs11613189	FAM101A	t/c	-0.03	0.03	2.27E-01	-0.005	0.001	6.04E-06	-0.016	0.002	2.01E-12	0.002	0.004	6.42E-01
rs2251069	DDHD1	c/t	0.01	0.02	7.29E-01	-0.006	0.001	7.41E-08	-0.013	0.002	1.20E-09	0.001	0.004	7.11E-01
rs6598351	FAM169B*	t/c	-0.02	0.03	5.26E-01	0.006	0.001	2.80E-05	0.012	0.003	1.77E-05	-0.004	0.005	3.90E-01
rs11646917	SALL1	t/g	-0.01	0.03	6.65E-01	-0.009	0.001	4.83E-10	-0.015	0.003	4.76E-09	-0.015	0.005	1.30E-03
rs11867840	BCAS3	g/a	0.04	0.03	1.04E-01	-0.006	0.001	4.86E-06	-0.018	0.002	2.35E-13	0.011	0.004	1.00E-02
rs6054375	BMP2	t/g	0.03	0.03	2.45E-01	-0.010	0.001	6.92E-15	-0.018	0.002	1.83E-15	-0.003	0.004	4.74E-01
rs3791679	EFEMP1/PNPT	g/a	0.04	0.03	1.72E-01	-0.005	0.001	1.17E-04	-0.013	0.002	4.92E-08	0.003	0.004	5.14E-01
rs12494328	FLNB	a/g	0.04	0.03	1.52E-01	0.006	0.001	1.56E-06	0.016	0.002	6.03E-11	0.009	0.004	4.50E-02
6:36592986	CDKN1A	d/r	-0.02	0.03	5.32E-01	0.006	0.001	1.92E-05	0.015	0.003	1.12E-08	-0.006	0.005	2.09E-01
rs72852338	CDKN1A	c/a	-0.02	0.03	5.46E-01	0.006	0.001	3.29E-05	0.014	0.003	3.17E-08	-0.005	0.005	2.97E-01
rs1074407	TRIOBP	t/a	0.11	0.02	4.00E-06	0.006	0.001	3.32E-07	0.012	0.002	1.90E-08	0.008	0.004	3.92E-02

For these SNPs, the associations with the other traits are also included. Here the SNPs genome-wide significantly associated with vertical cup-disc ratio or cup area and that are Bonferroni significantly associated with VCDR or cup area are shown in bold (p-value <  $5.37 \times 10^{-4}$ ; 0.05/93).Nearest gene, reference NCBI build37; A1, reference allele; A2, other allele;  $\beta$ , effect size on effect size on the endophenotype (IOP, VCDR, cup area or disc area) based on allele A1; SE, standard error of the effect size; i, insertion; d, deletion; r, reference.\*Genome-wide associated in the European-only meta-analysis

