

THE UNIVERSITY of EDINBURGH

Edinburgh Research Explorer

The braincase and inner ear of 'Metriorhynchus' cf. 'M.' brachyrhynchus – implications for aquatic sensory adaptations in crocodylomorphs

Citation for published version:

Schwab, J, Young, M, Herrera, Y, Witmer, LM, Walsh, S, Katsamenis, OL & Brusatte, S 2021, 'The braincase and inner ear of 'Metriorhynchus' cf. 'M.' brachyrhynchus – implications for aquatic sensory adaptations in crocodylomorphs', *Journal of Vertebrate Paleontology*, vol. 41, no. 1, e1912062 . https://doi.org/10.1080/02724634.2021.1912062

Digital Object Identifier (DOI):

10.1080/02724634.2021.1912062

Link:

Link to publication record in Edinburgh Research Explorer

Document Version: Peer reviewed version

Published In: Journal of Vertebrate Paleontology

Publisher Rights Statement:

© by the Society of Vertebrate Paleontology

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Édinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



1	The braincase and inner ear of 'Metriorhynchus' cf. 'M.' brachyrhynchus - implications for
2	aquatic sensory adaptations in crocodylomorphs
3	
4	JULIA A. SCHWAB, ^{1,*} MARK T. YOUNG, ¹ YANINA HERRERA, ² LAWRENCE M.
5	WITMER, ³ STIG A. WALSH, ⁴ ORESTIS L. KATSAMENIS, ⁵ and STEPHEN L.
6	BRUSATTE ¹
7	
8	¹ School of GeoSciences, Grant Institute, James Hutton Road, The King's Buildings,
9	University of Edinburgh, Edinburgh, EH9 3FE, United Kingdom, julia.schwab@ed.ac.uk;
10	² CONICET. División Paleontología Vertebrados, Museo de La Plata, FCNyM, UNLP, La
11	Plata, Argentina;
12	³ Department of Biomedical Sciences, Heritage College of Osteopathic Medicine, Ohio
13	University, Athens, Ohio 45701, USA;
14	⁴ National Museum of Scotland, Chambers Street, Edinburgh, EH 1 1JF, United Kingdom;
15	⁵ µ-VIS X-ray Imaging Centre, Faculty of Engineering and Physical Sciences, University of
16	Southampton, SO17 1BJ, Southampton, United Kingdom
17	
18	Running head instruction:
19	Schwab et al. — Braincase and inner ear of 'Metriorhynchus' cf. 'M.' brachyrhynchus
20	
21	*Corresponding author
22	
23	
24	
25	

ABSTRACT-During their long evolutionary history crocodylomorphs achieved a great 26 diversity of body sizes, ecomorphotypes and inferred feeding ecologies. One unique group of 27 28 crocodylomorphs are the thalattosuchians, which lived during the Jurassic and Cretaceous (ca. 191 – 125 Ma). They transitioned from shallow marine species, like teleosauroids into 29 30 fully pelagic forms with paddle shaped limbs and a vertically orientated tail fluke, the 31 metriorhynchids. The osteological adaptations that allowed metriorhynchids to live in the 32 water are generally well understood, but less is known about their neurosensory and endocranial systems, such as the brain, inner ears, sinuses and cranial nerves and how they 33 34 were related to their aquatic lifestyle. Based on micro-computed tomography (µCT) data and three-dimensional models we here describe the braincase and endocranial anatomy of a fully 35 marine metriorhynchid, 'Metriorhynchus' cf. 'M.' brachyrhynchus (NHMUK PV OR 32617). 36 We found several neuroanatomical features that likely helped this species function in its 37 marine environment. This includes a unique flexure in the brain endocast not seen in other 38 39 thalattosuchians. Other features that have previously been seen in thalattosuchians include enlarged cerebral hemispheres; a hypertrophied venous sinus system; enlarged internal 40 carotid arteries and foramina; and closed/absent lateral pharyngotympanic foramina. The 41 42 specimen also possesses a pelagic metriorhynchid bony labyrinth morphology, with a compact and dorsoventrally short shape, thick semicircular canals, an enlarged vestibule and 43 44 potentially a short cochlear duct. A review of character distribution confirms that some of these features evolved at the base of Thalattosuchia in semiaquatic species, long before 45 metriorhynchids became pelagic, suggesting that endocranial anatomy helped allow 46 47 metriorhynchoids colonize the ocean realm.

48

49

INTRODUCTION

50

Mesozoic oceans were inhabited by various marine reptiles such as ichthyosaurs, mosasaurs, sauropterygians, and sea turtles. These groups achieved incredible morphofunctional diversity, often being at the top of their respective food chains. To successfully adapt to the marine environments, these reptile groups needed to fundamentally change their anatomy and physiology relative to their terrestrial ancestors, evolving a body plan well suited for life in the open ocean (e.g. Massare, 1988; Motani, 2009; Kelley and Pyenson, 2015; Foffa et al., 2018).

One of these marine reptile groups was Metriorhynchidae, a group of thalattosuchian 58 59 crocodylomorphs that included the only known obligately aquatic archosaurs (Herrera, 2015; Herrera et al., 2017; Fig. 1). Metriorhynchidae evolved during the Middle Jurassic, and 60 diversified throughout the Jurassic into highly successful marine predators, becoming extinct 61 during the Early Cretaceous (Fraas, 1902; Andrews, 1913; Lepage et al., 2008; Pol and 62 Gasparini, 2009; Young et al., 2010; Larsson et al., 2011; Chiarenza et al., 2015; Herrera and 63 Vennari, 2015; Fernández et al., 2019; Sachs et al., 2020). As they transitioned from 64 semiaquatic basal metriorhynchoids into open-ocean swimming metriorhynchids, their body 65 plan underwent a radical change, evolving paddle-shaped limbs, a hypocercal tail fin, and 66 hypertrophied nasal salt glands, while losing their osteoderms (Fraas, 1902; Andrews, 1913; 67 Fernández and Gasparini, 2008; Young et al., 2010; Wilberg, 2015; Ősi et al., 2018). 68 Osteological modifications in their pelvis suggest that metriorhynchids gave birth to live 69 70 young (Herrera et al., 2017).

Besides their osteological changes, thalattosuchian sensory systems played an important role in their evolutionary transition, and these systems have only recently become a focus of research. Computed tomography (CT) scans and three-dimensional visualization of internal cranial anatomy has helped us better understand the land-to-sea transition in Thalattosuchia (Brusatte et al., 2016; Pierce et al., 2017; Herrera et al., 2018; Schwab et al.,

2020). Neuroanatomical and internal cranial features, such as the brain, sinuses, and sensory
systems like the inner ear, provide unique insight into how thalattosuchians—and in
particular metriorhynchids—adapted to their aquatic environment.

79 Here we describe the braincase and endocranial anatomy of the Middle Jurassic metriorhynchid 'Metriorhynchus' cf. 'M.' brachyrhynchus from the Vaches Noires cliffs of 80 Normandy (France), based on an isolated but well preserved and, rare for thalattosuchian 81 82 fossils, uncrushed specimen. CT scanning allows analysis of the brain endocast, inner ear, vasculature, and pneumatic sinuses of this metriorhynchid, which we compare with other 83 84 extant and extinct crocodylomorphs. This allows us to better understand the morphological and physiological features that allowed these unique crocodylomorphs to adapt to their ocean 85 realm. 86

Institutional Abbreviations—FMNH, Field Museum of Natural History, Chicago,
Illinois, USA; IVPP, Institute of Vertebrate Paleontology and Paleoanthropology of the
Chinese Academy of Sciences, Beijing, China; MLP, Museo de La Plata, La Plata,
Argentina; NHMUK, Natural History Museum, London, England, United Kingdom.

93

92

MATERIALS AND METHODS

94 Fossil Specimen

The braincase of the basal geosaurine thalattosuchian '*Metriorhynchus*' cf. '*M*.' *brachyrhynchus* (NHMUK PV OR 32617; Fig. 2) was purchased in 1857 by the British Museum, and now forms part of the Tesson Collection in the Natural History Museum London. The specimen was discovered from the Vaches Noires cliffs of Normandy, France, where late Callovian/early Oxfordian formations are exposed (Brignon, 2016). Although

100 fragmentary, the braincase is relatively well preserved with no evidence of shearing or101 dorsoventral compression.

