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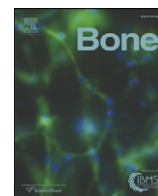
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Review

New insights into NPP1 function: Lessons from clinical and animal studies

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ABSTRACT

The recent elucidation of rare human genetic disorders resulting from mutations in ectonucleotide pyrophosphatase/phosphodiesterase (*ENPP1*), also known as plasma cell membrane glycoprotein 1 (PC-1), has highlighted the vital importance of this molecule in human health and disease.

Generalised arterial calcification in infants (GACI), a frequently lethal disease, has been reported in recessive inactivating mutations in *ENPP1*. Recent findings have also linked hypophosphataemia to a lack of NPP1 function. A number of human genetic studies have indicated that NPP1 is a vital regulator that influences a wide range of tissues through various signalling pathways and when disrupted can lead to significant pathology.

The function of *Enpp1* has been widely studied in rodent models, where both the mutant tiptoe walking (*ttw/ttw*) mouse and genetically engineered *Enpp1*^{−/−} mice show significant alterations in skeletal and soft tissue mineralisation, calcium/phosphate balance and glucose homeostasis. These models therefore provide important tools with which to study the potential mechanisms underpinning the human diseases associated with altered NPP1.

This review will focus on the recent advances in our current knowledge of the actions of NPP1 in relation to bone disease, cardiovascular pathologies and diabetes. A fuller understanding of the mechanisms through which NPP1 exerts its pathological effects may stimulate the development of novel therapeutic strategies for patients at risk from the devastating clinical outcomes associated with disrupted NPP1 function.

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Introduction

Rare human genetic disorders resulting from loss-of-function mutations in the ectonucleotide pyrophosphatase/phosphodiesterase

(*ENPP1*) gene, also known as plasma cell membrane glycoprotein 1 (PC-1), have highlighted the importance of this molecule in human health and disease. Generalised arterial calcification in infants (GACI) and severe hypophosphataemia have been reported in recessive inactivating mutations in the *ENPP1* gene [1–4]. Together with the association between polymorphisms in *ENPP1* and *ALPL*, the gene encoding for tissue non-specific alkaline phosphatase (TNAP), and reduced bone size and mineral density in the Caucasian population [5] these findings indicate that the *ENPP1* gene is required for normal inhibition of ectopic

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mineralisation while also being essential for mineralisation in the bone. Furthermore, levels of *ENPP1* expression have been reported to be elevated in humans showing high levels of insulin resistance [6–8] suggesting an important role in glucose homeostasis and insulin signalling. These human studies indicate that the NPP1 protein is a vital regulator that influences a wide range of tissues through various signalling pathways and when disrupted can lead to significant pathology.

The function of NPP1 has been widely studied in rodent models, where both the mutant tiptoe walking (*ttw/ttw*) mouse [9–14], and the transgenically engineered *Enpp1*^{−/−} mice [15,16], show changes in skeletal and soft tissue mineralisation, calcium/phosphate and glucose homeostasis, mimicking the diseases seen in human subjects. Furthermore, by acting remotely on the balance of circulating minerals and glucose, NPP1 has a wider reaching impact on both skeletal and soft tissue structure and metabolism. This review will focus on the recent advances in current understanding of the role of the NPP1 protein in these pathways and outline the importance of this research in bone diseases, cardiovascular diseases and diabetes.

Genetics and function of NPP1

The nucleoside pyrophosphatase/phosphodiesterases (NPPs) are an important group of enzymes with an extensive functional range that are distributed widely and are highly conserved between species. In humans the NPP family consists of 5 proteins of which NPP1 and NPP3 show similar structure and function and the genes encoding for these two proteins have been mapped to human chromosome 6q22–23 [17,18]. Despite the close sequence homology of the *NPP* genes between species it has been reported that the 5′ flanking region is far less conserved, leading to different regulation and gene expression patterns in different species [19].

The NPP1 protein is a membrane spanning homodimer and, when cleaved, the extracellular domain can function as a secreted circulating protein. In a very revealing review Bollen and colleagues have discussed the biochemistry of the NPP family and have summarised the localisation of *ENPP1* gene expression [19]. *ENPP1* is expressed in a wide range of tissues including cartilage, heart, kidney, parathyroid and

skeletal muscle, and it is highly expressed in vascular smooth muscle cells (VSMCs), osteoblasts and chondrocytes [20–22].

NPPs have wide substrate specificity, and the hydrolysis of pyrophosphate bonds (for example, in ATP) and phosphodiester bonds (for example, in oligonucleotides) to produce nucleoside 5′-monophosphates makes NPPs extremely important in extracellular nucleotide metabolism and extracellular signalling. NPP1 (EC3.1.4.1) is a 104 kDa type II transmembrane protein consisting of a small intracellular region (between 10 and 80 residues) and a larger extracellular domain (830 residues) which contains the catalytic site [23]. Phosphodiesterases are classified as enzymes that hydrolyse diesters of phosphoric acid into phosphomonoesters, and can be classified into two main groups – those that act on lipids or on nucleotides. Pyrophosphatases are acid anhydride hydrolases that catalyse the breakdown of diphosphate bonds and are biologically important in the cleavage of ATP. NPP1 hydrolyses ATP to generate either inorganic pyrophosphate (PP_i) plus AMP or inorganic phosphate (P_i) plus ADP in a two stage process via either ADP or a phosphate bound intermediate, respectively (Fig. 1) [19,24]. It has also been reported that NPPs can convert AMP into adenosine and P_i [25,26] although conflicting reports suggest that AMP competitively inhibits NPP activity [27]. All of the products of these hydrolysis reactions are essential in cellular signalling and function, the effects of which vary between tissues.

