THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Soil carbon loss by experimental warming in a tropical forest

Citation for published version:

Nottingham, A, Meir, P, Velasquez, E & Turner, BL 2020, 'Soil carbon loss by experimental warming in a tropical forest', *Nature*. <https://doi.org/10.1038/s41586-020-2566-4>

Digital Object Identifier (DOI):

[10.1038/s41586-020-2566-4](https://doi.org/10.1038/s41586-020-2566-4)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Nature

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Article to Nature; accepted 6 March with only editorial corrections needed.

Soil carbon loss from a tropical forest in response to *in situ* experimental warming

Andrew T. Nottingham^{1,2}, Patrick Meir^{1,3}, Esther Velasquez², Benjamin L. Turner²

¹School of Geosciences, University of Edinburgh, Crew Building, Kings Buildings, Edinburgh EH9 3FF, UK

²Smithsonian Tropical Research Institute, 0843-03092, Balboa, Ancon, Republic of Panama

³Research School of Biology, Australian National University, Canberra, ACT 2601, Australia

Tropical soils contain a third of global soil carbon¹, so destabilization of soil organic matter caused by the approximate 4°C warming predicted for tropical regions this century could accelerate climate change by releasing additional carbon dioxide (CO₂) to the atmosphere²⁻⁵. Theory predicts that warming should cause only modest carbon loss in tropical soils relative to those at higher latitudes^{4,6}, but there have been no warming experiments in tropical forests to test this prediction⁷. Here we show that *in situ* experimental warming of a lowland tropical forest soil on Barro Colorado Island, Panama, caused an unexpectedly large increase in soil CO₂ emissions. Two years of warming of the whole soil profile by 4°C increased CO₂ emission by 55% compared to soils at ambient temperature. The additional CO₂ originated from heterotrophic rather than autotrophic sources and equated to a loss of 8.2 ± 4.2 (± 1 SE) Mg C ha⁻¹ yr⁻¹ from the breakdown of soil organic matter. During this time, we detected no acclimation of respiration rates, no thermal compensation or change in temperature sensitivity of enzyme activities, and no change in microbial carbon-use efficiency. These results demonstrate a high sensitivity of soil carbon in tropical forests to warming, which represents a potentially substantial positive feedback to climate change.

28 Tropical forests play a large role in the global carbon (C) cycle, because they exchange more
29 CO₂ with the atmosphere than any other ecosystem, contain over two-thirds of terrestrial plant
30 biomass⁸ and harbour over a quarter of global soil C (ref. 1). Between 30 and 50% of the C respired
31 from tropical forests originates from soil, most of which is derived from the decomposition of organic
32 matter⁹⁻¹¹. Thus, even a small increase in respiration from tropical forest soils could have a large effect
33 on atmospheric CO₂ concentrations, with consequences for global climate.

34 There is considerable concern that increased global temperatures will destabilize soil C and
35 increase the flux of CO₂ from soil to the atmosphere²⁻⁵. Experiments in temperate and arctic regions
36 have consistently found that short-term (< 2 years) warming increases the soil CO₂ efflux by an average
37 of $46 \pm 8\%$ compared to soil at ambient temperature^{2,3,5,12}. For the tropics, it is expected that the
38 response of soil C to warming will be smaller than at higher latitudes, because kinetic theory predicts
39 that the intrinsic temperature sensitivity of reaction rates is reduced at higher temperatures^{4,13}, and
40 meta-analyses of warming experiments have shown that the temperature sensitivity of soil C loss
41 increases with latitude⁶. However, the extent to which intrinsic temperature sensitivity translates into
42 actual ('apparent') temperature sensitivity depends on co-variation of other environmental factors that
43 influence respiration, such as soil moisture and substrate availability^{4,14}. As there have been no *in situ*
44 warming experiments conducted in tropical forests, the apparent temperature sensitivity of soil organic
45 matter in this biome remains unknown. As a result, earth-system models continue to use kinetic theory
46 to define the temperature sensitivity of soil C¹⁵, limiting how they predict the response of tropical
47 forests to global environmental change^{16,17}.