While Lydekker (1888:96) initially referred NHMUK PV OR 32617 to Metriorhynchus 102 103 superciliosus (now referred to the genus Thalattosuchus; Young et al., 2020), we refer it to 'M.' cf. 'M.' brachyrhynchus. This is due to (1) the strongly concave supraoccipital that is 104 noticeably inset relative to the occipital surface of the cranium, (2) the pronounced nuchal 105 106 crest along the midline of the supraoccipital, (3) the basicccipital tuberosities not being proportionally enlarged, (4) and the occipital condyle being as wide mediolaterally as it is 107 108 high dorsoventrally. These characters match those seen in the English 'M.' brachyrhynchus specimens (NHMUK PV R 3699, NHMUK PV R 3700, NHMUK PV R 3804; Andrews, 109 1913). The distinctly concave supraoccipital is a readily identifiable character, and it is not 110 seen in the Vaches Noires specimens of Thalattosuchus superciliosus or Suchodus 111 durobrivensis (see Lepage et al., 2008). Nor is this feature seen in Maledictosuchus riclaensis 112 113 (Parrilla-Bel et al., 2013), Dakosaurus andiniensis (Pol and Gasparini, 2009), D. cf. andiniensis (Herrera and Vennari 2015), Plesiosuchus manselii (Young et al., 2012), or 114 Torvoneustes coryphaeus (Young et al., 2013). Purranisaurus potens also has a concave 115 supraoccipital (Herrera et al., 2015). However, there the concavity is 'open' laterally, being 116 contiguous with the concave dorsal-half of the otoccipital. 117

118

119 Computed Tomography and Visualisation

The braincase of '*Metriorhynchus*' cf. '*M.*' *brachyrhynchus* (NHMUK PV OR 32617)
was imaged by means of microfocus Computed Tomography (μCT) at the μ-VIS X-ray
Imaging Centre at the University of Southampton, using the custom designed Nikon/Metris
dual source high energy microfocus walk-in enclosure system. A 450 kVp source was used,
coupled with a 1621 PerkinElmer caesium-iodide detector. Peak voltage was set at 430 kVp

and the current at $120 \,\mu$ A. A total of 3142 projections (4 frames per projection) were collected during a 360° rotation, with each projection occurring over an exposure time of 250 ms.

128 Source to detector and source to object distances were set to 684.0 mm and 186.8 mm respectively, resulting in a radiograph pixel-size of 54.6 µm and a field of view of c. 90 mm x 129 90 mm. Imaging of the whole specimen (W \approx 65 mm, D \approx 85 mm, H \approx 128 mm) was 130 131 achieved in two halves as shown in (Fig. S1) allowing for an overlap of approximately 20%. The raw projection data for each scan were reconstructed into 3D volumes (32-bit raw) using 132 133 an isotropic voxel-size (with an edge length) of 54.6 µm by means of Nikon's reconstruction software (CT Pro 3D, v. XT 2.2 SP10), which uses a filtered back projection algorithm. 134 The two raw reconstructed volumes were then concatenated using Fiji/ImageJ 135 (Schindelin et al., 2012) matching one of the overlapped slices of bottom and top volumes, to 136 generate a single volume that contained the whole specimen which assured anatomy 137 continuity (Supplementary Video SV1). 138

To allow for further analysis and segmentation the CT volumes were contrast-139 enhanced using a custom-made algorithm, which was developed and optimised for this 140 specimen. The script was written in ImageJ macro language (IJ1 Macro), to generate and 141 apply a contrast and edge enhancement filter on a slice-by-slice basis. The macro 1. 142 duplicated the scan volume and run a bandpass filter (parameters: filter large =15; 143 filter small=5; source: https://imagej.nih.gov/ij/docs/menus/process.html#fft) on it. It then 144 ran a 3D median filter (2 x 2 x 2 pixel) to improve on signal to noise ratio, followed by an 145 unsharp mask (blur radius = 1 pixel; filter weighting = 0.7). The resulted "filter" volume was 146 then multiplied with the original in 32-bit domain to generate the enhanced "filtered" volume. 147 which was then sampled down to 16-bit to reduce size while maintaining a wide dynamic 148 range (Fig. S2). 149

150	The filtered volumes were then further analysed using the software Materialise Mimics
151	20.0. The lasso tool was used for manual image segmentation of anatomical structures. For
152	structures with minimal shape changes we interpolated between slides. Three dimensional
153	models of the brain cavity, endosseous labyrinth, pneumatic sinuses, cranial nerves, veins,
154	and arteries were created based on those segmentations.
155	Three dimensional models have been uploaded to Morphosource
156	(https://www.morphosource.org/) and can be accessed here:
157	https://www.morphosource.org/Detail/ProjectDetail/Show/project_id/1209.
158	
159	RESULTS
160	
161	External Cranial Anatomy
162	In our description of the braincase of 'Metriorhynchus' cf. 'M.' brachyrhynchus
163	(NHMUK PV OR 32617) we do not include the quadrate and squamosal, as they are largely
164	incomplete and not well preserved (Fig. 2, 3).
165	Frontal-The frontal is largely incomplete, with only the frontal participation to the
166	intertemporal bar, supratemporal fossae and part of the lateral processes present (Fig. 2A,
167	3A). The left and right elements are fused, with no evidence internally or externally for a
168	midline suture. The frontal contacts the parietal along its posterior margin and the
169	laterosphenoids ventrolaterally. The frontal forms the majority of the anteromedially
170	expanded supratemporal fossae, as in other metriorhynchids (e.g. Andrews, 1913; Young et
171	al., 2013). The preserved part of the frontal is higher on the lateral edge than in the medial
172	part (Fig. 2A, 3A).
470	Derivated The manifest is subting the small measure does does not be a first of the state of t

Parietal—The parietal is relatively well preserved and is a single fused element with
no evidence of an interparietal suture internally or externally. It forms part of the dorsal and

posterolateral wall of the braincase, also contributing to the posterior and medial margins of 175 the supratemporal fenestrae. It is a single Y-shaped element, with the anterior process 176 177 forming the posterior half of the intertemporal bar and the two lateral processes diverge posteriorly, where the supraoccipital fits between them (Fig. 2A, 3A). The parietal bears a 178 sagittal crest, as in most other thalattosuchians (e.g. Andrews, 1913; Young et al., 2013; 179 180 Brusatte et al., 2016; Herrera et al., 2018). The parietal contacts the frontal along its anterior margin, and within the supratemporal fenestrae along its ventral margin, it contacts 181 laterosphenoid anteroventrally and the prootic posteroventrally (Fig. 2B, 3B). In occipital 182 183 view, the parietal contacts the otoccipital and the supraoccipital ventrally (Fig. 2C, 3F). **Prootics**—The prootics are located on the lateral wall of the braincase and form the 184 posteromedial region of the supratemporal fenestra, as in other thalattosuchians (e.g. 185 Andrews, 1913; Young et al., 2013; Herrera et al., 2018; Fig. 2B, E, 3B, E). Each prootic 186 dorsally contacts the parietal, posteroventrally the quadrate, and anteriorly the laterosphenoid. 187 This subtriangular bone forms part of the dorsal margin of the trigeminal fossa and is broadly 188 visible in dorsal view (Fig. 2A, 3A). The prootic and otoccipital house the cavity of the inner 189 ear internally. 190

Laterosphenoids—The laterosphenoids form the lateral walls of the braincase (Fig. 191 2B, E, 3B, E). Internally, they house the cerebral hemispheres. The laterosphenoid forms the 192 anteromedial margin of the supratemporal fenestra. Ventrally, each laterosphenoid contacts 193 194 the basisphenoid, anterodorsally the frontal, posterodorsally the parietal, and posteriorly the prootic. The laterosphenoid forms the anterior and anteroventral margins of the trigeminal 195 196 foramen (Fig. 2B, 3B). The trigeminal foramen is bilobate, or hour-glass-shaped, as is the case in metriorhynchids generally (Herrera et al., 2018), differing from the more commonly 197 found circular foramen of other crocodylomorphs. Of the two lobes, the posterodorsal one is 198 smaller than the anteroventral one. 199

200 Supraoccipital—The supraoccipital forms the dorsal part of the occipital region of the braincase in posterior view, where it is a singular median bone dorsal to the foramen magnum 201 but does not participate in its margins (Fig. 2C, 3F). It is lateromedially broader than 202 dorsoventrally tall and exhibits two dorsoventrally long grooves, separated by a pronounced 203 nuchal rest running along the midline of the bone. This has been noted previously in the 204 teleosauroid Plagiophthalmosuchus cf. gracilirostris ('Steneosaurus' cf. gracilirostris in 205 206 Brusatte et al., 2016), but in Cricosaurus araucanensis the supraoccipital is flat in this region (Herrera et al., 2018). Dorsally the supraoccipital contacts the parietal and ventrolaterally 207 208 articulates with the otoccipital, with the latter bones excluding the supraoccipital from the foramen magnum. Participation of the supraoccipital in the dorsal margin of the foramen 209 magnum is variable in Thalattosuchia, and most thalattosuchians have a midline otoccipital 210 contact dorsal to the foramen magnum that excludes the supraoccipital. However, Brusatte et 211 al., (2016) suggested that the supraoccipital overlays the otoccipital and this could explain 212 why in some specimens the supraoccipital forms the dorsomedial margin of the foramen 213 magnum. 214

Otoccipital—The otoccipital is a single element, comprised of the fused exoccipital 215 and opisthotic on either side of the braincase. It contacts the supraoccipital dorsomedially and 216 the parietal along the dorsal margin, lateral to the supraoccipital (Fig. 2C, 3F). 217 Ventromedially it contacts the basioccipital and ventrolaterally the quadrate, but in this 218 219 specimen the quadrate is largely missing. The otoccipital forms the dorsal and lateral margins of the foramen magnum, which is oval with the major axis mediolaterally oriented. The 220 221 otoccipital contacts the basioccipital at the dorsolateral corners of the occipital condyle. The paroccipital processes are not preserved in this specimen. The foramen for the cranial nerve 222 XII (hypoglossal foramen) is located on the occipital surface of the braincase ventrolateral to 223 the foramen magnum. It appears to be a single opening as in most other thalattosuchians 224

(Jouve, 2009; Fernández et al., 2011; Brusatte et al., 2016; Herrera et al., 2018) although the
foramen for the second hypoglossal canal might not be preserved in this specimen. The
foramina for cranial nerves IX-XI are not preserved in this specimen. The carotid foramen is
located ventrolateral to the occipital condyle. It is enlarged as in other thalattosuchians (e.g.
Jouve, 2009; Brusatte et al., 2016; Pierce et al., 2017; Herrera et al., 2018).