			ЮР			VCDR			Cup area	a	Disc area			
SNP	Nearest gene	A1/A	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
rs1192414	CDC7/TGFBR	a/g	0.06	0.03	5.66E-02	0.014	0.001	1.78E-23	0.007	0.003	1.12E-02	0.087	0.005	7.44E-71
rs10753787	F5	t/c	-0.03	0.02	1.69E-01	-0.007	0.001	2.48E-09	-0.005	0.002	2.14E-02	-0.019	0.004	1.60E-06
rs2920293	PSCA	g/c	0.00	0.02	8.57E-01	-0.006	0.001	5.04E-09	-0.007	0.002	9.17E-04	-0.015	0.004	9.94E-05
rs4658101	CDC7/TGFBR	a/g	0.06	0.03	4.46E-02	0.013	0.001	5.19E-23	0.007	0.003	1.13E-02	0.089	0.005	8.01E-77
1:16953052	F5/SELP	i/r	0.02	0.03	4.22E-01	0.007	0.001	7.20E-07	0.005	0.003	5.44E-02	0.033	0.005	1.49E-12
rs2239854	F5/SELP	a/g	0.03	0.03	2.64E-01	0.006	0.001	8.37E-07	0.005	0.002	5.04E-02	0.030	0.004	7.60E-13
rs9843102	ABI3BP	a/g	0.00	0.03	9.84E-01	-0.006	0.002	2.18E-04	-0.002	0.003	5.88E-01	-0.036	0.005	1.35E-11
8:88744441	DCAF4L2	d/r	-0.01	0.02	6.98E-01	0.006	0.001	6.66E-07	0.006	0.002	4.53E-03	0.026	0.004	2.04E-11
rs6468996	DCAF4L2	t/c	0.00	0.02	9.12E-01	0.005	0.001	2.52E-07	0.006	0.002	2.14E-03	0.025	0.004	5.16E-11
rs61101201	ELP4/PAX6	g/t	0.02	0.03	5.51E-01	0.006	0.001	2.27E-06	0.005	0.002	4.51E-02	0.028	0.004	1.53E-10
rs56385951	CARD10	a/g	-0.06	0.04	9.08E-02	0.011	0.002	1.87E-11	0.008	0.003	8.83E-03	0.047	0.006	1.49E-16
1:3046430	PRDM16	i/r	-0.04	0.04	4.14E-01	0.007	0.002	5.35E-04	-0.002	0.004	7.15E-01	0.044	0.007	1.79E-09
rs12028027	PRDM16	c/t	-0.03	0.04	4.97E-01	0.007	0.002	2.15E-04	-0.001	0.004	8.58E-01	0.043	0.007	1.46E-09

Table2c. Single nucleotide polymorphisms (SNPs) that are genome-wide significantly associated with vertical cup-disc ratio or with disc area

For these SNPs, the associations with the other traits are also included. Here the SNPs genome-wide significantly associated with vertical cup-disc ratio or with disc area that are Bonferroni significantly associated with the VCDR or disc area are shown in bold (p-value <  $5.37 \times 10^{-4}$ ; 0.05/93).Nearest gene, reference NCBI build37;  $\beta$ , effect size on effect size on the endophenotype (IOP, VCDR, cup area or disc area) based on allele A1; SE, standard error of the effect size; i, insertion; d, deletion; r, reference.

				IOP			VCDR			Cup area			Disc area	
SNP	Nearest gene	A1/A2	β	SE	Р	β	SE	Р	β	SE	Р	В	SE	Р
1:227562773	CDC42BPA	d/r	-0.10	0.05	3.01E-02	0.003	0.002	2.37E-01	0.024	0.004	8.05E-09	-0.055	0.008	3.65E-13
rs73102394	CDC42BPA	t/c	-0.09	0.05	4.34E-02	0.003	0.002	1.62E-01	0.022	0.004	4.16E-08	-0.053	0.007	5.01E-13
rs11811982	CDC42BPA	a/c	-0.12	0.05	1.35E-02	0.004	0.002	5.54E-02	0.027	0.004	2.31E-10	-0.062	0.008	2.02E-15
rs10021731	UGT8	c/t	0.01	0.02	8.23E-01	-0.002	0.001	5.56E-02	-0.002	0.002	2.68E-01	-0.020	0.004	7.48E-07
rs12220165	CTNNA3*	g/c	0.02	0.03	5.88E-01	-0.004	0.002	1.47E-02	-0.004	0.003	1.92E-01	-0.023	0.005	2.51E-05
rs787541	U6, GADD45A	c/g	0.07	0.03	7.08E-03	0.002	0.001	7.47E-02	0.002	0.002	4.82E-01	0.023	0.004	6.66E-08
rs1367187	DIRC3	c/t	-0.07	0.03	9.74E-03	0.002	0.001	2.46E-01	-0.002	0.003	4.87E-01	0.026	0.005	1.03E-08
rs2443724	VGLL4	c/g	0.00	0.02	8.62E-01	-0.003	0.001	1.53E-02	0.000	0.002	9.15E-01	-0.022	0.004	4.72E-08
rs1013830	CTNNA3	t/c	0.00	0.05	9.49E-01	-0.007	0.002	4.80E-03	-0.004	0.005	4.10E-01	-0.046	0.008	5.45E-08