48 Several factors could influence the apparent temperature sensitivity of soil organic matter. For
49 example, soil warming is typically accompanied by soil drying, which can either reduce respiration in
50 aerobic soils by reducing water availability, or increase respiration in waterlogged soils by increasing
51 oxygen availability^{4,7,14}. Warming can also affect respiration rates by inducing changes in biotic
52 processes, such as the physiological response or community composition of microbes, or changes in

53 substrate availability to decomposers¹³. In experiments performed at higher latitudes, temperature-
54 adaptive or compensatory responses of microbial communities and enzyme activities have been shown
55 to modulate the effect of warming on the soil C cycle^{2,13}. In the tropics, future novel warm temperature
56 maxima could exceed critical biochemical thresholds^{7,18}, with added complexity emerging from altered
57 interactions among species-rich plant and microbial communities¹⁹, and from covarying changes in
58 hydrological and nutrient cycles⁷.

59 Here, we present results from the first soil warming experiment in a lowland tropical forest
60 (SWELTR: Soil Warming Experiment in Lowland Tropical Rainforest). The experiment tests the
61 response of the whole soil profile to the 4°C warming predicted for tropical latitudes by the end of this
62 century¹⁶ (Fig. 1A; Extended Data Figs. 1-3). SWELTR consists of five pairs of circular control and
63 warmed plots, evenly distributed within approximately 1 ha of seasonally-moist lowland tropical forest
64 on Barro Colorado Island, Panama. The soils are moderately weathered Dystric Eutrudepts
65 (Inceptisols) that have developed on the volcanic facies of the Bohio Formation (Extended Data Table
66 1, see methods). Each warmed plot has a ground surface area of ~20 m² heated to 1.2 m depth, resulting
67 in a total of 120 m³ of warmed soil across the experiment.

68 Two years of experimental warming increased soil CO₂ emissions by 55%, from 18.8 ± 1.9 Mg
69 C ha⁻¹ yr⁻¹ in control plots to 29.2 ± 5.0 Mg C ha⁻¹ yr⁻¹ in warmed plots (treatment effect, $p < 0.05$; Fig.
70 2; Extended Data Table 2). The soil CO₂ emission rate from unheated (i.e., control) plots is
71 representative of tropical forests worldwide (8–40 Mg C ha⁻¹ yr⁻¹), including in the Amazon basin (12–
72 24 Mg C ha⁻¹ yr⁻¹)²⁰. Using exclusion and ingrowth cores to partition respiration from heterotrophic
73 (soil-derived) and autotrophic (root-derived) sources, we find that the increase in CO₂ efflux was
74 derived predominantly from heterotrophic sources, whether from the decomposition of fresh-litter
75 inputs or pre-existing soil organic matter (Figs. 2-3, Extended Data Fig. 5). Soil-derived respiration
76 increased from 12.0 ± 2.1 Mg C ha⁻¹ yr⁻¹ in control plots to 20.1 ± 4.2 Mg C ha⁻¹ yr⁻¹ in warmed plots
77 (a 68% increase of 8.2 Mg C ha⁻¹ yr⁻¹; treatment effect, $p < 0.05$), while root-derived respiration was

not altered significantly ($p = 0.21$; 6.8 ± 1.2 and 9.0 ± 3.4 Mg C ha⁻¹ yr⁻¹ in control and warmed plots, respectively; Fig. 3, Extended Data Table 3).

It is possible that the large warming-induced increase in soil CO₂ efflux was due in part to soil drying, because the warmed plots were slightly drier than the controls, particularly in the early wet season (Extended Data Fig. 3). In wet soils, soil drying can increase respiration by increasing the supply of oxygen to heterotrophic microbes. In contrast, soil drying under aerobic conditions can reduce heterotrophic respiration by promoting water limitation²⁰. Here, we find a marginally non-significant effect of soil moisture on CO₂ efflux across seasons (Extended Data Table 2; annual $p = 0.69$, wet-season $p = 0.07$ and dry-season $p = 0.06$), consistent with the parabolic relationship of CO₂ flux with soil moisture for this site²⁰. However, there was no direct effect of warming on soil moisture (for surface soils $p = 0.19$, whole-profile $p = 0.24$; Extended Data Tables 4-5), and the interaction between soil moisture and warming in the CO₂ efflux model was not significant (annually and for individual seasons; $p > 0.2$; Extended Data Table 2), indicating that the warming effect on CO₂ efflux was not influenced by soil moisture. Furthermore, soil moisture was not correlated with soil CO₂ efflux in the warmed plots (Extended Data Fig. 3); and drying during the early wet season in warmed soil (Extended Data Fig. 3) should decrease rather than increase CO₂ efflux, because the soil was aerobic during this period and below the moisture content of 0.45 m³ m⁻³ at which soil CO₂ efflux peaks in this forest²⁰. Our data thus show that although soil moisture influenced soil CO₂ efflux and that warmed plots were slightly drier than control plots, particularly during the early wet season, this did not contribute significantly to the increased CO₂ efflux from warmed soil.