Basic mechanisms of bone formation and the role of NPP1 in skeletal mineralisation

In order to understand the functions of NPP1 it is important to appreciate the physiological process of mineralisation in bone. This relies on the deposition of hydroxyapatite (HA) onto a collagenous matrix, and is a highly regulated process that requires the correct concentration of calcium (Ca²⁺) and P_i to precipitate as HA crystals. Mineralisation is thought to be a two stage process, the first of which occurs within matrix vesicles (MVs) [28] where the conditions are optimal for the initial precipitation of HA. The second stage consists of the propagation of HA formation onto the extracellular matrix

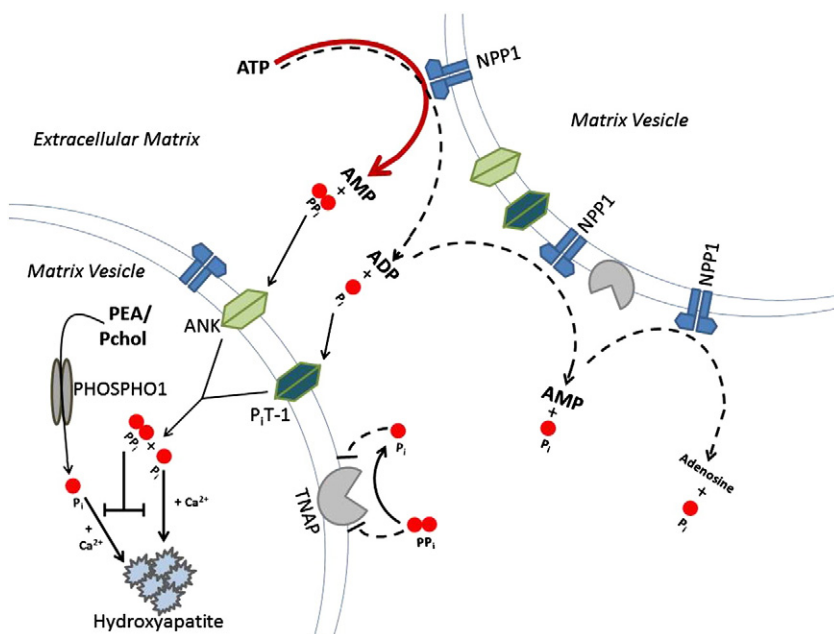


Fig. 1. Schematic showing the role of NPP1 in ATP hydrolysis and the downstream effects on bone mineralisation. The primary function of NPP1 is the hydrolysis of ATP into AMP and PP_i, although it is involved in further degradation of pyrophosphate bonds to generate ADP, adenosine and P_i (secondary reactions denoted by dotted lines). PP_i is converted into P_i by TNAP and the transport of PP_i and P_i through the cell membrane is mediated by ANK and P_i-T-1 respectively. Within the matrix vesicle PHOSPHO1 can generate further P_i by the hydrolysis of PEA and Pchol. PP_i acts to inhibit hydroxyapatite formation, while P_i promotes this process, thus the balance of these two mediators is highly important in regulating mineralisation.

(ECM) following the disruption of the MVs. While P_i acts to promote precipitation of HA crystals, PP_i has a dual role as an inhibitor of HA formation and a precursor to P_i . The ratio of P_i to PP_i is controlled by a complex interaction between the following enzymes: NPP1, tissue-nonspecific alkaline phosphatase (TNAP), phosphoethanolamine/phosphocholine phosphatase (PHOSPHO1), type III sodium-dependent P_i co-transporter 1 (P_i T-1) and ankylosis protein (ANK) (Fig. 1) [29–35]. NPP1 extracellularly generates PP_i and AMP by hydrolysis of ATP [36]. Intracellularly to extracellular channelling of P_i and PP_i is mediated by ANK [37,38] and P_i T-1. TNAP, which hydrolyses PP_i in the ECM to release P_i and PHOSPHO1, which hydrolyses phosphocholine (Pchol) and phosphoethanolamine (PEA) to produce P_i inside the MVs, act to control the presence of each substrate during the two stages of mineralisation [35]. Further feedback signalling allows mediation of the mineralisation process; both P_i and PP_i inhibit the enzymatic activity of TNAP [39], and both exogenous P_i and PP_i induce osteopontin (OPN), a bone sialoprotein which inhibits mineral formation through limiting HA crystal precipitation and growth [30,32,39].

The link between defective NPP1 expression and altered mineralisation was initially demonstrated in the mutant “tiptoe walking” (*ttw/ttw*) mouse model. These animals are homozygous for a GRT substitution resulting in the introduction of a stop codon in the NPP1 coding sequence. The subsequent truncated protein leads to the loss of a vital calcium binding domain and two putative glycosylation sites [13]. The *ttw/ttw* mouse phenotype includes the postnatal development of progressive ankylosing intervertebral and peripheral joint hyperostosis, as well as spontaneous arterial and articular cartilage calcification and increased vertebral cortical bone formation [9,11–14]. Transgenic mice that are homozygous for a disruption in Exon 9 of the *Enpp1* gene exhibit abnormalities that are almost identical to those present in *ttw/ttw* mice [15]. These include decreased levels of extracellular PP_i , with phenotypic features including significant alterations in bone mineralisation in long bones and calvariae, and pathologic, severe peri-spinal soft tissue and arterial calcification [16,30,32].

The calvariae of *Enpp1*^{−/−} mice are hyper-calcified in vivo, and calvarial osteoblasts derived from *Enpp1*^{−/−} mice show reduced extracellular PP_i levels, and a concomitant increase in calcification in vitro [30]. These abnormalities can be rescued by transfection with NPP1 but not with NPP3. A significant reduction in the mineralisation inhibitor OPN has also been observed in *Enpp1*^{−/−} osteoblasts, indicating that NPP1 not only has a direct effect on PP_i concentration, but also has an indirect effect on the process of calcification by regulating the expression of other cellular regulators [32].

Enpp1^{−/−} mice also show significant defects in long-bone mineralisation [16,40]. *Enpp1*^{−/−} mice have reduced trabecular bone mass (Fig. 2) and cortical thickness of both the tibia and femur, characterised by disruption of the structural and mechanical properties, the severity of which increases with age [40]. This is likely to be a

direct effect of lack of NPP1 activity, but the reduced body weight observed in *Enpp1*^{−/−} mice will reduce the loading on the bones and thus may also have an effect on their structure.