Table 3 Single nucleotide polymorphisms (SNPs) that are genome-wide significantly associated with optic nerve head parameters (cup area and disc area)

For these SNPs, the associations with the other traits are also included. SNPs that are Bonferroni significantly associated with other traits are shown in bold (p-value <  $5.37 \times 10^{-4}$ ; 0.05/93). In the first rows, the SNPs genome-wide significantly associated with cup area are shown. Next, SNPs associated with only disc area, are shown. Nearest gene, reference NCBI build37; A1, reference allele; A2, other allele;  $\beta$ , effect size on the endophenotype (IOP, VCDR, cup area or disc area) based on allele A1; SE, standard error of the effect size; i, insertion; d, deletion; r, reference. \*Genome-wide associated in the European-only meta-analysis.

## Table 4. Association with primary open-angle glaucoma in a meta-analysis of four independent glaucoma case-

control studies (ANZRAG, NEIGHBORHOOD, Singapore, and Southampton).

	Nearest gene	Δ1/Δ2	OR (95% CI)	OR	P-value	P-value	Direction	12	P-value of
	incurest gene	//1//		(R)	· value	(R)	Direction		heterogeneity
				(14)		(1)			neterogeneity
IOP SNPs				1			1		1 005 01
rs10918274	TMC01	t/c	1.39 (1.30-1.50)	1.39	2.75E-19	1.37E-09	++++	38.4	1.82E-01
rs/635832	FNDC3B	g/t	0.89 (0.83-0.95)	0.91	1.41E-03	3.65E-02	?	33.9	2.20E-01
rs10281637	CAV1/CAV2	c/t	1.13 (1.07-1.20)	1.13	2.32E-05	2.32E-05	++++	0	4.89E-01
rs2487048	ABCA1	a/g	1.26 (1.19-1.33)	1.26	2.65E-15	3.82E-03	++++	82.9	5.53E-04
rs81/6/41	ABU	a/g	1.07 (0.99-1.17)	1.04	7.36E-02	5.25E-01	-+-+	58.5	6.51E-02
15/944/35	Many genes	c/g	1.06 (1.01-1.13)	1.07	2.99E-02	2.99E-02	++++	0	8.99E-01
11.120257425	(NUP160, PTPRJ)	d/r	1 16 (1 00 1 22)	1 10	1 525 06	2.025.02		02.2	
11:120357425	ARHGEF12	u/r +/c	1.10 (1.09-1.23)	1.19	1.52E-06	3.02E-02	++++	83.Z	4.05E-04
rc0012011	ADAIVITS8		1.07 (0.94-1.24)	1.17	2.72E-01	4.46E-01	+:	78.0	9.35E-03
135513511	0437	g/a	0.80 (0.70-0.84)	0.80	1.081-17	1.081-17		0	7.502-01
rs1925953	RPE65	t/a	1 07 (1 02-1 13)	1 10	4 21F-03	2 01F-02	++++	46.7	1 31F-01
rs1192414	CDC7/TGFBR3	a/g	1.08 (1.02-1.15)	1.10	9 26F-03	9 26F-03	++++	0	7 27F-01
rs10753787	ES F5	t/c	0.97 (0.93-1.03)	0.97	3.67F-01	3.67F-01		0	9.92F-01
rs6804624	COL8A1	c/t	0.99 (0.94-1.05)	0.99	8.14E-01	8.14E-01	+	0	8.42E-01
rs72759609	PDZD2	c/t	0.90 (0.83-0.99)	0.91	3.20E-02	3.20E-02		0	9.53E-01
rs114503346	DUSP1	t/c	1.00 (0.80-1.25)	1.00	9.99E-01	8.80E-01	+?-+	42	1.78E-01
rs4960295	RREB1	a/g	0.99 (0.95-1.05)	1.00	9.50E-01	9.09E-01	-+-+	4.6	3.70E-01
rs10274998	DGKB	t/c	1.03 (0.98-1.10)	1.04	2.16E-01	2.16E-01	++-+	0	5.38E-01