Occipital Condyle—The occipital condyle is round and mediolaterally as wide as 230 dorsolaterally tall (Fig. 2C, 3F). It is mostly formed by the basioccipital centrally and the 231 otoccipital, which forms the left and the right dorsolateral corners. The dorsal margin forms 232 233 the ventral floor of the foramen magnum. In essence, there is a gap between the two otoccipital corners, which is filled by the basioccipital. This is the case in most other 234 thalattosuchians (Andrews, 1913; Pierce and Benton, 2006; Lepage et al., 2008; Jouve, 2009; 235 Foffa and Young, 2014). In derived geosaurine metriorhynchids such as Plesiosuchus 236 manselii (Young et al., 2012) and Torvoneustes corvphaeus (Young et al., 2013), however, 237 the otoccipital covers the entire dorsal margin of the occipital condyle, the left and right 238 otoccipitals meet to exclude the basioccipital from the dorsal margin of the condyle, and thus 239 the floor of the foramen magnum. 240

Basioccipital—The basioccipital, a midline bone, forms the ventromedial portion of occipital region of the braincase, including most of the occipital condyle. In occipital view, it contacts the otoccipital dorsolaterally (Fig. 2C, 3F), and in ventral view, it contacts the basisphenoid anteriorly (Fig. 2D, 3D). It forms the left and right basal tubera, located ventrolateral to the occipital condyle, which extend in the posteroventral direction. Medial to tuberosities is the median pharyngeal foramen (Fig. 2D, 3D).

Basisphenoid—The basisphenoid is partially preserved. In ventral view, the
basisphenoid contacts the basioccipital along its posterior margin. The basisphenoid forms
the anterior margin of the median pharyngeal foramen (Fig. 2D, 3D). Here the lateral

pharyngeal foramina (pharyngotympanic or Eustachian tube) are absent, as in *Cricosaurus araucanensis* and other metriorhynchids (e.g. *Purranisaurus potens*, *Thalattosuchus superciliosus*, supporting the hypothesis that metriorhynchids lacked these foramina, at least in morphologically mature individuals, see Herrera et al., 2018). The basisphenoid is partially visible in occipital view, ventral to basal tubera of the basioccipital (Fig. 2C, 3F). The basisphenoid continues anteriorly as a narrow structure along the midline. The basisphenoid is expected to house the orbital arteries, but the braincase is broken in this region.

257

258 Internal Cranial Anatomy

Three-dimensional models of the brain endocast, pneumatic sinuses, bony labyrinth, cranial nerves and vasculature are described in detail below (Figs. 4–7, S3).

Cranial Endocast—The cranial endocast is incomplete due to a break in the anterior 261 part. The break is located in the forebrain region and hence the olfactory tract and bulbs are 262 missing. NHMUK PV OR 32617 shows a concave dorsal curvature in lateral view in the 263 forebrain/midbrain region and it seems that the dorsal surface of the cerebellum forms a dural 264 peak, which might be part of the venous sinus (Fig. 5). Other thalattosuchian brain endocasts 265 have been generally described as elongated, straight and narrow, however Cricosaurus 266 araucanensis shows a slight curvature in its forebrain region as well (Wharton, 2000; 267 Fernández et al., 2011; Herrera, 2015; Herrera and Vennari, 2015; Brusatte et al., 2016; 268 Pierce et al., 2017; Herrera et al., 2018).Extant crocodylians, in contrast, have greater flexure 269 270 between the midbrain and hindbrain (cephalic flexure) and between the midbrain and 271 hindbrain (pontine flexure) (Colbert, 1946; Witmer et al., 2008; Dufeau and Witmer, 2015). Outside of Crocodylomorpha, cephalic and pontine flexure has also been found in 272 ornithosuchids, aetosaurs, erpetosuchids and phytosaurs (Baczko and Desojo, 2016; 273 Lautenschlager and Butler, 2016; Baczko et al., 2018; Nesbitt et al., 2018). The cerebrum is 274

enlarged laterally, symmetrical, positioned behind the olfactory tract and is more laterally 275 expanded than in other thalattosuchians (Brusatte et al., 2016; Pierce et al., 2017; Herrera et 276 al., 2018). Metriorhynchids generally have larger cerebral hemispheres than teleosauroids and 277 basal metriorhynchoids such as *Pelagosaurus typus* (Brusatte et al., 2016; Pierce et al., 2017; 278 Herrera et al., 2018). The optic lobes are positioned posterior to the cerebral region. They are 279 not discrete in modern adult crocodylians but are more defined in juveniles (Dufeau and 280 Witmer, 2015; Brusatte et al., 2016). The cerebellum starts at the level of cranial nerve V and 281 extends to the anterior semicircular canal of the bony labyrinth (Fig. 5B, E). The pituitary 282 283 fossa is ventral to the midbrain and posteroventral to the cerebrum, and the anterior part is connected to the brain cavity in the region of the cerebrum (Fig. 5B, E). In most other 284 thalattosuchians it is elongated anteroposteriorly and dorsoventrally low (Brusatte et al., 285 2016; Pierce et al., 2017). In NHMUK PV OR 32617 it appears more bulbous and rounded 286 than in other thalattosuchians. It is positioned parallel to the brain endocast, but in extant 287 crocodylians it is oriented anterodorsally, such that the internal carotid vessels are noticeably 288 ventral to where the orbital vessels would exit anteriorly. The brain endocast is ventrally 289 more bulbous than in other thalattosuchians (Fig. 5D), but this shape also characterises 290 modern crocodylians. The rostral middle cerebral vein forms a ridge at the dorsal region of 291 the brain endocast, in the midbrain region (Fig. 5A-C, D-E). It exits the braincase through 292 the trigeminal foramen, contributing to its hour-glass shape. 293

Dorsal Dural Venous Sinus—Above the hindbrain, there is a pair of large venous sinuses. They merge with the brain endocast dorsally, between the where cranial nerve V emerges and the anterior semicircular canal of the bony labyrinth (Fig. 5). The sinuses are positioned dorsal to the crus commune of the bony labyrinth, then bend ventrolaterally to exit the braincase through the temporo-orbital foramen. The parietal forms the dorsal, the prootic the ventral and the squamosal the lateral margins of the temporo-orbital foramen, which leads

into the supratemporal fenestra. Based on comparisons with extant crocodylians (Porter et al.,
2016), this canal would have housed the caudal middle cerebral vein. The hypertrophy of this
venous sinus system is present in all other digitally segmented and natural endocasts of
thalattosuchians (Fernández et al., 2011; Brusatte et al., 2016; Pierce et al., 2017; Herrera et
al., 2018), and is never seen in this hypertrophied form in modern crocodylians (Witmer et
al., 2008; Dufeau and Witmer, 2015).

Pneumatic Sinuses—The braincase of NHMUK PV OR 32617 is pneumatized, with internal air sinuses surrounding the brain endocast (Fig. 4, S3). Those sinuses can be divided into two main systems (following Dufeau and Witmer, 2015): (1) the median pharyngeal sinus system ventral to the brain endocast and (2) the pharyngotympanic sinus system lateral to the endocast.