Previous evaluation of the mineralisation of bones from 10-day-old *Enpp1*^{−/−} and [*Enpp1*^{−/−}; *Akp2*^{−/−}] double knockout mice has indicated that the effects of *Enpp1* ablation on an *Akp2* null background is site-specific [16]. Thus, in contrast to the normalisation of the degree of mineralisation seen in the joints, calvariae, vertebrae and soft tissues as a consequence of ablating both NPP1 and TNAP function, the long bones of these double knockout mice appeared to remain hypomineralised. Furthermore, calcified nodule formation and mineral deposition are inhibited to a higher extent in osteoblasts isolated from *Enpp1*^{−/−} bone marrow than calvarial osteoblasts isolated from the same animal, further indicating that loss of NPP1 activity affects skeletal sites in a site-specific manner [16]. The hypomineralisation observed in the long bones of *Enpp1*^{−/−} mice may be related to relatively low levels of endogenous PP_i when compared to the calvaria [16]. Thus, in long bones, the complete deletion of NPP1 activity would further reduce extracellular PP_i to abnormally low levels. This would result in insufficient PP_i substrate for TNAP to generate P_i for normal mineral formation.

It has been widely reported that PP_i functions to regulate both osteoblast and chondrocyte differentiation. However, it has recently been shown that NPP1 regulates osteoblastic gene expression and cellular differentiation in calvarial osteoblasts independent of PP_i and P_i [41]. Nam and colleagues have provided evidence that NPP1 is an inducer of osteoblast differentiation, demonstrating that FGF-2 signalling induces *Enpp1* expression in pre-osteoblasts but not in differentiated osteoblasts. Furthermore, MC3T3E1(C4) cells that over-expressed *Enpp1* showed enhanced osteoblastic gene expression. Conversely, defective osteoblast differentiation was observed in both calvariae extracted from *Enpp1*^{−/−} mice and MC3T3E1(C4) cells treated with *Enpp1* targeting short hairpin RNAs. Therefore inhibition of osteoblast differentiation due to lack of NPP1 activity may also contribute to the reduced mineralisation in the long bones observed in *Enpp1*^{−/−} mice.

Wild-type mice show reduced bone resorption with advancing age which is consistent with the attainment of the adult skeleton. Interestingly *Enpp1*^{−/−} mice maintain similar levels of osteoclast activity at 6 and 22 weeks of age, indicating an increase in functional activity of osteoclasts [40], the cells that mediate bone resorption. Furthermore, treatment of *ttw/ttw* mice with calcitonin, a known inhibitor of osteoclast function and putative suppressor of osteoblastic bone formation, has been shown to reverse the osteopenic phenotype [14]. This study indicates that accelerated periosteal bone formation in *ttw/ttw* mice is suppressed by calcitonin but does not assess the role of osteoclasts in the correction of the osteopenic phenotype.

The role of NPP1 in soft tissue calcification

Ectopic calcification occurs throughout the body causing clinical complications particularly when seen in the aorta, cardiac valves and in the myocardium, where mineralisation is a serious risk factor in cardiovascular disease. It is also observed in tendons, cartilages and ligaments where severe osteoarthritis and ankylosis can occur. Mutations in the *ENPP1* gene have been associated with several rare human diseases, demonstrating the importance of NPP1 in maintaining normal tissue function [1,3,4,42]. There is a complex interaction between a wide range of molecular and genetic factors that inhibit calcification of the soft tissue and a breakdown in these pathways can lead to severe pathology. These genetic factors have been recently reviewed [43] therefore this review will focus on the roles of NPP1 in ectopic calcification in human disease, and the relevant rodent models used to study these pathological conditions.

The study of diseases such as GACI and pseudoxanthoma elasticum (PXE), which show overlapping clinical pathology in a wide range of

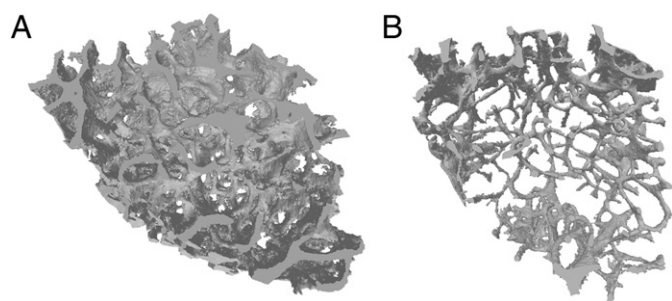


Fig. 2. Disruption of long bone mineralisation in *Enpp1*^{−/−} mice. Micro-computed tomography CT analysis of the femur of a (A) wild-type and (B) *Enpp1*^{−/−} mouse at 22 weeks of age. These reconstructions illustrate decreased trabecular bone mass in the *Enpp1*^{−/−} mice as reported in Mackenzie et al. [40].

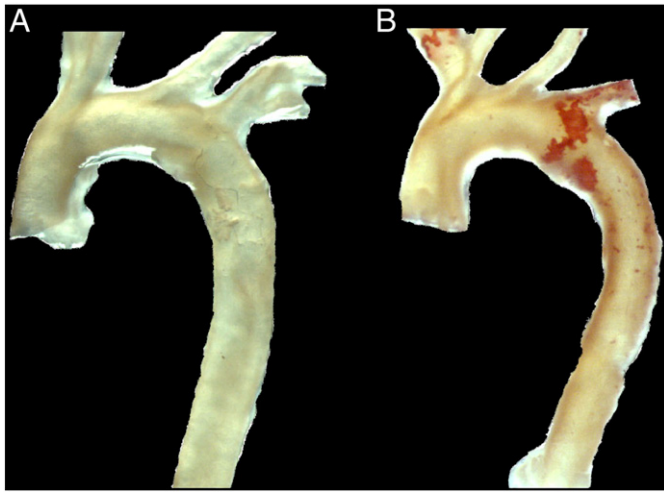


Fig. 3. Aortic calcification in *Enpp1*^{-/-} mice. Alizarin red staining of the aorta of a (A) wild-type and (B) *Enpp1*^{-/-} mouse at 22 weeks of age. Severe calcification of the aortic arch is observed in the *Enpp1*^{-/-} mouse.

tissues [42], has highlighted the extent of pathology caused by disrupted *ENPP1* expression.