There was no moderation of the warming-induced increase in soil CO₂ efflux over the two years of the experiment. Such a moderation might be expected in the long-term, whether through substrate limitation, adaptation of microbial communities (through changes in microbial carbon-use-efficiency; CUE) or thermal compensation of enzyme activities (reduced $V_{\max}$ at higher temperatures)^{2,13,21}. We found no reduction in extractable or mineralized nitrogen or phosphorus with

warming, as would be expected under nutrient limitation (Extended Data Figs. 6-7). Almost all hydrolytic enzymes were unaffected by warming (Extended Data Figs. 6-7), except for β -xylanase – an enzyme involved in hemicellulose degradation – for which activity increased with warming during the wet season, an opposite response to that predicted by thermal compensation¹⁹. The temperature sensitivity of enzyme activity (Q_{10} of $V_{\max}$) was unaffected by warming (Extended Data Fig. 8), indicating no dampening effect on soil C breakdown as a result of decreased enzyme flexibility (which would cause decreased affinity of the enzyme for its substrate), as expected at warmer temperatures²². Microbial CUE, which broadly represents C stabilised in biomass relative to C lost in respiration and can promote long-term (decadal) soil C loss^{2,21}, was unaffected by warming (Treatment effect, $p = 0.37$; Extended Data Figs. 6-7). Indeed, microbial C increased with warming at the annual scale (treatment effect, annual scale, $p = 0.02$; with a marginally non-significant increase at the seasonal scale, $p < 0.1$ Extended Data Fig. 7), suggesting slightly increased growth in response to greater organic matter turnover in the absence of nutrient constraints to C degradation, which did not translate into changes in CUE.

Our finding that tropical forest soil C has a high apparent temperature sensitivity challenges the prevailing expectation that the temperature sensitivity of soil C is lower in the tropics compared to cooler ecosystems at higher latitudes^{4,6}. The 55% increase in total soil CO₂ emissions we report here following two years of 4°C whole-profile warming (18.8 to 29.2 Mg C ha⁻¹ yr⁻¹) is larger than that found in a temperate forest using a similar whole-soil-profile experimental design (34–37% increase over two years of 4°C whole-profile warming; 13 to 17.5 Mg C ha⁻¹ yr⁻¹)⁵. In addition, the rate of additional soil C loss (8.2 Mg C ha⁻¹ yr⁻¹) is greater than for all the studies in a recent meta-analysis of surface-only soil warming experiments at higher latitude sites (all loss rates < 5 Mg C ha⁻¹ yr⁻¹)³. The expectation that the temperature sensitivity of soil C breakdown is lower in the tropics compared to higher latitudes, based in part on kinetic theory and commonly used to describe soil C responses in earth-system models¹⁵, is therefore not consistent with the (‘apparent’)^{4,14} temperature sensitivity of

128 the breakdown of tropical forest soil C reported here. Our results are, however, consistent with recent
129 atmospheric and satellite measurements, which have shown a high sensitivity of ecosystem-scale C
130 cycling in tropical regions in response to interannual temperature variation^{23,24}. Our findings suggest
131 that soils contribute a major component of these ecosystem-scale responses to warming.

132 This high apparent temperature sensitivity of tropical forest soil C under *in situ* experimental
133 warming must arise through the temperature response of covarying ecosystem properties rather than
134 as the sole consequence of intrinsic kinetic processes. Although our data do not provide conclusive
135 mechanistic evidence for the marked increase in soil C loss from warmed soil, several findings point
136 to a possible explanation: i) the general absence of thermal compensation in enzyme activities (no
137 decrease in $V_{\max}$); ii) no change in the temperature sensitivity of enzymes under warming (no decrease
138 in Q_{10} of $V_{\max}$); and iii) no moderating thermal response of microbial CUE (Extended Data Figs. 6-
139 7)^{21,25,26}. Together, our results indicate that organic matter degradation increased under warming with
140 no moderating responses or acclimation during our study period among microbial communities, or the
141 enzymes they synthesise.