The median pharyngeal sinus is located within the basisphenoid. It enters the braincase 311 at the posteroventral external surface at the median pharyngeal foramen, which is located 312 313 ventral to the foramen magnum, between the basioccipital and the basisphenoid (Fig. 4B, E). The median pharyngeal sinus tube then extends anteriorly and connects to the 314 pharyngotympanic sinus system via two laterally diverging canals. The median pharyngeal 315 316 sinus diverticulum is a midline structure extending anteroventrally from the basisphenoid of the median pharyngeal sinus in the anteroventral direction, as in Plagiophthalmosuchus cf. 317 gracilirostris (Brusatte et al., 2016). 318

The pharyngotympanic sinus is the more extensive system, being located lateral to the brain endocast and connected to the median pharyngeal sinus. It is less extensive compared to teleosauroids (Brusatte et al., 2016; Herrera et al., 2018) and the basal metriorhynchoid *Pelagosaurus typus* (Pierce et al., 2017). This sinus system has been divided into eight main diverticula in the modern crocodylian species *Alligator mississippiensis* (Dufeau and Witmer, 2015). The basioccipital diverticulum is absent in NHMUK PV OR 32617 but has been

reported in the teleosauroid Macrospondylus bollensis and the basal metriorhynchoid 325 Pelagosaurus typus (Pierce et al., 2017; Steneosaurus bollensis in Herrera et al., 2018). The 326 327 recessus epitubaricum is absent in the specimen described here, as in most other 328 thalattosuchians. Ventral to the brain endocast, at the base of the pharyngotympanic sinus, the pterygoid diverticulum is located anteriorly (Fig. 4B). This diverticulum seems to be absent, 329 or at least highly reduced, in most other thalattosuchians but might be present in 330 Macrospondylus bollensis (Herrera et al., 2018), however it is worth noting that due to its 331 small size this might be a segmentation error or a resolution issue. The intertympanic 332 333 diverticulum and the parietal diverticulum are absent here, as in all other thalattosuchians. It has been debated, whether the prootic diverticulum is present in thalattosuchians. It has been 334 reported in Plagiophthalmosuchus cf. gracilirostris and Pelagosaurus typus (Brusatte et al., 335 2016; Pierce et al., 2017), but suggested to be absent in thalattosuchians by Herrera et al. 336 (2018) due to its position and forming an isolated recess in modern crocodylians (Dufeau and 337 Witmer, 2015). Laterally the pharyngotympanic sinus system is incomplete due to the 338 absence of the squamosal and quadrate, and hence a potential suspensorium diverticulum is 339 not preserved, as it is in more complete specimens like those of Cricosaurus araucanensis 340 and *Pelagosaurus typus* (Pierce et al., 2017; Herrera et al., 2018). The pharyngotympanic 341 sinus is divided at its posterior end into two diverticula, the otoccipital diverticulum dorsally 342 and the pharyngotympanic (Eustachian) tubes ventrally (Fig. 4B, E). 343

Endosseous Labyrinth—The endosseous labyrinth resembles those of other metriorhynchids, as it has a dorsoventrally short and compact appearance with thick canal cross-sectional diameters, which has been associated with their pelagic lifestyle (Schwab et al., 2020; Fig. 6). This differs from the dorsoventrally taller labyrinths, with thinner canals, that are present in semiaquatic and terrestrial crocodylomorphs. The right labyrinth is completely preserved, including all three semicircular canals, the vestibule and the cochlear

duct. The left labyrinth, however, is difficult to trace in the CT scan and hence we did not 350 include it here. The endosseous labyrinth of NHMUK PV OR 32617 has an overall triangular 351 352 appearance and the canal region takes the form of an 'M' shape from the anterior to the posterior canal, as seen in most crocodylians (Brusatte et al., 2016). The anterior canal 353 extends slightly higher dorsally than the posterior one and is oval shaped, whereas the 354 posterior canal is more triangular. The lateral semicircular canal is the smallest. All three 355 semicircular canals are more or less perpendicular to each other, and the crus commune is 356 slightly bent anteriorly and short. The anterior ampulla appears to be larger than in most other 357 358 crocodylomorphs (Schwab et al., 2020). The vestibule does not dorsally expand beyond the lateral canal. No columellae or endolymphatic ducts are preserved. The cochlear duct 359 (lagena) projects straight ventrally in lateral view and is relatively short. Metriorhynchids 360 generally have shorter cochlear ducts than other crocodylomorphs (Schwab et al., 2020), but, 361 due to the fact that the end of the lagena is not fully enclosed by bone, as in most archosaur 362 clades (Evers et al., 2019), this is just the minimum cochlear duct length. 363

Cranial Nerves—Canals for cranial nerves (CN) V (trigeminal), VI (abducens), and 364 XII (hypoglossal) were recognized and reconstructed (Fig. 5). Usually they are most 365 discernible close to the brain endocast or external braincase surface. Due to the missing 366 anterior part of the braincase, CN I (olfactory), CN II (optic), and CN III (oculomotor) are 367 missing, and the CN IV (trochlear), CN VII (facial) and CN VIII (vestibulocochlear) cannot 368 be traced in the scan but could have potentially shared a canal with other cranial nerves. The 369 largest of the nerves is the CNV. It is positioned at the lateral surface of the midbrain, 370 371 posterior to cerebrum/optic lobes. Other crocodylomorphs also exhibit a large trigeminal ganglion in this position (e.g., Dufeau and Witmer, 2015; Brusatte et al., 2016). The 372 trigeminal fossa is generally very large in metriorhynchids, but due to the lack of the quadrate 373 it is difficult to interpret its exact size and shape in the specimen described here (e.g. 374

Fernández et al., 2011; Herrera et al., 2018). The canals of CN VI are positioned lateral to the 375 pituitary fossa, ventral to CNV, and pass in an anteroventral direction. CN XII is paired and 376 exits the posterior surface of the braincase, laterally to the foramen magnum. Following 377 anteriorly from its external foramen, it meets the endocast posteroventrally to the ampulla of 378 the posterior semicircular canal of the bony labyrinth. A left second hypoglossal canal is 379 380 present (with the right one not being preserved in this specimen) and can easily be seen at the posterior surface of the braincase, ventrolateral to the foramen magnum in the anterior 381 direction. 382

383 Vasculature—The internal carotid canals were completely visible and reconstructed in the scan (Fig. 5). These hypertrophied canals enter at the posterior surface of the braincase, 384 ventrolateral to the foramen magnum in occipital view. Anteriorly from their exit foramina, 385 the internal carotid canals are visible through the entire scan and contacting the 386 pharyngotympanic sinus. Having the internal carotids pass through the pharyngotympanic 387 sinus has only been reported previously for metriorhynchids in Cricosaurus araucanensis 388 previously (Herrera et al., 2018), but modern crocodylians (Dufeau and Witmer, 2015) and 389 non-metriorhynchoid thalattosuchians (Brusatte et al., 2016; Pierce et al., 2017; Herrera et al., 390 2018) also have the medial portion of these canals pass through the pharyngotympanic sinus. 391 The internal carotid canals run parallel to the brain until the cochlear duct where they turn in 392 a ventromedial direction. The canals then pass over into the upper posterior end of the 393 pituitary fossa, with the left and right canals entering the fossa through a single midline 394 foramen. However, in most other thalattosuchians (Fernández et al., 2011; Brusatte et al., 395 396 2016; Pierce et al., 2017; Herrera et al., 2018) and modern crocodylians (Dufeau and Witmer, 2015) the carotid canals enter the pituitary fossa through two separate foramina anteriorly. 397 The orbital vessels are not preserved due to the break in the braincase but, it seems that the 398 orbital canals exit the pituitary fossa through two separate foramina (Fig. 5B, D). This is the 399

400	case for other thalattosuchians, which like NHMUK PV OR 32617, have hypertrophied
401	orbital arteries, similar in dimension to the internal carotid canals (Brusatte et al., 2016;
402	Pierce et al., 2017; Herrera et al., 2018).
403	
404	DISCUSSION
405	
406	Braincase
407	One of the most notable features of the braincase of NHMUK PV OR 32617 is the
408	unique shape of the brain endocast (Fig. 5). Thalattosuchians are generally characterised by a
409	straight endocast (Herrera, 2015; Herrera and Vennari, 2015; Brusatte et al., 2016; Pierce et
410	al., 2017; Herrera et al., 2018). Most extinct and extant crocodylomorphs have a flexure in
411	their mid/hindbrain region (Colbert, 1946; Witmer et al., 2008; Kley et al., 2010; Leardi et
412	al., 2017). NHMUK PV OR 32617, however, is unique in having a pronounced brain flexure
413	in the forebrain/midbrain region and a curved dorsal margin. This flexure may be present in
414	C. araucanensis, albeit in a less pronounced manner, and therefore could be a unique
415	characteristic of metriorhynchids. Previously there have been two ideas proposed for why
416	thalattosuchians lack the brain flexures (Herrera et al., 2018). First, most thalattosuchians are
417	characterised by a long tubular snout, with a very flat transition between the skull roof and
418	the snout. Another proposed explanation is that the extended dorsal venous sinus hides the
419	midbrain/hindbrain flexure. However, this does not explain the unusual kinked brain
420	morphology of NHMUK PV OR 32617, where there is no evidence for particularly large
421	sinuses dorsal to the forebrain/midbrain region, yet it is still obviously flexed. One other
422	interesting feature of the NHMUK PV OR 32617 brain endocast is that the pituitary fossa is
423	more bulbous and round compared to other thalattosuchians (Brusatte et al., 2016; Pierce et
424	al., 2017; Herrera et al., 2018). Reptilian pituitary glands are correlated with water

metabolism (Heller, 1942, 1950) and hence an enlarged pituitary fossa in thalattosuchians might be correlated with their water regulation to prevent dehydration (Pierce et al., 2017). An alternative hypothesis is that the pituitary fossa was enlarged due to hypertrophy of the cavernous sinus, as the cavernous sinus is located within the pituitary fossa in extant crocodylians (see Porter et al., 2016). Given the hypertrophy of the transverse sinus and stapedial canals in thalattosuchians (Fernández et al., 2011; Brusatte et al., 2016; Pierce et al., 2017; Herrera et al., 2018), hypertrophy of the cavernous sinus would not be unexpected.