Generalised arterial calcification of infancy and pseudoxanthoma elasticum: disease models of ectopic tissue calcification

Generalised arterial calcification of infancy (GACI) is a rare autosomal recessive disease characterised by calcification of large and medium-sized arteries and arterial stenosis caused by intimal proliferation (Fig. 4). Most affected children die within the first 6 months of life from the sequelae of end-organ damage including myocardial infarction [44]. In a subset of patients, peri-articular calcification of the greater joints also occurs. The finding of low systemic levels of inorganic pyrophosphate in one affected proband [45] due to defective activity of the PP_i-generating enzyme NPP1 [2] prompted the search for mutations in the NPP1 encoding gene, and indeed, most of the patients known so far with the classical GACI phenotype were found to carry bi-allelic mutations in *ENPP1* [46]. The understanding of the disease as caused by the deficiency of an inhibitor of hydroxyapatite crystal deposition, namely inorganic pyrophosphate, has paved the way for the use of bisphosphonates, i.e., synthetic analogues of PP_i to effectively treat GACI patients [46,47]. The retrospective observational analysis of 55 subjects affected by generalised arterial calcification of infancy by Rutsch and colleagues showed that treatment with bisphosphonates was associated with a regression of the calcifications and an increased survival rate [46]. However, spontaneous regression of ectopic calcifications also occurs in GACI patients [48,49]. Most

recently, mutations in *ENPP1* were also detected in a subset of patients with generalised arterial calcification and pseudoxanthoma elasticum: up to date, a total of four patients have been described, who presented typical signs of GACI in infancy and who later developed typical signs of PXE, including angioid streaks of the retina and pseudoxanthomatous skin lesions [42,50]. Pseudoxanthoma elasticum, a rare disease associated with soft tissue calcification at different sites including the eye, the kidneys, the arterial wall and the skin had been previously demonstrated to be caused by loss of function mutations in *ABCC6* encoding MRP6, a transport protein of hitherto unknown function [51,52]. Interestingly, *ABCC6* mutations have also been found to be associated with the GACI phenotype [42]. The finding of genocopy and phenocopy in GACI and PXE points to a close relationship between these two diseases and suggests common downstream mediators of ectopic tissue calcification in MRP6 and NPP1 deficiency [43].

Mouse models elucidating the role of NPP1 in tissue calcification

It has been widely described that mouse models with disrupted or genetically ablated *Enpp1* expression show high levels of ectopic calcification and subsequent cardiovascular pathology and hyperostosis of the joints [9,11,13–16,30,32,40]. Given that NPP1 is the primary producer of PP_i, an important inhibitor of HA crystallisation and chondrocyte differentiation [53], it is unsurprising that widespread soft tissue calcification is observed when NPP1 function is disrupted.

In the mutant mouse model, designated the *ttw/ttw* mouse, a phenotype including postnatal development of progressive ankylosing intervertebral and peripheral joint hyperostosis; increased vertebral cortical bone formation; spontaneous articular cartilage and arterial calcification is observed [9,11–14]. This mouse model provides a useful model for ossification of posterior lateral ligament (OPLL), a human condition characterised by pathological cartilage calcification in the spine and disrupted phosphate metabolism, associated with single nucleotide polymorphisms in the *ENPP1* gene [54–56].

A number of studies have demonstrated that *Enpp1*^{-/-} mice develop extensive arterial calcification (Fig. 3) [57]. The regulation of the phenotypic transition of VSMCs during aortic calcification is likely to involve reduced NPP1 activity and subsequent PP_i levels, with *Enpp1*^{-/-} VSMCs showing an up-regulation of molecules associated with chondrogenic, osteoblastic and osteocytic phenotypes [57]. Recent research has also demonstrated that NPP1 activity modulates arterial calcification through the mediation of receptor for advanced glycation of end-products (RAGE) signalling [58]. Membrane bound RAGE promotes nuclear factor-kappaB (NF-κB) and oxidative stress signalling, causing an up-regulation of aortic matrix remodelling. This signalling pathway has been implicated in patients suffering from aortic aneurysms and calcific aortic valve stenosis (CAVS) [59,60]. The production of sRAGE – a soluble endogenous suppressor of RAGE signalling – has been shown to be reduced in *Enpp1*^{-/-} aortic ring cultures. Additionally, treatment of cultures with sRAGE inhibits

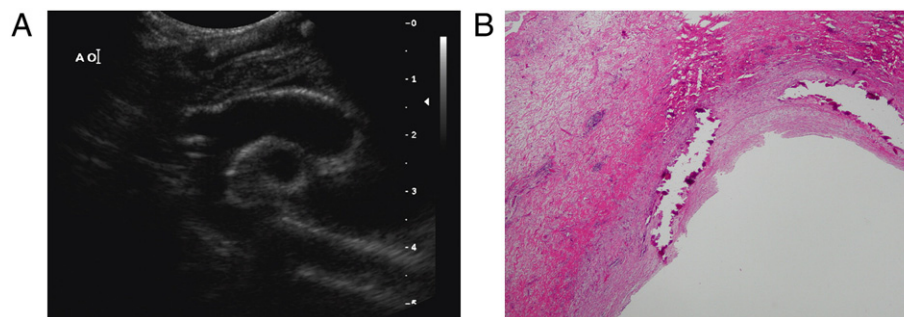


Fig. 4. Manifestations of generalised arterial calcification of infancy. Increased echogenicity of the calcified aortic arch in an infant carrying bi-allelic mutations in *ENPP1*, who died at the age of 8 days (ultrasonography, suprasternal view) (A). Calcification of the disrupted lamina elastica interna and intima proliferation of the aorta of another infant with GACI (haematoxylin-eosin, 10×) (B).

calcification and chondrogenic trans-differentiation [58]. Furthermore, the *Rage*^{-/-}/*Enpp1*^{-/-} double knockout mouse shows reduced arterial calcification when compared to the *Enpp1*^{-/-} mouse. It is, however, important to note that this double knockout mouse did not show a rescue of skeletal phenotype seen in *Enpp1*^{-/-} mice, suggesting that the changes in RAGE signalling mediated by loss of NPP1 activity may be specific to vascular smooth muscle cells.