142 This surprisingly large loss of soil C from warmed soil represents a substantial positive climate
143 feedback over the period of this study. The additional C loss from warming observed here is of similar
144 magnitude to annual litterfall-C inputs at this site ($5\text{--}7 \text{ Mg C ha}^{-1} \text{ yr}^{-1}$)²⁷ and is equivalent to
145 approximately 13% of the total soil C stock, or 30% of gross primary productivity ($27.5 \text{ Mg C ha}^{-1} \text{ yr}^{-1}$)²⁸. Extrapolation of the first two years of C loss in our experiment across the entire tropical forest
147 soil C stock (502 Pg C)¹ indicates a global loss of $>65 \text{ Pg C}$ with 4°C warming this century, which is
148 consistent with estimated C loss based on a five-year soil translocation experiment in tropical forests
149 elsewhere²⁹. In the light of these findings, earlier estimates of global soil C loss under 4°C warming,
150 which were based on experiments performed at higher latitudes ($120\text{--}190 \text{ Pg C}$)^{2,30}, underestimate the
151 magnitude of this global earth-atmosphere feedback.

We expect that the rate of soil C loss will eventually decline in warmed soils as substrate limitation increases, but we do not know how long this will take, nor whether the long-term soil C balance will be affected by plant-soil interactions or changes in soil microbial communities as they adapt to warmer temperatures^{2,12,13,21}. The nature of these longer-term responses will determine the strength of this positive earth-atmosphere feedback, already significant in the short-term, in contributing to further climate warming.

References

- 1 Jackson, R. B. *et al.* The ecology of soil carbon: pools, vulnerabilities, and biotic and abiotic controls. *Annual Review of Ecology, Evolution, and Systematics* **48**, 419-445, doi:10.1146/annurev-ecolsys-112414-054234 (2017).
- 2 Melillo, J. M. *et al.* Long-term pattern and magnitude of soil carbon feedback to the climate system in a warming world. *Science* **358**, 101-104, doi:10.1126/science.aan2874 (2017).
- 3 van Gestel, N. *et al.* Predicting soil carbon loss with warming. *Nature* **554**, E4-E5, doi:10.1038/nature20150 (2018).
- 4 Davidson, E. A. & Janssens, I. A. Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* **440**, 165-173, doi:10.1038/nature04514 (2006).
- 5 Hicks Pries, C. E., Castanha, C., Porras, R. C. & Torn, M. S. The whole-soil carbon flux in response to warming. *Science* **355**, 1420-1422, doi: 10.1126/science.aal1319 (2017).
- 6 Carey, J. C. *et al.* Temperature response of soil respiration largely unaltered with experimental warming. *P Natl Acad Sci USA* **113**, 13797-13802, doi:10.1073/pnas.160536511 (2016).
- 7 Wood, T. E. *et al.* in *Ecosystem Consequences of Soil Warming: Microbes, Vegetation, Fauna and Soil Biogeochemistry* (ed J. Mohan) Ch. 14, 385-439 (Academic Press, 2019).
- 8 Pan, Y. *et al.* A large and persistent carbon sink in the world's forests. *Science* **333**, 988-993, doi:10.1126/science.1201609 (2011).
- 9 Anderson-Teixeira, K. J., Wang, M. M. H., McGarvey, J. C. & LeBauer, D. S. Carbon dynamics of mature and regrowth tropical forests derived from a pantropical database (TropForC-db). *Global Change Biol* **22**, 1690-1709, doi:10.1111/gcb.13226 (2016).
- 10 Malhi, Y. The productivity, metabolism and carbon cycle of tropical forest vegetation. *J Ecol* **100**, 65-75, doi:10.1111/j.1365-2745.2011.01916.x (2012).
- 11 Chambers, J. Q. *et al.* Respiration from a tropical forest ecosystem: Partitioning of sources and low carbon use efficiency. *Ecol Appl* **14**, S72-S88 (2004). doi: 10.1890/01-6012.
- 12 Romero-Olivares, A. L., Allison, S. D. & Treseder, K. K. Soil microbes and their response to experimental warming over time: A meta-analysis of field studies. *Soil Biol Biochem* **107**, 32-40, doi:10.1016/j.soilbio.2016.12.026 (2017).