rs2157719	CDKN2B-AS1	c/t	0.69 (0.66-0.74)	0.69	1.29E-40	1.29E-40		0	5.67E-01
rs1900005	ATOH7	a/c	1.01 (0.96-1.07)	1.01	6.98E-01	6.77E-01	+-++	5.1	3.67E-01
10:96008348	PLCE1	d/r	1.02 (0.97-1.09)	1.04	3.38E-01	3.15E-01	+-+?	35.3	2.13E-01
rs1346	SSSCA1	t/a	0.90 (0.85-0.97)	0.91	2.41E-03	2.41E-03		0	9.04E-01
rs4936099	ADAMTS8	c/a	0.94 (0.9-1.00)	0.94	5.75E-02	5.75E-02		0	9.63E-01
rs324780	TMTC2	g/a	0.93 (0.89-0.99)	0.93	1.35E-02	1.35E-02		0	7.69E-01
13:36629905	DCLK1	d/r	0.99 (0.94-1.05)	0.99	7.53E-01	8.00E-01	+-	6.2	3.62E-01
rs8015152	SIX6	t/c	1.21 (1.16-1.28)	1.19	3.90E-15	7.08E-05	++++	62.4	4.62E-02
rs4299136	ASB7	c/g	1.03 (0.97-1.10)	1.03	3.55E-01	3.55E-01	++-+	0	8.29E-01
16:51461915	SALL1	i/r	0.94 (0.89-1.00)	0.94	3.85E-02	3.85E-02		0	7.82E-01
rs6107845	BMP2	a/g	0.89 (0.85-0.94)	0.91	1.02E-05	6.94E-03		43.1	1.53E-01
rs5752773	CHEK2	g/c	0.92 (0.88-0.98)	0.92	4.63E-03	4.63E-03		0	9.12E-01
rs2092172	CARD10	a/g	0.97 (0.92-1.04)	0.98	4.35E-01	4.35E-01	+-	0	7.76E-01
rs6764184	FLNB	t/g	1.07 (1.02-1.13)	1.02	5.73E-03	7.66E-01	++-+	86.1	8.14E-05
rs7717697	VCAN	c/t	0.98 (0.93-1.04)	0.98	5.26E-01	5.26E-01	?	0	7.30E-01
rs2920293	PSCA	g/c	1.03 (0.98-1.09)	1.03	2.25E-01	2.25E-01	++-?	0	3.79E-01
rs1681/39	ENO4	t/c	1.02 (0.97-1.08)	1.03	3.92E-01	3.99E-01	++	49.2	1.16E-01
rs/311936	FAM101A	c/g	0.99 (0.95-1.04)	1.00	8.12E-01	8.59E-01	+	11	3.38E-01
14:23388793	KBIVI23	r/d	1.03 (0.98-1.10)	1.03	1.83E-01	1.83E-01	+++?	0	4.61E-01
Cup area SNDs									
reases	DEDE	+/c	1 11 (1 06 1 10)	1 1 1		2 975 02		E0.2	6 10E 02
rs4946112			1.11(1.00-1.10)	1.11	5.70E-05	2.87E-02	++-+	0	5.10E-02
1.227562772	CDCA2RDA	d/g d/r	0.93 (0.91-1.01)	0.90	1.18L-01	2 11E-01	2	18.6	1.43E-01
rs12016882	TRIR?		1.08(1.03-1.14)	1.08	1.14L-02	4 25E-03	2	40.0	8.63E-01
rs2508/282		c/s	1.08 (1.05-1.14)	1.00	6 72E-01	6 72E-01	+2-+	0	3 01E-01
rs117598310	CRISPI D1	t/σ	1.04 (0.85-1.29)	1.05	5 39F-02	5 39F-02	+++	0	8 01F-01
rs1360589	CDKN2B-AS1	0/5 c/t	0.69 (0.66-0.73)	0.69	1 90F-42	1 90F-42		0	6.47F-01
rs10998036	ATOH7	c/g	1.01 (0.96-1.08)	1.02	5.42F-01	5.72F-01	+	26	2.55E-01
10.96008348	PLCE1	d/r	1.02 (0.97-1.09)	1.02	3 38F-01	3 15E-01	+-+?	25 3	2.33E 01
rs1346	SSSCA1	t/a	0.90 (0.85-0.97)	0.91	2.41F-03	2.41F-03		0	9.04F-01
rs482507	TMTC2	c/t	0.94 (0.89-0.99)	0.94	2.03F-02	2.03F-02		0	7.46E-01
rs11613189	FAM101A	t/c	0.99 (0.95-1.05)	0.99	8.25E-01	7.77E-01	++	18.5	2.98E-01
rs7323428	DCLK1	t/g	0.99 (0.94-1.05)	1.00	7.83E-01	8.87E-01	+-+-	13.6	3.25E-01
rs2251069	DDHD1	c/t	0.95 (0.91-1.00)	0.96	7.62E-02	7.62E-02	+-	0	4.08E-01
rs4436712	SIX6	t/g	1.24 (1.19-1.31)	1.23	5.77E-18	1.52E-07	++++	48.8	1.19E-01
					. = = = 9				–