The generation of PP_i by NPP1 also upregulates OPN expression, which can further inhibit mineralisation [61–64]. The complex interplay between OPN and NPP1 during ectopic calcification is confounded by the pro-atherogenic activity of OPN [65,66], and the recent finding that NPP1 promotes atherosclerotic plaque formation through OPN [20]. Furthermore, recent studies by Cote and colleagues have demonstrated that over-expression of *ENPP1* can also induce mineralisation in human valve interstitial cells [67]. The authors show not only that *ENPP1* expression is increased in human stenotic valve samples, but also that when over-expressed in vitro, NPP1 acts to increase apoptosis and mineralisation through a mechanism involving disrupted signalling of the P2Y2 and PI3-kinase/Akt pathways. These data indicate that expression of *ENPP1* must be maintained within a physiological range, and when altered, either by a reduction or increase in *ENPP1* expression, ectopic mineralisation may occur. Thus the precise role that NPP1 plays in modulating vascular calcification has yet to be fully defined, and requires further investigation.

Calcium phosphate homeostasis

The recent demonstration of elevated expression and circulating levels of fibroblast growth factor 23 (FGF-23) in *Enpp1*^{-/-} mice [40] is consistent with human genetic studies that have shown that mutations in *ENPP1* can cause hypophosphataemic rickets resulting from increased levels of FGF-23 [4]. These findings add to a growing number of single gene mutations whose activation impairs bone mineralisation and leads to changes in *Fgf-23* gene transcription [68]. As well as in *ENPP1*, mutations in other regulators of phosphate homeostasis, including phosphate regulating endopeptidase homolog, X-linked (*PHEX*) and dentin matrix protein-1 (*DMP1*), cause hypophosphatemic disorders and stimulate expression of FGF-23 [69,70]. This indicates that levels of bone metabolism and systemic phosphate homeostasis are tightly coordinated.

FGF-23 is a phosphaturic hormone that controls phosphate homeostasis, calcium homeostasis and bone mineralisation. FGF-23 binds to FGF receptors (mainly FGFR1) and the co-receptor KLOTHO in the kidney and promotes excretion of P_i, which leads to reduced serum P_i [71,72] and stimulation of Cyp24 and inhibition of Cyp27b1 in the kidney to reduce circulating 1,25(OH)₂D levels. Thus, the decreases in circulating calcium and phosphate levels reported in *Enpp1*^{-/-} mice are consistent with increased FGF-23 [40]. The mechanism whereby *Fgf-23* gene transcription in bone is stimulated by NPP1 inactivation has yet to be defined, however, recent studies have indicated that alterations in matrix mineralisation induced by other single gene mutations in osteoblasts lead to stimulation of *Fgf-23* expression via FGF receptor activation [73]. It is not clear whether the increase in FGF-23 observed in *Enpp1*^{-/-} bone is intrinsic and due to pathways similar to *PheX* and *Dmp1* mutations [69,70] or as a result of distinct signalling pathways. The increases in serum FGF-23 levels reported in *Enpp1*^{-/-} mice [40] may regulate the *Enpp1*^{-/-} bone phenotype through the bone–kidney axis or through local effects on bone cells. There is also controversial evidence that indicates that FGF-23 may directly affect skeletal mineralisation, independent of phosphate homeostasis [74], which further confounds the relationship between NPP1 and FGF-23 in *Enpp1*^{-/-} mice. Further research is required in order to fully elucidate the mechanisms through which NPP1 and FGF-23 are acting to modulate bone mineralisation.

Furthermore, the role of the FGF-23/KLOTHO axis in mediating vascular calcification is a subject of increasing interest. Although the

interaction between NPP1 and FGF-23 has not been investigated during vascular calcification it is interesting to note that there is an association between FGF-23 levels and calcium accumulation in the aorta and coronary arteries of patients with chronic kidney disease (CKD) [75–77]. Indeed, elevated FGF-23 levels in patients with CKD have also been associated with the presence of widespread atherosclerosis [78] and left ventricular hypertrophy [79,80]. High levels of ectopic calcification and disrupted bone structure have been described in *Fgf-23*^{-/-} mice [81,82], similar to the phenotype described in *Enpp1*^{-/-} mice. *Fgf-23* over-expressing mice also show a disrupted bone phenotype, with no ectopic calcification [83–85]. Recent evidence suggests that FGF-23 plays a protective role in vascular smooth muscle cells [86] but the precise actions of FGF-23, and its possible relationship with NPP1, during vascular calcification remain unclear and require further investigation.

Insulin signalling and glucose homeostasis

The link between NPP1 and insulin signalling was first described in a seminal study by Maddux and colleagues nearly two decades ago. NPP1 activity was shown to be increased in dermal fibroblast cultures from patients with non-insulin-dependent type 2 diabetes and severe insulin resistance [6]. Defective insulin-stimulated autophosphorylation of the insulin receptor (IR) was also observed in these cells, leading to the hypothesis that NPP1 acts as an inhibitor of the IR [87]. Subsequently, NPP1 has been shown to directly interact with the receptor α -subunit of the IR, blocking the insulin signalling pathway [88]. Additional studies in humans have also revealed that increased NPP1 expression in muscle correlates with increased body mass index and decreased insulin stimulation of muscle glucose transport [7,89], indicating a possible link between levels of NPP1 in muscle and insulin resistance.