The cerebral hemispheres appear enlarged, as in other metriorhynchids (Herrera et al., 432 433 2018). In birds, enlarged cerebral hemispheres have been interpreted as an adaptation to process increased sensory inputs, and to coordinate increasingly complex behaviours (Rogers, 434 1999; Walsh and Milner, 2011; Balanoff et al., 2013). The enlarged cerebral hemispheres in 435 NHMUK PV OR 32617 and other metriorhynchids could be linked to the pelagic lifestyle, as 436 these animals had to deal with more complex body movements and a greater sensory input 437 than terrestrial and semiaquatic relatives, because they lived and moved in a three-438 dimensional environment. 439

It should also be noted that the brain endocast does not accurately represent the actual 440 brain morphology. The brain does not fill the entire endocranial cavity in crocodylomorphs, 441 as parts of the brain are covered by overlaying venous sinuses (Hopson, 1979). Hence the 442 brain endocast does not perfectly mirror the actual brain, which results in size and shape 443 444 differences between the endocast and brain (the latter of which is not preserved in fossils, so we can never assess the quality of the match). However, it has been demonstrated that the 445 446 difference between the actual brain and the endocast in alligators is comparable with intraspecific variation in brain shape and therefore endocasts can be used for large scale 447 studies of brain morphology in archosaurs (Watanabe et al., 2019). In fossils it is challenging 448 to study the actual neural organs as preservation allows us to study only their preserved bony 449

450 cavities. However, some parts of the endocast more adequately reflect the actual size and 451 shape of their corresponding brain structures, such as the olfactory bulbs and the cerebral 452 hemispheres, whereas the hindbrain region does not show the same correspondence (Witmer 453 et al., 2008; Watanabe et al., 2019). Thus, we are confident that the enlarged cerebral 454 hemispheres that we note in NHMUK PV OR 32617 and other metriorhynchids is a genuine 455 feature.

'Metriorhynchus' cf. 'M.' brachyrhynchus has the thalattosuchian synapomorphy of 456 enlarged internal carotid canals and foramina (Pol and Gasparini, 2009; Brusatte et al., 2016; 457 458 Pierce et al., 2017; Herrera et al., 2018). It was suggested for metriorhynchids that the enlargement of the internal carotid canals and foramina indicates an increase in blood flow to 459 salt glands (Herrera et al., 2013). Subsequently, based on CT scans of the teleosauroid 460 Plagiophthalmosuchus cf. gracilirostris, Brusatte et al. (2016) hypothesised that the enlarged 461 cerebral carotid arteries also supplied large nasal salt glands in this taxon. Thalattosuchians 462 also evolved a hypertrophied venous sinus system. Based on the blood flow patterns of extant 463 species (Porter et al., 2016), thalattosuchians would have had far greater blood flow entering 464 and exiting the orbital and nasal regions. This corresponds to both their proportionally large 465 orbits and salt glands, and hints that the salt glands evolved at the base of Thalattosuchia 466 (Brusatte et al., 2016). It has been hypothesized that this enhanced blood supply was related 467 to thermoregulation, as larger arteries can carry a larger amount of blood, which could then 468 be used for heat exchange (Herrera et al., 2018; Porter et al., 2019). This could be linked to 469 the hypothesis that metriorhynchids possibly evolved a non-homeothermic form of 470 471 endothermy as an adaptation to active pelagic hunting (Séon et al., 2020).

472 Another interesting feature is that the lateral pharyngeal foramina are closed/absent in 473 this specimen, as in most metriorhynchids (*Cricosaurus araucanensis*; the metriorhynchid 474 specimen from Mörnsheim Formation (BSPG 1973 I195); *Purranisaurus potens*;

Thalattosuchus superciliosus; 'Metriorhynchus' westermanni; see Herrera et al., 2018), while 475 non-metriorhynchid thalattosuchians retain the plesiomorphic condition of having these 476 477 foramina. The closure of these foramina would have limited the communication between the middle ear cavities and the pharynx to just the median pharyngeal foramen (Herrera et al., 478 2018). Loss of the lateral pharyngeal (Eustachian) foramina is significant in that the 479 480 pharyngotympanic (Eustachian) tube is a highly conserved attribute that arises embryonically from the first pharyngeal pouch. In other tetrapods, this communication is used to equalize 481 pressure of the middle ear and the surrounding environment, and hence losing the lateral 482 483 plesiomorphic connection, leaving only the novel median connection, could be a physiological adaptation to the pelagic lifestyle. 484

485

486 Vestibular System

The vestibular system of the inner ear is the sensory system involved in balance and 487 equilibrium (Sipla and Spoor, 2008). It is furthermore involved in important functions 488 relating to environment such as head and gaze stabilization during body movements. For 489 secondarily aquatic animals, such as metriorhynchids, the environment puts different 490 demands on the vestibular system than is the case with their terrestrial relatives. Unlike in 491 most terrestrial settings locomotion in an aquatic environment offers movements in all three 492 dimensions of space. To adapt to such new environments, their body plan needs to change. 493 Metriorhynchids, for example evolved flippers and lost their osteoderms to allow the animal 494 more flexible body movements and possibly axial undulation (Massare, 1988). Strong 495 496 buoyancy forces reduce the effects of weight and allow more complex body movements in a highly viscous aquatic medium and are some of the factors that might influence 497 morphological changes in the labyrinth organ in a variety of secondarily aquatic vertebrates. 498 Pelagic sauropterygians and sea turtles have more bulbous labyrinths with thicker 499

semicircular canals than other reptiles (Georgi and Sipla, 2008; Neenan et al., 2017; Evers et
al., 2019), cetaceans miniaturized their labyrinth drastically compared to their body size
(Spoor et al., 2002), and other aquatic mammals such as sirenians and seals also reduced their
semicircular canals but to a lesser degree (Hyrtl, 1845; Loza et al., 2017). Hence it is clear
that specific aspects of their labyrinth morphologies convergently evolved in independent
pelagic lineages.

In crocodylomorphs three different labyrinth morphologies have been recognized, each of which is characteristic of a particular habitat: terrestrial, semiaquatic, and pelagic (Schwab et al., 2020; Fig. 7). Schwab et al. (2020) used statistical tests to demonstrate the existence of distinctive ear morphologies for these three habitat categories, but only briefly described and figured representative inner ears of each type. We here describe and figure them in more detail (Fig. 7).

The terrestrial morphology is present in early crocodylomorphs, such as sphenosuchiangrade taxa like *Junggarsuchus sloani* (Fig. 7A-D). It is characterised by very long, thin and round semicircular canals and a thin crus commune. The anterior and posterior canals embay the characteristic 'M' shape and the anterior canal extends higher dorsally. The cochlear duct is relatively long compared to the rest of the labyrinth and medially bends in *Junggarsuchus sloani*, but is straight in other non-crocodyliform crocodylomorphs.

The semiaquatic labyrinth morphology is seen in modern crocodylians as well as nonmetriorhynchid thalattosuchians like teleosauroids. This labyrinth morphology is an intermediate form with thicker semicircular canals and wider crus commune, and it is overall dorsoventrally shorter than in the terrestrial forms (Fig. 7E-H, M-P). Non-metriorhynchid thalattosuchians have a reduced curvature and triangular shaped semicircular canals compared to modern crocodylians, where the curvature appears more rounded and the overall canal is oval in cross section. Teleosauroids have the 'M' shape which is mostly lost in

525 modern crocodylians. The cochlear duct is straight and shorter than in the terrestrial 526 morphology, but longer than in the pelagic one. Modern crocodylians have a twisted cochlear 527 duct.

528 The third labyrinth morphology is seen in pelagic taxa, including the NHMUK PV OR 32617 specimen that we describe here. This unique labyrinth morphology is present in all 529 sampled metriorhynchids and characterised by a dorsoventrally compact labyrinth, thick 530 semicircular canals, an enlarged vestibule and a presumably short cochlear duct (the end of 531 the cochlear duct is not enclosed by bone and hence just a minimum length can be estimated). 532 533 This morphology is most extreme in the derived metriorhynchids, like Cricosaurus (Fig. 7I-L) and Torvoneustes. The canals have more pronounced curvature compared to non-534 metriorhynchid thalattosuchians and the anterior ampulla is more bulbous. Unlike the 535 terrestrial and semiaquatic labyrinths, the anterior canal of the pelagic labyrinths becomes 536 smaller and just slightly exceeds the posterior canal dorsally. 537