	Nearest gene	A1/A2	OR (95% CI)	OR	P-value	P-value	Direction	12	P-value of
				(R)		(R)			heterogeneity
			•						
Cup area SNPs									
rs6598351	FAM169B	t/c	0.99 (0.93-1.06)	0.99	8.06E-01	8.06E-01	-+	0	7.11E-01
rs11646917	SALL1	t/3g	0.98 (0.93-1.04)	0.98	5.49E-01	5.49E-01	++	0	5.97E-01
rs11867840	BCAS3	g/a	1.06 (1.01-1.13)	1.06	1.83E-02	2.12E-02	++++	8.3	3.51E-01
rs6054375	BMP2	t/g	0.89 (0.85-0.94)	0.91	8.52E-06	9.93E-03		47.1	1.29E-01
rs738722	CHEK2	t/c	0.93 (0.89-0.99)	0.93	1.26E-02	1.26E-02		0	9.05E-01
rs3791679	EFEMP1/PNPT1	a/g	0.96 (0.92-1.02)	0.96	2.23E-01	2.23E-01		0	5.51E-01
rs12494328	FLNB	a/g	1.13 (1.07-1.20)	1.13	1.28E-05	5.89E-04	++-+	26.9	2.50E-01
rs6804624	COL8A1	c/t	0.99 (0.94-1.05)	0.99	8.14E-01	8.14E-01	+	0	8.42E-01
6:36592986	CDKN1A	d/r	1.14 (1.09-1.21)	1.15	7.74E-07	1.04E-04	++++	36.6	1.93E-01
rs2684249	HSF2	c/t	0.92 (0.88-0.97)	0.94	1.08E-03	1.66E-01	+	63.3	4.25E-02
rs8176672	ABO	t/c	1.00 (0.91-1.11)	1.00	9.49E-01	9.49E-01	-+-?	0	3.69E-01
rs4936099	ADAMTS8	c/a	0.94 (0.90-1.00)	0.94	5.75E-02	5.75E-02		0	9.63E-01
rs34222435	ASB7	t/c	1.03 (0.97-1.10)	1.03	3.66E-01	3.66E-01	++-+	0	8.74E-01
rs1074407	TRIOBP	t/a	1.04 (1.00-1.10)	1.04	4.92E-02	8.66E-02	++++	32.9	2.15E-01
Disc area SNPs									
rs4658101	CDC7/TGFBR3	a/g	1.08 (1.02-1.16)	1.08	7.81E-03	7.81E-03	++++	0	7.22E-01
1:169530520	F5/SELP	i/r	1.01 (0.96-1.08)	1.02	5.40E-01	5.40E-01	++-?	0	7.14E-01
rs11811982	CDC42BPA	a/c	0.87 (0.80-0.97)	0.90	1.19E-02	8.28E-02	++	20.5	2.87E-01
rs9843102	ABI3BP	a/g	0.92 (0.86-0.98)	0.92	1.37E-02	1.37E-02		0	6.24E-01
rs10021731	UGT8	c/t	1.01 (0.96-1.06)	1.01	6.82E-01	6.82E-01	++	0	6.50E-01
8:88744441	DCAF4L2	d/r	1.03 (0.99-1.09)	1.04	0.1225	1.39E-01	++-+	4.9	3.68E-01
rs12220165	CTNNA3	g/c	1.08 (1.01-1.16)	1.09	1.14E-02	1.14E-02	++++	0	9.04E-01
rs7916410	ATOH7	t/c	1.00 (0.96-1.06)	1.00	7.63E-01	7.45E-01	+-++	3.9	3.73E-01
rs61101201	ELP4/PAX6	g/t	1.00 (0.94-1.06)	1.00	9.77E-01	9.77E-01	-+-?	0	9.63E-01
rs442376	TMTC2	c/t	1.04 (0.99-1.10)	1.05	7.94E-02	7.94E-02	-+++	0	6.82E-01
rs1345467	SALL1	g/a	1.07 (1.01-1.14)	1.07	1.86E-02	1.86E-02	++++	0	8.73E-01
rs5762752	CHEK2	c/g	0.92 (0.88-0.98)	0.92	4.90E-03	4.90E-03		0	8.29E-01
rs56385951	CARD10	a/g	0.99 (0.92-1.07)	1.00	9.15E-01	9.15E-01	+-+-	0	9.88E-01
1:3046430	PRDM16	i/r	0.97 (0.87-1.10)	0.98	7.13E-01	8.72E-01	+?	63.9	6.28E-02
rs787541	U6, GADD45A	c/g	0.98 (0.94-1.04)	0.98	6.10E-01	9.06E-01	++	50.7	1.08E-01
rs1367187	DIRC3	c/t	0.95 (0.90-1.01)	0.96	1.11E-01	4.12E-01	+-+-	46.1	1.35E-01
rs2443724	VGLL4	c/g	0.91 (0.87-0.97)	0.91	1.04E-03	2.61E-02	+-	38	1.84E-01
rs11129176	RARB	a/g	0.99 (0.94-1.05)	1.00	8.85E-01	9.93E-01	+	40.4	1.69E-01
rs1997404	COL8A1	g/t	1.00 (0.95-1.06)	1.00	9.60E-01	9.60E-01	-+++	0	6.18E-01
rs34935520	SIX6	g/a	1.26 (1.20-1.33)	1.26	2.82E-20	6.73E-14	++++	21.5	2.81E-01
rs60779155	ASB7	a/g	1.02 (0.96-1.10)	1.03	4.52E-01	4.52E-01	++	0	5.02E-01