Studies in animal models have shown that NPP1 regulates insulin signalling in both in vitro and in vivo settings. Transgenic mice with liver specific over-expression of human *ENPP1* show insulin resistance and glucose intolerance, although the animals are not overtly diabetic [90]. However, transgenic mice with human *ENPP1* over-expressed in both liver and muscle have fed and fasting hyperglycaemia with hyperinsulinaemia, suggesting that NPP1 may play a role in the insulin resistance and hyperglycaemia of type 2 diabetes. These findings have been further supported by murine studies demonstrating that in the presence of a high-fat diet, *Enpp1* over-expression in adipocytes induces fatty liver, hyperlipidaemia, and dysglycaemia, thus recapitulating key manifestations of the metabolic syndrome [91].

The majority of animal studies to date have focused on the effects on insulin signalling induced by over-expression of *Enpp1*. However a study by Zhou and colleagues [92] investigated the biological effect of NPP1 suppression. This research demonstrated that knockdown of *Enpp1* with siRNA significantly increases insulin-stimulated Akt phosphorylation in HuH7 human hepatoma cells. In vivo studies utilising the *db/db* mouse model of diabetes revealed that *db/db* mice treated with *Enpp1*-1 short hairpin RNA adenovirus showed reduced hepatic *Enpp1* mRNA levels and decreased fed and fasting plasma glucose, with a concomitant improved oral glucose tolerance. Taken together, these results demonstrate that suppression of *Enpp1* expression improves insulin sensitivity, supporting the proposition that NPP1 inhibition is a potential therapeutic approach for the treatment of type 2 diabetes.

Multiple linkage studies have associated the chromosome locus mapping *ENPP1*, to insulin resistance [90,93,94], hyperglyceridaemia [95], childhood and adult obesity and increased risk of type 2 diabetes [8]. Furthermore, specific polymorphisms have been identified, of which Lys121Gln (K121Q) [96] is the most widely investigated. Over-expression of the NPP1 Gln121 variant in vitro has been shown to have increased IR inhibition activity in cell lines representing the

liver (HepG2) and skeletal muscle (L6) when compared to the over-expression of the Lys121 variant [93]. This study showed that the Gln121 has a higher affinity to the IR, leading to a stronger inhibition of autophosphorylation. In the pancreatic B-cell line INS1E, over-expression of the Gln121 variant induced a significant increase in apoptosis, and almost abolished glucose induced insulin secretion, however the mechanism by which NPP1 mediates this reduction was not investigated. It is of particular interest that the over-expression of *ENPP1* alone, regardless of the variant, induced an 80% reduction in insulin secretion in INS1E cells, and a 20% and 50% decrease in IR autophosphorylation in HepG2 and L6 cells respectively [93].

Despite the existing evidence from *in vitro* studies on the increased susceptibility to insulin resistance of the Gln121 variant, there are now an increasing number of population association studies that show conflicting data about the linkage of this variant to insulin resistance, type 2 diabetes and obesity, which was extensively reviewed by Goldfine et al. [97]. Two recent studies have shown no association of Gln121 with type 2 diabetes in the Iranian and northern Chinese populations while previous studies on a Finnish population showed a strong linkage to early onset type 2 diabetes [98]. However, the largest Lys121Gln meta-analysis in type 2 diabetes to date, conducted on European populations, showed a modest increase of the Gln allele to risk of type 2 diabetes [99]. It is therefore likely that ethnic origin and environmental factors influence the development of type 2 diabetes, therefore confounding the role of NPP1 as a risk factor.

A fuller appreciation of the role of NPP1 in regulating insulin signalling and glucose homeostasis in newly defined metabolic tissues such as bone, as well as in established endocrine organs such as the pancreas and liver, is essential for the advancement of new potential strategies for the prevention and control of diabetes.

Conclusions

NPP1 is known to play vital roles in calcium/phosphate regulation, and repression of soft tissue mineralisation, as well as maintaining skeletal structure and function. A greater understanding of the actions of NPP1 in novel pathways such as insulin signalling in bone, in concurrence with the full elucidation of the mechanisms underpinning and connecting the known effects of NPP1, may stimulate the development of novel therapeutic treatments for patients with bone diseases, cardiovascular pathologies and diabetes.