One possible explanation for the change in morphology in the labyrinths of pelagic 538 species (which evolved from semiaquatic ancestors) is their wider range of body movements 539 in a three-dimensional aquatic context, not simply because they lived in the water. For 540 example, cetaceans are highly agile and reduced their semicircular canals, however sirenians 541 (manatees and dugongs) are also fully aquatic but are not very agile and hence do not have 542 reduced canals (Ekdale, 2013). The reduced cetacean semicircular canals may also prevent 543 544 overstimulation during exaggerated head movements associated with swimming and diving, as it is generally assumed that longer and more arching semicircular canals are more sensitive 545 546 to rotations in space (Spoor et al., 2002; Ekdale, 2016). This could also be the case for marine reptiles such as metriorhynchids. Another explanation could be that because of neck 547 shortening in metriorhynchids (they had five instead of seven postaxial cervical vertebrae; 548 Andrews, 1913; Young and Andrade, 2009) head and gaze stabilization (vestibulo-ocular and 549

vestibulo-collic reflexes) became less important, which resulted in a reduction of the 550 semicircular canals, as has been suggested for cetaceans and plesiosaurs (Spoor et al., 2002; 551 Neenan et al., 2017). A third hypothesis for the morphological shift seen in pelagic labyrinths 552 relates to diving. Thickening of the semicircular canals could have been driven by either an 553 increased membranous duct diameter or an increased perilymphatic space (space between the 554 membranous and bony labyrinth, filled with perilymphatic fluid). Diving results in changes in 555 hydrostatic pressure, which must be compensated for via fluid movement within the 556 perilymphatic duct system. Thicker canals could have increased the perilymphatic space and 557 558 functioned as a buffer to protect the sensory system. This hypothesis has been proposed for pelagic deep diving turtles (Evers et al., 2019). Hence, the change in morphology could have 559 been a response to changing sensory requirements as secondarily aquatic vertebrates, such as 560 metriorhynchids, moved into deeper, more open waters. 561

562

563

CONCLUSION

564

Our computed tomography (CT) study of the braincase of the marine thalattosuchian 565 'Metriorhynchus' cf. 'M.' brachyrhynchus provides one of the best looks yet at the 566 neurosensory system and internal cranial anatomy of a fully aquatic, pelagic, fast-swimming 567 metriorhynchid. Key neuroanatomical features described herein include a unique flexure in 568 the brain endocast that has not been noted in other thalattosuchians or marine reptiles, the 569 function of which is currently unknown. NHMUK PV OR 32617 also had enlarged cerebral 570 571 hemispheres to potentially process increased sensory input and a more rounded pituitary fossa that could be related to water regulation in the ocean or housed an enlarged venous sinus. The 572 well-developed vasculature system noted here has previously been reported for other 573 574 metriorhynchids as well as basal thalattosuchians, suggesting it evolved early in

575	thalattosuchian history. Whether it functioned to support salt glands and/or for aide
576	thermoregulation is currently unclear. Furthermore, a pelagic bony labyrinth morphology is
577	present, with an overall dorsoventrally short morphology, thick semicircular canals and an
578	enlarged vestibule. This might have allowed the animal to better interact with its three
579	dimensional aquatic environment, it may have helped with pressure changes during diving or
580	might be due to reduced neck and head and gaze stabilisation. These new insights into the
581	endocranial evolution of thalattosuchians allow us to better understand their transition into
582	the open oceans. Some of these features (e.g., salt glands) seem to have evolved prior to
583	metriorhynchids becoming fully pelagic, and were thus 'exaptations' that helped transition
584	them to life in the water, whereas others (e.g., pelagic labyrinth morphology) apparently
585	developed only after metriorhynchids moved into the open oceans.
586	
587	ACKNOWLEDGMENTS
588	
589	We are grateful to S. Maidment (Natural History Museum London) for providing
590	access to the specimen. We thank the editor and two anonymous reviewers for their helpful
591	suggestions. This project is supported by a Leverhulme Trust Research Project grant (RPG-
592	2017-167) to PI S.L.B., which funds J.A.S. and M.T.Y. Y.H was partially supported by the
593	ANPCyT-PICT-2016-0267 and ANPCyT-PICT 2016-1039. L.M.W. acknowledges support
594	from United States National Science Foundation Grants IOB-0517257, IOS-1050154, and
595	IOS-1456503.
596	
597	LITERATURE CITED
508	

- Andrews, C. W. 1913. Descriptive Catalogue of the Marine Reptiles of the Oxford Clay, Part
 Two. British Museum (Natural History), London, 206 pp.
- 601 Baczko, M. B. von and J. B. Desojo. 2016. Cranial anatomy and palaeoneurology of the
- 602 archosaur *Riojasuchus tenuisceps* from the Los Colorados Formation, La Rioja,
- Argentinia. PLoS ONE 11:e0148575.
- 604 Baczko, M. B. von, J. R. A. Taborda, and J. B. Desojo. 2018. Paleoneuroanatomy of the
- 605 aetosaur Neoaetosauroides engaeus (Archosauria: Pseudosuchia) and its
- paleobiological implications among archosauriforms. PeerJ 6:e5456.
- Balanoff, A., G. S. Bever, T. B. Rowe, and M. A. Norell. 2013. Evolutionary origins of the
 avian brain. Nature 501:93–97.
- Brignon, A. 2016. L'abbé Bacheley et la découverte des premiers dinosaures et crocodiliens
 marins dans le Jurassique des Vaches Noires (Callovien/Oxfordien, Normandie).
- 611 Comptes Rendus Palevol 15: 595–605.
- Brusatte, S. L., A. Muir, M. T. Young, S. Walsh, L. Steel, and L. M. Witmer. 2016. The
- braincase and neurosensory anatomy of an Early Jurassic marine crocodylomorph:
- 614 implications for crocodylian sinus evolution and sensory transitions. Anatomical
 615 Record 299:1511–1530.
- Chiarenza, A., D. Foffa, M. T. Young, G. Insacco, A. Cau, G. Carnevale, and R. Catanzariti.
 2015. The youngest record of metriorhynchid crocodylomorphs, with implications for
 the extinction of Thalattosuchia. Cretaceous Research 56:608–616.
- 619 Colbert, E. H. 1946. Sebecus, representative of a peculiar suborder of fossil Crocodylia from
- 620 Patagonia. Bulletin of the American Museum of Natural History 87:2017–2270.
- 621 Dufeau, D. L., and L. M. Witmer. 2015. Ontogeny of the middle-ear air-sinus system in
- 622 Alligator mississippiensis (Archosauria: Crocodylia). PLoS ONE 10:e0137060.

- Ekdale, E. G. 2013. Comparative Anatomy of the Bony Labyrinth (Inner Ear) of Placental
 Mammals. PLoS ONE 8:e66624.
- Ekdale, E. G. 2016. Form and function of the mammalian inner ear. Journal of Anatomy
 228:324–337.
- 627 Evers, S., J. M. Neenan, G. S. Ferreira, I. Werneburg, P. M. Barett, and R. B. J. Benson.
- 628 2019. Neurovascular anatomy of the protostegid turtle Rhinochelys pulchriceps and
- 629 comparisons of membranous and endosseous labyrinth shape in an extant turtle.
- 630 Zoological Journal of the Linnean Society 20:1–29.
- 631 Fernández, M., and Z. Gasparini. 2008. Salt glands in the Jurassic metriorhynchid Geosaurus:
- 632 implications for the evolution of osmoregulation in Mesozoic crocodyliforms.
- 633 Naturwissenschaften 95:79–84.
- Fernández, M. S., A. P. Carabajal, Z. Gasparini, and G. Chong Diaz. 2011. A metriorhynchid
 crocodyliform braincase from northern Chile. Journal of Vertebrate Paleontology
 31:369–377.
- Fernández, M. S., Y. Herrera, V. V. Vennari, L. Campos, M. de la Fuente, M. Talevi, and B.
 Aguirre-Urreta. 2019. Marine reptiles from the Jurassic/Cretaceous transition at the
- High Andes, Mendoza, Argentina. Journal of South American Earth Sciences 92:658–
 640 673.
- Foffa, D., and M. T. Young. 2014. The cranial osteology of *Tyrannoneustes lythrodectikos*(Crocodylomorpha: Metriorhynchidae) from the Middle Jurassic of Europe. PeerJ
 2:e608.
- Foffa, D., M. T. Young, T. L. Stubbs, K. G. Dexter, and S. L. Brusatte. 2018. The long-term
 ecology and evolution of marine reptiles in a Jurassic seaway. Nature Ecology and
 Evolution 2:1548–1555.

Fraas, E. 1902. Die Meer-Krocodilier (Thalattosuchia) des oberen Jura unter specieller
berücksichtigung von *Dacosaurus* und *Geosaurus*. Palaeontographica 49:1–72.

649 Georgi, J. A., and J. S. Sipla. 2008. Comparative and Functional Anatomy of Balance in

- 650 Aquatic Reptiles and Birds; pp. 233–256 in J. G. M. Thewissen and S. Nummela (ed.),
- 651 Sensory Evolution on the Threshold: Adaptations in Secondarily Aquatic Vertebrates.

652 University of California Press, Berkeley, California.

653 Heller, H. 1942. The posterior pituitary principles of a species of reptile (Tropidonotus

654 *natrix*) with some remarks on the comparative physiology of the posterior pituitary

gland generally. The Journal of Physiology 101:317–3262.