- 192 13 Tang, J. *et al.* in *Ecosystem Consequences of Soil Warming: Microbes, Vegetation, Fauna*
193 *and Soil Biogeochemistry* (ed J. Mohan) Ch. 8, 175-201 (Academic Press, 2019).
- 194 14 Nottingham, A. T. *et al.* Climate warming and soil carbon in tropical forests: insights
195 from an elevation gradient in the Peruvian Andes. *Bioscience* **65**, 906-921,
196 doi:10.1093/biosci/biv109 (2015).
- 197 15 Todd-Brown, K. E. O. *et al.* Causes of variation in soil carbon predictions from CMIP5
198 Earth system models and comparison with observations. *Biogeosciences* **10**, 1717–1736,
199 doi:10.5194/bgd-9-14437-2012 (2013).
- 200 16 Hoegh-Guldberg, O. *et al.* in *Global Warming of 1.5°C. An IPCC Special Report on the*
201 *impacts of global warming of 1.5°C above pre-industrial levels and related global*
202 *greenhouse gas emission pathways, in the context of strengthening the global response to*
203 *the threat of climate change, sustainable development, and efforts to eradicate poverty*
204 (Cambridge University Press, 2018).
- 205 17 Cox, P. M. *et al.* Sensitivity of tropical carbon to climate change constrained by carbon
206 dioxide variability. *Nature* **494**, 341-344, doi:10.1038/Nature11882 (2013).
- 207 18 Mora, C. *et al.* The projected timing of climate departure from recent variability. *Nature*
208 **502**, 183, doi: 10.1038/Nature12540 (2013).
- 209 19 Steidinger, B. S. *et al.* Climatic controls of decomposition drive the global biogeography
210 of forest-tree symbioses. *Nature* **569**, 404, doi:10.1038/s41586-019-1128-0 (2019).
- 211 20 Rubio, V. E. & Detto, M. Spatiotemporal variability of soil respiration in a seasonal
212 tropical forest. *Ecol Evol* **7**, 7104-7116, doi:10.1002/ece3.3267 (2017).
- 213 21 Frey, S. D., Lee, J., Melillo, J. M. & Six, J. The temperature response of soil microbial
214 efficiency and its feedback to climate. *Nat Clim Change* **3**, 395-398,
215 doi:10.1038/Nclimate1796 (2013).
- 216 22 Bradford, M. A. Thermal adaptation of decomposer communities in warming soils. *Front*
217 *Microbiol* **4**, doi:Artn 333, doi 10.3389/Fmicb.2013.00333 (2013).
- 218 23 Wang, X. H. *et al.* A two-fold increase of carbon cycle sensitivity to tropical temperature
219 variations. *Nature* **506**, 212–215, doi:10.1038/nature12915 (2014).
- 220 24 Liu, J. J. *et al.* Contrasting carbon cycle responses of the tropical continents to the 2015-
221 2016 El Nino. *Science* **358**, doi:10.1126/science.aam5690 (2017).
- 222 25 Karhu, K. *et al.* Temperature sensitivity of soil respiration rates enhanced by microbial
223 community response. *Nature* **513**, 81-84, doi:10.1038/nature13604 (2014).
- 224 26 Dacal, M., Bradford, M. A., Plaza, C., Maestre, F. T. & Garcia-Palacios, P. Soil microbial
225 respiration adapts to ambient temperature in global drylands. *Nat Ecol Evol* **3**, 232-+,
226 doi:10.1038/s41559-018-0770-5 (2019).
- 227 27 Wieder, R. K. & Wright, S. J. Tropical forest litter dynamics and dry season irrigation on
228 Barro Colorado Island, Panama. *Ecology* **76**, 1971-1979, doi: 10.2307/1940727 (1995).
- 229 28 Chave, J. *et al.* Spatial and temporal variation of biomass in a tropical forest: results from
230 a large census plot in Panama. *J Ecol* **91**, 240-252, doi: 10.1046/j.1365-
231 2745.2003.00757.x (2003).
- 232 29 Nottingham, A. T. *et al.* Microbial responses to warming enhance soil carbon loss
233 following translocation across a tropical forest elevation gradient. *Ecol Lett* **22**, 1889-
234 1899, doi:10.1111/ele.13379 (2019).
- 235 30 Crowther, T. W. *et al.* Quantifying global soil carbon losses in response to warming.
236 *Nature* **540**, 104-108, doi:10.1038/nature20150 (2016).
- 237

238 **Supplementary Information** is linked to the online version of the paper

Acknowledgements | This study was supported by a European Union Marie-Curie Fellowship FP7-2012-329360 and a Smithsonian Tropical Research Institute Tupper Fellowship to ATN, a Smithsonian Institution Scholarly Studies Grant to BT and Klaus Winter, and a NERC grant NE/K01627X/1 (NERC HTMF-BALI) to PM, who was also supported by an Australian Research Council (ARC) grant DP170104091. We thank Oris Acevado, Dayana Agudo, Aleksandra Bielnicka, Melissa Cano, David Dominguez, Milton Garcia, Matthew Larsen, Maria Jose Montero, Julio Rodriguez, Irene Torres, William Weislo, Klaus Winter and Joe Wright for their contributions to SWELTR.