Results are shown for the most significantly associated single nucleotide polymorphisms from the endophenotype analyses. Nearest gene, reference NCBI build37; A1, reference allele; A2, other allele; OR, estimated odds ratio for allele A1; 95% CI, confidence interval; OR (R), estimated odds ratio for allele A1 in random effect meta-analysis; P-value (R), p-value in random effect meta-analysis; I<sup>2</sup> statistic measuring heterogeneity on a scale of 0% to 100%; i, insertion; d, deletion; r, reference.

## Abbreviations

Abbreviation	Explanation
A1	Reference allele
A2	Other allele
Chr	Chromosome
CI	confidence interval
d	Deletion
FDR	False Discovery Rate
GWAS	Genome-wide association studies
h <sup>2</sup>	Heritability
i	Insertion
<sup>2</sup>	Statistic measuring heterogeneity on a scale of 0% to 100%
IGGC	International Glaucoma Genetics Consortium
IOP	Intraocular pressure
LCA2	Leber Congenital Amaurosis type 2
LD	Linkage disequilibrium
MAF	Minor allele frequency
OR	Estimated odds ratio for allele A1
OR (R)	Estimated odds ratio for allele A1 in random effect meta-analysis
POAG	Primary open-angle glaucoma
r	Reference
RP	Retinitis Pigmentosa
SE	Standard error
SNPs	Single nucleotide polymorphisms
VCDR	Vertical cup-disc ratio
β	Effect size