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References

- [1] Rutsch F, Ruf N, Vaingankar S, Toliat MR, Suk A, Hohne W, et al. Mutations in *ENPP1* are associated with 'idiopathic' infantile arterial calcification. *Nat Genet* 2003;34:379–81.
- [2] Rutsch F, Vaingankar S, Johnson K, Goldfine I, Maddux B, Schauer P, et al. PC-1 nucleoside triphosphate pyrophosphohydrolase deficiency in idiopathic infantile arterial calcification. *Am J Pathol* 2001;158:543–54.
- [3] Levy-Litan V, Hershkovitz E, Avizov L, Leventhal N, Bercovich D, Chalifa-Caspi V, et al. Autosomal-recessive hypophosphatemic rickets is associated with an inactivation mutation in the *ENPP1* gene. *Am J Hum Genet* 2010;86:273–8.
- [4] Lorenz-Depiereux B, Schnabel D, Tiosano D, Hausler G, Strom TM. Loss-of-function *ENPP1* mutations cause both generalized arterial calcification of infancy and autosomal-recessive hypophosphatemic rickets. *Am J Hum Genet* 2010;86:267–72.
- [5] Ermakov S, Toliat MR, Cohen Z, Malkin I, Altmueller J, Livshits G, et al. Association of *ALPL* and *ENPP1* gene polymorphisms with bone strength related skeletal traits in a Chuvashian population. *Bone* 2010;46:1244–50.
- [6] Maddux BA, Sbraccia P, Kumakura S, Sasson S, Youngren J, Fisher A, et al. Membrane glycoprotein PC-1 and insulin-resistance in non-insulin-dependent diabetes-mellitus. *Nature* 1995;373:448–51.
- [7] Frittitta L, Youngren J, Vigneri R, Maddux BA, Trischitta V, Goldfine ID. PC-1 content in skeletal muscle of non-obese, non-diabetic subjects: relationship to insulin receptor tyrosine kinase and whole body insulin sensitivity. *Diabetologia* 1996;39:1190–5.
- [8] Meyre D, Bouatia-Naji N, Tounian A, Samson C, Lecoeur C, Vatin V, et al. Variants of *ENPP1* are associated with childhood and adult obesity and increase the risk of glucose intolerance and type 2 diabetes. *Nat Genet* 2005;37:863–7.
- [9] Sakamoto M, Hosoda Y, Kojimaha K, Yamazaki T, Yoshimura Y. Arthritis and ankylosis in TWY mice with hereditary multiple osteochondral lesions — with special reference to calcium deposition. *Pathol Int* 1994;44:420–7.
- [10] Terakado A, Tagawa M, Goto S, Yamazaki M, Moriya H, Fujimura S. Elevation of alkaline-phosphatase activity induced by parathyroid-hormone in osteoblast like cells from the spinal hyperostotic mouse TWY (twy/twy). *Calcif Tissue Int* 1995;56:135–9.
- [11] Baba H, Furusawa N, Fukuda M, Maezawa Y, Imura S, Kawahara N, et al. Potential role of streptozotocin in enhancing ossification of the posterior longitudinal ligament of the cervical spine in the hereditary spinal hyperostotic mouse (twy/twy). *Eur J Histochem* 1997;41:191–202.
- [12] Furusawa N, Baba H, Imura S, Fukuda M. Characteristics and mechanism of the ossification of posterior longitudinal ligament in the tip-toe walking Yoshimura (twy) mouse. *Eur J Histochem* 1996;40:199–210.
- [13] Okawa A, Nakamura I, Goto S, Moriya H, Nakamura Y, Ikegawa S. Mutation in *Npps* in a mouse model of ossification of the posterior longitudinal ligament of the spine. *Nat Genet* 1998;19:271–3.
- [14] Okawa A, Goto S, Moriya H. Calcitonin simultaneously regulates both periosteal hyperostosis and trabecular osteopenia in the spinal hyperostotic mouse (twy/twy) *in vivo*. *Calcif Tissue Int* 1999;64:239–47.
- [15] Sali A, Favaloro J, Terkeltaub R, Goding J. Germline deletion of the nucleoside triphosphate pyrophosphohydrolase (NTPPH) plasma cell membrane glycoprotein-1 (PC-1) produces abnormal calcification of periarticular tissues. In: Vanduffel L, Lemmings R, editors. *Ecto-ATPases and related ectoenzymes*. Shaker Publishing; 1999. p. 267–82.
- [16] Anderson HC, Harmey D, Camacho NP, Garimella R, Sipe JB, Tague S, et al. Sustained osteomalacia of long bones despite major improvement in other hypophosphatasia-related mineral deficits in tissue nonspecific alkaline phosphatase/nucleotide pyrophosphatase phosphodiesterase 1 double-deficient mice. *Am J Pathol* 2005;166:1711–20.
- [17] Buckley MF, Loveland KA, McKinstry WJ, Garson OM, Goding JW. Plasma-cell membrane glycoprotein PC-1 — cDNA cloning of the human molecule, amino acid sequence, and chromosomal location. *J Biol Chem* 1990;265:17506–11.
- [18] Funakoshi I, Kato H, Horie K, Yano T, Hori Y, Kobayashi H, et al. Molecular-cloning of cDNAs for human fibroblast nucleotide pyrophosphatase. *Arch Biochem Biophys* 1992;295:180–7.
- [19] Bollen M, Gijssels R, Ceulemans H, Stalmans W, Stefan C. Nucleotide pyrophosphatases/phosphodiesterases on the move. *Crit Rev Biochem Mol Biol* 2000;35:393–432.
- [20] Nitschke Y, Weissen-Plenz G, Terkeltaub R, Rutsch F. *Npp1* promotes atherosclerosis in ApoE knockout mice. *J Cell Mol Med* 2011;15:2273–83.
- [21] Terkeltaub RA. Inorganic pyrophosphate generation and disposition in pathophysiology. *Am J Physiol Cell Physiol* 2001;281:C1–C11.
- [22] Johnson K, Terkeltaub R. Inorganic pyrophosphate (PP_i) in pathologic calcification of articular cartilage. *Front Biosci* 2005;10:988–97.
- [23] Goding JW, Terkeltaub R, Maurice M, Deterre P, Sali A, Belli SL. Ecto-phosphodiesterase/pyrophosphatase of lymphocytes and non-lymphoid cells: structure and function of the PC-1 family. *Immunol Rev* 1998;161:11–26.
- [24] Stefan C, Jansen S, Bollen M. Modulation of purinergic signaling by NPP-type ectophosphodiesterases. *Purinergic Signal* 2006;2:361–70.
- [25] Clair T, Lee HY, Liotta LA, Stracke ML. Autotaxin is an exoenzyme possessing 5'-nucleotide phosphodiesterase/ATP pyrophosphatase and ATPase activities. *J Biol Chem* 1997;272:996–1001.
- [26] Uriarte M, Stalmans W, Hickman S, Bollen M. Phosphorylation and nucleotide-dependent dephosphorylation of hepatic polypeptides related to the plasma-cell differentiation antigen PC-1. *Biochem J* 1993;293:93–100.
- [27] Landt M, Butler LG. 5'-Nucleotide phosphodiesterase — isolation of covalently bound 5'-monophosphate, an intermediate in catalytic mechanism. *Biochemistry* 1978;17:4130–5.
- [28] Anderson HC. Matrix vesicles and calcification. *Curr Rheumatol Rep* 2003;5:222–6.
- [29] Johnson KA, Hesse L, Vaingankar S, Wennberg C, Mauro S, Narisawa S, et al. Osteoblast tissue-nonspecific alkaline phosphatase antagonizes and regulates PC-1. *Am J Physiol Regul Integr Comp Physiol* 2000;279:R1365–77.
- [30] Johnson K, Goding J, Van Etten D, Sali A, Hu SI, Farley D, et al. Linked deficiencies in extracellular PP_i and osteopontin mediate pathologic calcification associated with defective PC-1 and ANK expression. *J Bone Miner Res* 2003;18:994–1004.
- [31] Hesse L, Johnson KA, Anderson HC, Narisawa S, Sali A, Goding JW, et al. Tissue-nonspecific alkaline phosphatase and plasma cell membrane glycoprotein-1 are central antagonistic regulators of bone mineralization. *Proc Natl Acad Sci U S A* 2002;99:9445–9.
- [32] Harmey D, Hesse L, Narisawa S, Johnson KA, Terkeltaub R, Millan JL. Concerted regulation of inorganic pyrophosphate and osteopontin by Atp2, Enpp1, and Ank — an integrated model of the pathogenesis of mineralization disorders. *Am J Pathol* 2004;164:1199–209.
- [33] Harmey D, Johnson KA, Zelken J, Camacho NP, Hoylaerts MF, Noda M, et al. Elevated skeletal osteopontin levels contribute to the hypophosphatasia phenotype in Atp2(−/−) mice. *J Bone Miner Res* 2006;21:1377–86.
- [34] Sugita A, Kawai S, Hayashibara T, Amano A, Ooshima T, Michigami T, et al. Cellular ATP synthesis mediated by type III sodium-dependent phosphate transporter Pit-1 is critical to chondrogenesis. *J Biol Chem* 2011;286:3094–103.
- [35] Yadav MC, Simao AMS, Narisawa S, Huesa C, McKee MD, Farquharson C, et al. Loss of skeletal mineralization by the simultaneous ablation of PHOSPHO1 and alkaline