Author contributions | ATN conceived the study with BLT and PM. ATN, EV and BLT performed the study. ATN analysed the data and wrote the paper with BLT and PM.

Author Information | The authors declare no competing financial interests. Reprints and permissions information is available at www.nature.com/reprints. Correspondence and requests for materials should be addressed to A.T.N. (anotting@ed.ac.uk).

Data availability statement | The data supporting the findings of this study are available within the paper and its supplementary information files, with further information available from the corresponding author upon reasonable request.

Supplementary Information and Extended Data for this manuscript include the following:

Supplementary Materials and Methods

Extended Data Table 1 to 5

Extended Data Figs. 1 to 8

Figure 1 | Mean differences in (a) soil temperature and (b) soil moisture content between control and warmed plots by depth. Data are the means of continuous measurements from integrated soil temperature and moisture probes (Campbell CS655) for the two-year period after the warming treatment began (Dec 2016 – Dec 2018). The error bars represent one standard error of the temporal variation across five plots (Treatment effect $p < 0.001$ for all comparisons, based on the temporal variation across $n = 5$ plots). The vertical lines show the mean soil temperature across the soil profile, which is 26.18°C for control plots and 30.14°C for warmed plots (3.97°C difference).

Figure 2 | Soil CO₂ efflux from control and warmed soils over two years. Panel (a) shows the total soil CO₂ flux during the study period (2017-2019). Panel (b) shows the total soil CO₂ flux partitioned into soil-derived and root-derived components (b), relative to the beginning of the warming treatment. Measurements were made every one or two weeks. Points represent the mean value of five plots, with error bars representing one standard error of the spatial variation. The box plots represent the mean and temporal variation over sequential 100-day periods to show seasonal dynamics. The dotted vertical line (relative day = -150) is when installation and testing of warming plots began (during this period each plot was warmed by 4°C relative to controls for a period of 1-2 weeks); the dashed vertical line (relative day = 0; 1 November 2017) shows when all five warming plots were switched on permanently. The shaded areas represent dry seasons (1 January–1 April). Soil CO₂ efflux was significantly higher in warmed plots for annual data and for dry or wet seasons individually (Extended Data Table 2). The error bars for points represent one standard error of the spatial variation (n = 5 plots); the error bars for boxes represent one standard error of the temporal variation (averaged over 100 days).

296

297

298 **Figure 3 | The annual soil CO₂ efflux (total), partitioned into soil-derived and root-derived components.**

299 **a.** The pre-treatment period (Jan-May 2016; predominantly dry-season). **b.** The two years with warming. The
300 figures show total CO₂ (Total) and the root-derived and soil-derived components calculated using CO₂ efflux
301 from partition cores (equations 1-2). Differences between control and warmed plots are shown by asterisks
302 where $p \leq 0.05$ or as non-significant (ns); determined using mixed effect models (Extended Data Tables 2-3).
303 The error bars represent one standard error of the variation across the five plots over the study period.

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318 **METHODS**

319

320 *Site and experiment.* The experiment is situated in approximately 1 ha area of seasonally moist
321 lowland tropical forest on Barro Colorado Island, Panama³¹. Within the plot area the dominant tree
322 species include *Anacardium excelsum* and *Poulsenia armata*. The soils are Inceptisols (Fine,
323 isohyperthermic, Dystric Eutrudepts) that are rich in clay (~54% profile-weighted clay concentration)
324 and secondary metal oxides and are developed on the volcanic facies of the Bohio Formation, a basaltic
325 conglomerate of Oligocene age³². Although these moderately weathered Inceptisols are less infertile
326 than soils under large areas of lowland tropical forests, Inceptisols still account for 14% of total land
327 area in the tropics (Ultisols and Oxisols account for 20% and 23%, respectively)³³, and soil respiration
328 in the control plots in our experiment is comparable to that in lowland tropical forests in general,
329 including those on Ultisols and Oxisols (see main text, ref. 20).