- Heller, H. 1950. The comparative physiology of the neurohypophysis. Cellular and Molecular
 Life Sciences 6:368–376.
- 658 Herrera, Y. 2015. Metriorhynchidae (Crocodylomorpha: Thalattosuchia) from Upper

59 Jurassic-Lower Cretaceous of Neuquén Basin (Argentina), with comments on the

660 natural cast of the brain; pp. 159–171 in M. S. Fernández and Y. Herrera (ed.), Reptiles

661 Extintos-Volumen en Homenaje a Zulma Gasparini. Publicación Electrónica de la

662 Asociación Paleontológica, Buenos Aires, Argentina.

Herrera, Y., and V. V. Vennari. 2015. Cranial anatomy and neuroanatomical features of a
new specimen of Geosaurini (Crocodylomorpha: Metriorhynchidae) from west-central
Argentina. Historical Biology 27:33–41.

666 Herrera, Y., M. S. Fernandez, and Z. Gasparini. 2013. The snout of Cricosaurus

- *araucanensis*: a case study in novel anatomy of the nasal region of metriorhynchids.
 Lethaia 46:331–340.
- Herrera, Y., Z. Gasparini, and M. S. Fernández. 2015. *Purranisaurus potens* Rusconi, an
 enigmatic metriorhynchid from the Late Jurassic–Early Cretaceous of the Neuquén
 Basin. Journal of Vertebrate Paleontology 35:e904790.

- Herrera, Y., J. M. Leardi, and M. Fernandez. S. 2018 Braincase and endocranial anatomy of
 two thalattosuchian crocodylomorphs and their relevance in understanding their
 adaptations to the marine environment. PeerJ 6:e5686.
- Herrera, Y, M. S. Fernández, G. S. Lamas, L. Campos, M. Talevi, and Z. Gasparini. 2017.
- 676 Morphology of the sacral region and reproductive strategies of Metriorhynchidae: a 677 counter-inductive approach. Earth and Environmental Science Transactions of the
- 678 Royal Society of Edinburgh 106:247–255.
- Hopson, J. A. 1979. Paleoneurology; pp. 39–146 in C. Gans (ed.) Biology of the Reptilia.
 Academic Press, New York.
- Hyrtl, J. 1845. Vergleichend-anatomische Untersuchungen Über das innere Gehörorgan des
 Menschen und der Säugethiere. Ehrlich, Prague, 139 pp.
- Jouve, S. 2009. The skull of *Teleosaurus cadomensis* (Crocodylomorpha; Thalattosuchia),
 and phylogenetic analysis of Thalattosuchia. Journal of Vertebrate Paleontology 29:88–
 102.
- Kelley, N. P., and N. D. Pyenson. 2015. Evolutionary innovation and ecology in marine
 tetrapods from the Triassic to the Anthropocene. Science 348: aaa3716.
- 688 Kley, N. J., J. W. Sertich, A. H. Turner, D. W. Kause, P. M. O'Connor, and J. A. Georgi.
- 689 2010. Craniofacialmorphology of *Simosuchus clarki* (Crocodyliformes: Notosuchia)
- from the Late Cretaceous of Madagascar. Journal of Vertebrate Paleontology 30:13–98.
- 691 Lautenschlager, S. and R. J. Butler. Neural and endocranial anatomy of Triassic phytosaurian
- reptiles and convergence with fossils and modern crocodylians. PeerJ 4:e2251.
- 693 Larsson, H. C. E., T. A. Dececchi, and F. C. Montefeltro. 2011. A new metriorhynchid
- 694 (Crocodyliformes, Thalattosuchia) from the early cretaceous of Colombia (Rosa Blanco
- 695 Formation, late Valanginian). Ameghiniana 48:R86–R87.

- 696 Leardi, J. M., D. Pol, and J. M. Clark. 2017. Detailed anatomy of the braincase of
- *Macelognathus vagans* Marsh, 1884 (Archosauria, Crocodylomorpha) using high
 resolution tomography and new insights on basal crocodylomorph phylogeny. PeerJ
 5:e2801.
- Lepage, Y., E. Buffetaut, S. Hua, J. E. Martin, and J. Tabouelle. 2008. Catalogue descriptif,
 anatomique, géologique et historique des fossiles présentés à l'exposition « Les
 Crocodiliens fossiles de Normandie ». Bulletin de la Société Géologique de Normandie
 et des Amis du Muséum du Havre 95:5–152.
- 704 Loza, C. M., A. E. Latimer, M. R. Sánchez-Villagra, and A. A. Carlini. 2017. Sensory
- anatomy of the most aquatic of carnivorans: the Antarctic Ross seal, and convergenceswith other mammals. Biological Letters 13:20170489.
- 707 Lydekker, R. 1888. Catalogue of the Fossil Reptilia and Amphibia in the British Museum
- 708 (Natural History), Cromwell Road, S.W., Part 1. Containing the Orders Ornithosauria,
- 709 Crocodilia, Dinosauria, Squamata, Rhynchocephalia, and Proterosauria. British
- 710 Museum of Natural History, London, 309 pp.
- 711 Martill, D. M. 1986. The diet of Metriorhynchus, a Mesozoic marine crocodile. Neues
- 712 Jahrbuch für Geologie und Paläontologie 10:621–625.
- Massare, J. A. 1988. Swimming capabilities of Mesozoic marine reptiles: implications for
 method of predation. Paleobiology 14:187–205.
- 715 Motani, R. 2009. The Evolution of Marine Reptiles. Evo Edu Outreach 2:224–235.
- 716 Neenan, J. M., T. Reich, S. W. Evers, P. S. Druckenmiller, D. F. A. E. Voeten, J. N.
- 717 Choiniere, P. M. Barrett, S. E. Pierce, and R. B. J. Benson. 2017. Evolution of the
- sauropterygian labyrinth with increasingly pelagic lifestyles. Current Biology 27:3852-
- 719 3858.

720	Nesbitt, S. J., M. Stocker, W. G. Parker, T. A. Wood, C. A. Sidor and K. D. Angielczyk.
721	2018. The braincase and endocast of Parringtonia gracilis, a Middle Triassic suchian
722	(Archosauris: Pseudosuchia). Society of Vertebrate Paleotology 37:122-141.
723	Ősi, A., M. T. Young, A. Galácz, and M. Rabi. 2018. A new large-bodied thalattosuchian
724	crocodyliform from the lower Jurassic (Toarcian) of Hungary, with further evidence of
725	the mosaic acquisition of marine adaptations in Metriorhynchoidea. PeerJ 6:e4668.
726	Parrilla-Bel, J., M. T. Young, M. Moreno-Azanza, and J. I. Canudo. 2013. The first
727	metriorhynchid crocodyliform from the Middle Jurassic of Spain, with implications for
728	evolution of the subclade Rhacheosaurini. PLoS ONE 8:e54275.
729	Pierce, S. E. and M. J. Benton. 2006. Pelagosaurus typus Bronn, 1841 (Mesoeucrocodylia:
730	Thalattosuchia) from the Upper Lias (Toarcian, Lower Jurassic) of Somerset, England.
731	Journal of Vertebrate Paleontology 26:621-635.
732	Pierce, S. E., M. Williams, and R. B. J. Benson. 2017. Virtual reconstruction of the
733	endocranial anatomy of the early Jurassic marine crocodylomorph Pelagosaurus typus
734	(Thalattosuchia). PeerJ 5:e3225.
735	Pol, D., and Z. Gasparini. 2009. Skull anatomy of Dakosaurus andiniensis
736	(Thalattosuchia:Crocodylomorpha) and the phylogenetic position of Thalattosuchia.
737	Journal of Systematic Palaeontology 7:163-197.
738	Porter, W. R., J. C. Sedlmayr, and Witmer L. M. 2016. Vascular patterns in the heads of
739	crocodilians: blood vessels and sites of thermal exchange. Journal of Anatomy
740	229:800-824.
741	Porter, W. R., and L. M. Witmer. 2019. Vascular patterns in the heads of dinosaurs: evidence
742	for blood vessels, sites of thermal exchange, and their role in physiological
743	thermoregulatory strategies. Anatomical Record 303:1075-1103.

744	Rogers, S. W. 1999. Allosaurus, crocodiles, and birds: evolutionary clues from spiral
745	computed tomography of an endocast. The Anatomical Record 1999:162-173.
746	Sachs, S., M. T. Young, and J. Hornung. 2020. The enigma of Enaliosuchus, and a
747	reassessment of the Early Cretaceous fossil record of Metriorhynchidae. Cretaceous
748	Research 114:104479.
749	Schindelin, J., I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S.
750	Preibisch, C. Rueden, S. Saalfeld, and B. Schmid. 2012 Fiji: an open-source platform
751	for biological-image analysis. Nature Methods 9, 676-682.
752	Schwab, J. A., M. T. Young, J. M. Neenan, S. A. Walsh, L. M. Witmer, Y. Herrera, R.
753	Allain, C. A. Brochu, J. N. Choiniere, J. M. Clark, K. N. Dollman, S. Etches, G.
754	Fritsch, P. M. Gignac, A. Ruebenstahl, S. Sachs, A. H. Turner, P. Vignaud, E. W.
755	Wilberg, X. Xu, L. E. Zanno and S. L. Brusatte. 2020 Inner ear sensory system changes
756	as extinct crocodylomorphs transitioned from land to water. PNAS 117:10422-10428.
757	Séon, N., R. Amiot, J. Martin, M. T. Young, H. Middleton, F. Fourel, L. Picot, X. Valentin,
758	and C. Lécuyer. 2020. Thermophysiologies of Jurassic marine crocodylomorphs
759	inferred from the oxygen isotope composition of their tooth apatite. Philosophical
760	Transactions of the Royal Society B 375:20190139.
761	Sipla, J. S., and F. Spoor. 2008. The Physics and Physiology of Balance; pp. 227-232 in J. G.
762	M. Thewissen and S. Nummela (ed.), Sensory Evolution on the Threshold: Adaptations
763	in Secondarily Aquatic Vertebrates. University of California Press, Berkeley,
764	California.
765	Spoor, F., S. Bajpai, S. T. Hussain, K. Kumar, and J. G. M Thewissen. 2002. Vestibular
766	evidence for the evolution of aquatic behaviour in early cetaceans. Nature 417:163-
767	166.