330 The SWELTR experiment consists of 10 circular plots (five paired plots ‘warm’ and ‘control’).
331 Each plot measures 5 m diameter, with approximately 10 m between each plot-pair and a minimum of
332 20 m between different plot-pairs. The warmed plots contain two heating structures, each consisting
333 of eight 1-m long stainless-steel rods, connected by approximately 50 cm of flexible stainless-steel
334 conduit. We used stainless-steel T-junctions at the top of each rod (adjoining the flexible conduit) and
335 conical caps at the bottom of each rod. The final structure was 1.2 m tall. Inside each of the structures,
336 we threaded 25 m of heating cable (SLMCAB10120BF, Briskheat, Columbus, USA) and filled the
337 remaining space in the rods and conduit with quartz sand, selected for its high thermal conductivity.
338 The complete structure was welded to seal the heating cable and sand inside. Two of these structures
339 were buried around a 3.5 diameter circumference, with the top of the flexible conduit 5 cm
340 belowground. Thus, each plot contained 50 m of heating cable inserted to 1.2 m depth, encircling a 3.5
341 m diameter area; with an effective heated plot area of 5 m diameter. The experiment heats

342 approximately 120 m³ soil in total (5 plots x 5 m diameter by 1.2 m depth). The plot design and heating
343 methodology follows that of Hanson et al. (ref. 34) and Hicks Pries et al. (ref. 5).

344

345 ***Temperature control.*** Each warming and control plot was connected to a thermostat system, which
346 maintained soil temperature in the warmed plots at 4°C above ambient temperature. The thermostat
347 system consisted of three integrated temperature and moisture sensors per plot (CS655 Reflectometer,
348 Campbell Scientific) inserted to 0-20, 50-70 and 100-120 cm depth at the mid-radius point in each
349 plot, which were connected to a control unit (one control unit for each plot pair; five in total). The
350 control units consisted of waterproof (IP68) enclosures containing a solid-state power controller
351 (DA10-24C0-0000, Watlow), relay (12V single channel), datalogger (CR1000, Campbell Scientific)
352 and 12V and 120V power supply. Temperature in each warmed plot was therefore maintained at 4°C
353 above the temperature in each corresponding paired control plot, based on the average temperature
354 from 0-120 cm depth at the mid-radius point in each plot. The average temperature differential over
355 two years was 3.97°C, which was the average of 2.7°C at 0-20 cm depth, 4.0°C at 50-70 cm depth and
356 5.2°C at 100-120 cm depth.

357 This experimental design has been shown to warm the soil approximately uniformly across the
358 soil volume, with minor anomalies of warmer soil very close to the heating rods (< 10 cm) and slightly
359 cooler surface soils due to heat-transfer to the air^{5, 34}. Therefore, surface soils were slightly cooler
360 compared to subsoils, although the response of surface rather than subsoils will likely dominate the
361 warming response across the soil profile because they contain greater organic matter (two thirds of the
362 C stock occurs in the upper 50 cm of the soil profile; Extended data Table 1). The heating structures
363 were installed during May-July 2016, and plots were tested during June-October 2016. The testing
364 phase consisted of heating each plot by 4°C for approximately 2-week periods. The experiment was
365 switched on in full on 1 November 2016.

366

367 ***Soil gas-exchange and partitioning.*** Soil CO₂ efflux was measured every two weeks at four
368 systematically distributed locations within each plot from 2016 until 2019 using an infra-red gas
369 analyser (IRGA Li-8100; LI-COR Biosciences, Nebraska, USA). The soil collars for soil CO₂ efflux
370 measurements were assigned to zones within each plot ('centre 1', 'centre 2', 'side 1' and 'side 2') and
371 were relocated randomly within each zone every three months, for long-term within-plot spatial
372 independence. Soil CO₂ efflux was also measured every two weeks for four root-partition cores per
373 plot (2 × root-exclusion and 2 × root-ingrowth) to determine soil- and root-derived components of the
374 CO₂ efflux. At the same time as soil CO₂ efflux measurements, we measured soil temperature (using
375 a HI98509 thermometer probe; Hanna Instruments, USA) and soil moisture (using a Thetaprobe;
376 Delta-T, Cambridge, UK) at 0-20 cm soil depth for a random location within 1 m radius of each soil
377 collar, or within the root-partition cores.

378 Root-exclusion cores were made from PVC tubing (30 cm height, 10 cm diameter) with a 1
379 µm nylon mesh base for drainage. Root-ingrowth cores (i.e. disturbance controls) had additional
380 windows (~340 cm²) covered with 2 mm mesh around the sides³⁵. In each plot, two root-exclusion
381 cores and two root-ingrowth cores were buried within each plot, approximately 30 cm from the heating
382 cable (where the soil profile is warmed on average by 4°C).

383 The percentage contributions of fine roots and rhizo-microorganisms (root-derived) and free-
384 living heterotrophic microorganisms (soil-derived) to the total soil CO₂ efflux were calculated as
385 follows:

386 Equation 1: $\text{Soil-derived (\%)} = (\text{root-exclusion core CO}_2 \text{ efflux} / \text{root-ingrowth core CO}_2 \text{ efflux}) \times$
387 100

388 Equation 2: $\text{Root-derived (\%)} = 100 - \text{Soil-derived (\%)}$

389

390 Total soil CO₂ efflux measured for soil collars was multiplied by the results from equations 1–2 to
391 estimate the absolute contributions of root-derived (roots, rhizo-microbial and mycorrhizal) and soil-

392 derived (free-living microbial through the decomposition of litter and soil organic matter)
393 components³⁵. The partition cores were buried to 0-25 cm depth, where 95% fine roots occur³⁶ –
394 therefore the soil component consists of soil-derived CO₂ from the entire soil profile (with very
395 marginal contribution from fine roots at > 20 cm depth).

396

397 ***Soil properties.*** Soil was sampled before the experimental treatments began (to 100 cm depth;
398 Extended Data Table 1) and then every three months following the beginning of the experiment (0-10
399 cm; average responses in Extended Data Fig. 4) within the plots at a point where the surface soil is
400 evenly heated (at approximately 30 cm distance from the heating structure), and analysed for soil
401 properties: total elements, available nutrients, microbial CNP and enzyme activities using standard
402 procedures (see SI methods). We calculated microbial carbon-use-efficiency (CUE) using microbial
403 CNP and enzyme activity data using a stoichiometric method³⁷. Here we describe the responses
404 following two years of warming, by using the average change in soil properties over two years (average
405 of eight temporal measurements per plot, with n = 5 per plot). Full details on these analytical methods
406 are provided in SI.

407

408 ***Statistical analyses.*** Treatment (warming) effects on time-averaged total and partitioned CO₂
409 emissions and other soil properties (nutrients, microbial properties), were tested using ANOVA.
410 Treatment effects on soil CO₂ emissions were further tested using mixed effects models with CO₂
411 emission as the response variable and warming-treatment, soil moisture, season, warming × soil
412 moisture and season × soil moisture as fixed effects and plot number as random effect³⁸. We tested
413 both with and without a repeated measures effect because within-plot soil CO₂ efflux measurements
414 were partially spatially independent (within-plot locations of soil collars were changed every three
415 months, see soil gas exchange measurements above). Treatment effects on partitioned root- and soil-
416 derived CO₂ efflux components were tested using repeated measures mixed models with time and plot

417 number as random effects. Treatment effects on soil-surface moisture (0-20 cm depth) were tested
 418 using mixed effects models with soil moisture as the response variable and warming-treatment, season,
 419 warming $\times$ season as fixed effects and plot number as random effect. For whole-profile soil moisture
 420 we included time as an additional random effect (soil-surface moisture and temperature measurements
 421 were fully independent, whereas whole-profile measurements were repeated measures of fixed points).
 422 Further details on these approaches are provided in SI. All statistical analyses were performed in R
 423 (version 3.5.2).

424

- 425 31 Leigh, E. G. J. *Tropical Forest Ecology: A View from Barro Colorado Island*. (Oxford
 426 University Press, 1999).
- 427 32 Woodring, W. P. Geology of Barro Colorado Island. *Smithsonian Miscellaneous Collections*
 428 **135**, 1 – 39 (1958).
- 429 33 Sanchez, P. A. & Logan, T. J. Myths and Science About the Chemistry and Fertility of Soils
 430 in the Tropics. *SSSA Spec Publ* **29**, 35-46 (1992).
- 431 34 Hanson, P. J. *et al.* A method for experimental heating of intact soil profiles for
 432 application to climate change experiments. *Global Change Biol* **17**, 1083-1096,
 433 doi:10.1111/j.1365-2486.2010.02221.x (2011).
- 434 35 Nottingham, A. T., Turner, B. L., Winter, K., van der Heijden, M. G. A. & Tanner, E. V. J.
 435 Arbuscular mycorrhizal mycelial respiration in a moist tropical forest. *New Phytol* **186**,
 436 957-967, doi:10.1111/J.1469-8137.2010.03226.X (2010).
- 437 36 Cavelier, J. Fine-root biomass and soil properties in a semideciduous and a lower
 438 montane rain-forest in Panama. *Plant Soil* **142**, 187-201 (1992).
- 439 37 Sinsabaugh, R. L. *et al.* Stoichiometry of microbial carbon use efficiency in soils.
 440 *Ecological Monographs* **86**, 172-189, doi:10.1890/15-2110.1 (2016).
- 441 38 Zuur, A. F., Ieno, E. N. & Smith, G. M. Introduction to mixed modelling. *Stat Biol Health*,
 442 125-142 (2007).
- 443