768	Walsh, S., and A. Milner. 2011. Halcyornis toliapicus(Aves: Lower Eocene, England)
769	indicates advancedneuromorphology in Mesozoic Neornithes. Journal of Systematic
770	Palaeontology 9:173–181.

- 771 Watanabe, A., P. M. Gignac, A. M. Balanoff, T. L. Green, N. J. Kley, and M. A. Norell.
- 2019. Are endocasts good proxies for brain size and shape in archosaurs throughoutontogeny?. Journal of Anatomy 234:291-305.
- Wharton, D. S. 2000. An enlarged endocranial venous system in *Steneosaurus pictaviensis*(Crocodylia: Thalattosuchia) from the Upper Jurassic of Les Lourdines, France.
- Comptes Rendus de l'Académie des Sciences—Series IIA—Earth and Planetary
 Science 331:221–226.
- Wilberg, E. W. 2015. A new metriorhynchoid (Crocodylomorpha, Thalattosuchia) from the
 Middle Jurassic of Oregon and the evolutionary timing of marine adaptations in
- thalattosuchian crocodylomorphs. Journal of Vertebrate Paleontology 35:e902846.
- 781 Wilkinson, L. E., M. T. Young, and M. T. Benton. 2008. A new metriorhynchid crocodylian
- 782 (Mesoeucrocodylia: Thalattosuchia) from the Kimmeridgian (Upper Jurassic) of
- 783 Wiltshire, UK. Palaeontology 51:1307–1333.
- Witmer, L. M., R. C. Ridgely, D. L. Dufeau, and M. C. Semones. 2008. Using CT to peer
 into the past: 3Dvisualization of the brain and ear regions of birds,crocodiles and
 nonavian dinosaurs; pp. 67–87 in H. Endo and R. Frey (ed.), Anatomical Imaging:
 Towards A New Morphology. Springer, Berlin.
- Young, M. T., and M. B. Andrade. 2009. What is *Geosaurus*? Redescription of *Geosaurus giganteus* (Thalattosuchia: Metriorhynchidae) from the Upper Jurassic of Bayern,
- Germany. Zoological Journal of the Linnean Society 157:551-585.
- 791 Young, M. T., S. L. Brusatte, M. Ruta, and M. B. Andrade. 2010. The evolution of
- 792 Metriorhynchoidea (Mesoeucrocodylia, Thalattosuchia): an integrated approach using

geometrics morphometrics, analysis of disparity and biomechanics. Zoological Journal
of the Linnean Society 158:801–859.

Young, M. T., M. B. Andrade, S. Etches, and B. L. Beatty. 2013. A new metriorhynchid

796 crocodylomorph from the Lower Kimmeridge Clay Formation (Late Jurassic) of England, with implications for the evolution of dermatocranium ornamentation in 797 Geosaurini. Zoological Journal of the Linnean Society 169:820-848. 798 Young, M. T., A. Brignon, S. Sachs, J. J. Hornung, D. Foffa, J. J. N. Kitson, M. M. Johnson, 799 and L. Steel. 2020. Cutting the Gordian knot: a historical and taxonomic revision of the 800 801 Jurassic crocodylomorph Metriorhynchus. Zoological Journal of the Linnean Society XX, 1-44. Young, M. T., S. L. Brusatte, M. B. Andrade, J. B. Desojo, B. L. Beatty, L. 802 Steel, M. S. Fernández, M. Sakamoto, J. I. Ruiz-Omeñaca, and R. R. Schoch. 2012. The 803 cranial osteology and feeding ecology of the metriorhynchid crocodylomorph genera 804 Dakosaurus and Plesiosuchus from the Late Jurassic of Europe. PLoS ONE 7:e44985. 805 806 Submitted August 28, 2020; revisions received November 29, 2020; accepted Month DD, 807

808 YYYY.

809

795

810 FIGURE CAPTIONS

811 FIGURE 1. Phylogenetic relationships of crocodylomorphs with 'Metriorhynchus' cf. 'M.'

brachyrhynchus (NHMUK PV OR 32617) in blue. Phylogeny based on Young et al. (2020).
Size (182x125mm)

814

815 FIGURE 2. Photographs and line drawing of the braincase of 'Metriorhynchus' cf. 'M.'

816 brachyrhynchus (NHMUK PV OR 32617). A, dorsal; B, right lateral; C, posterior; D,

817 ventral; E, left lateral. Abbreviations: bo, basioccipital; bs, basisphenoid; cf, carotid

foramen; fm, foramen magnum; fr, frontal; ls, laterosphenoid; mpf, median pharyngeal
foramen; oc, occipital condyle; ot, otoccipital; pa, parietal; pro, prootic; so, supraoccipital;
tof, temporo-orbital foramen. Roman numerals designate cranial nerve openings. Roman
numerals designate cranial nerves. Scale bar equals 5cm. Size (182x100mm)

822

FIGURE 3. Digital reconstruction of the braincase of '*Metriorhynchus*' cf. '*M*.'

brachyrhynchus (NHMUK PV OR 32617). A, dorsal; B, right lateral; C, anterior; D, ventral;
E, left lateral; F, posterior. Abbreviations: bo, basioccipital; bs, basisphenoid; cf, carotid
foramen; fm, foramen magnum; fr, frontal; ls, laterosphenoid; mpf, median pharyngeal
foramen; oc, occipital condyle; ot, otoccipital; pa, parietal; pro, prootic; so, supraoccipital;
tof, temporo-orbital foramen. Roman numerals designate cranial nerve openings. Scale bar
equals 5cm. Size (182x106.17mm)

830

FIGURE 4. Internal endocranial anatomy of 'Metriorhynchus' cf. 'M.' brachyrhynchus 831 (NHMUK PV OR 32617). A, dorsal; B, right lateral; C, anterior; D, ventral; E, left lateral; F, 832 posterior. Abbreviations: bl, bony labyrinth; cer, cerebrum; dvs, dural venous sinus; ic, 833 internal carotid artery; mps, median pharyngeal sinus; mpt, median pharyngeal tube; mpsd, 834 median pharyngeal sinus diverticulum; otd, otoccipital diverticulum; pf, pituitary fossa; ptd, 835 pterygoid diverticulum; pts, pharyngotympanic sinus; ptt, pharyngotympanic tubes; shc, 836 837 second hypoglossal canal. Roman numerals designate cranial nerves. Scale bar equals 5cm. Size (182x112mm) 838

839

840 FIGURE 5. Internal endocranial anatomy of 'Metriorhynchus' cf. 'M.' brachyrhynchus

841 (NHMUK PV OR 32617). A, dorsal; B, right lateral; C, anterior; D, ventral; E, left lateral; F,

842 posterior. Abbreviations: bl, bony labyrinth; cer, cerebrum; dvs, dural venous sinus; ic,

internal carotid artery; oa, orbital artery; pf, pituitary fossa; rmcv, rostral middle cerebral
vein; shc, second hypoglossal canal. Roman numerals designate cranial nerves. Scale bar
equals 5cm. Size (182x112mm)

846

FIGURE 6. Right bony labyrinth of '*Metriorhynchus*' cf. '*M.*' *brachyrhynchus* (NHMUK PV
OR 32617). A, lateral; B, medial; C, anterior; D, posterior; E, dorsal. Abbreviations: asc,
anterior semicircular canal; cc, crus commune; cd, cochlear duct; lsc, lateral semicircular
canal; psc, posterior semicircular canal; ve, vestibule. Scale bar equals 1cm. Size
(182x54.61mm)

852

853 FIGURE 7. Comparative right bony labyrinths. A-D, Junggarsuchus sloani (IVPP V14010);

854 E-H, Osteolaemus tetraspis (FMNH 98936); I-L, Cricosaurus araucanensis (MLP 72-IV-7-

1); M-P, Plagiophthalmosuchus cf. gracilirostris (NHMUK PV OR 33095). Orientations of

the labyrinths are lateral, anterior, posterior and dorsal from left to right. Coloured boxes

857 indicate habitat, red, terrestrial; orange, semiaquatic; blue, pelagic. Scale bars equal 1cm. Size

858 (182x90mm)