- phosphatase function: a unified model of the mechanisms of initiation of skeletal calcification. *J Bone Miner Res* 2011;26:286–97.
- [36] Terkeltaub R, Rosenbach M, Fong F, Goding J. Causal link between nucleotide pyrophosphohydrolase overactivity and increased intracellular inorganic pyrophosphate generation demonstrated by transfection of cultured fibroblasts and osteoblasts with plasma-cell membrane glycoprotein-1 – relevance to calcium pyrophosphate dihydrate deposition disease. *Arthritis Rheum* 1994;37:934–41.
- [37] Ho AM, Johnson MD, Kingsley DM. Role of the mouse ank gene in control of tissue calcification and arthritis. *Science* 2000;289:265–70.
- [38] Hakim FT, Cranley R, Brown KS, Eanes ED, Harne L, Oppenheim JJ. Hereditary joint disorder in progressive ankylosis (ank/ank) mice. I. Association of calcium hydroxyapatite deposition with inflammatory arthropathy. *Arthritis Rheum* 1984;27:1411–20.
- [39] Addison WN, Azari F, Sorensen ES, Kaartinen MT, McKee MD. Pyrophosphate inhibits mineralization of osteoblast cultures by binding to mineral, up-regulating osteopontin, and inhibiting alkaline phosphatase activity. *J Biol Chem* 2007;282:15872–83.
- [40] Mackenzie NCW, Zhu D, Milne EM, van 't Hof R, Martin A, Quarles DL, et al. Altered bone development and an increase in FGF-23 expression in *Enpp1*^{−/−} mice. *PLoS One* 2012;7:e32177.
- [41] Nam HK, Liu J, Li Y, Kragor A, Hatch NE. Ectonucleotide pyrophosphatase/phosphodiesterase-1 (ENPP1) protein regulates osteoblast differentiation. *J Biol Chem* 2011;286:39059–71.
- [42] Nitschke Y, Baujat G, Botschen U, Wittkamp T, du Moulin M, Stella J, et al. Generalized arterial calcification of infancy and pseudoxanthoma elasticum can be caused by mutations in either *ENPP1* or *ABCC6*. *Am J Hum Genet* 2012;90:25–39.
- [43] Rutsch F, Nitschke Y, Terkeltaub R. Genetics in arterial calcification: pieces of a puzzle and cogs in a wheel. *Circ Res* 2011;109:578–92.
- [44] Moran JJ. Idiopathic arterial calcification of infancy: a clinicopathologic study. *Pathol Annu* 1975;10:393–417.
- [45] Rutsch F, Schuete P, Kalhoff H, Petrarulo M, August C, Diekmann L. Low levels of urinary inorganic pyrophosphate indicating systemic pyrophosphate deficiency in a boy with idiopathic infantile arterial calcification. *Acta Paediatr* 2000;89:1265–9.
- [46] Rutsch F, Boeyer P, Nitschke Y, Ruf N, Lorenz-Depierreux B, Wittkamp T, et al. Hypophosphatemia, hyperphosphaturia, and bisphosphonate treatment are associated with survival beyond infancy in generalized arterial calcification of infancy. *Circ Cardiovasc Genet* 2008;1:133–40.
- [47] Ramjan KA, Roscioli T, Rutsch F, Silience D, Munns CF. Generalized arterial calcification of infancy: treatment with bisphosphonates. *Nat Clin Pract Endocrinol Metab* 2009;5:167–72.
- [48] Sholler GF, Yu JS, Bale PM, Hawker RE, Celermajer JM, Kozlowski K. Generalized arterial calcification of infancy – 3 case reports, including spontaneous regression with long-term survival. *J Pediatr* 1984;105:257–60.
- [49] Ciana G, Trappan A, Bambi B, Benettoni A, Maso G, Zennaro F, et al. Generalized arterial calcification of infancy: two siblings with prolonged survival. *Eur J Pediatr* 2006;165:258–63.
- [50] Li Q, Schumacher W, Jablonski D, Siegel D, Uitto J. Cutaneous features of pseudoxanthoma elasticum in a patient with generalized arterial calcification of infancy due to a homozygous missense mutation in the *ENPP1* gene. *Br J Dermatol* 2012;166:1107–11.
- [51] Le Saux O, Urban Z, Tschuch C, Csiszar K, Bacchelli B, Quaglini D, et al. Mutations in a gene encoding an ABC transporter cause pseudoxanthoma elasticum. *Nat Genet* 2000;25:223–7.
- [52] Ringpfeil F, Lebwohl MG, Christiano AM, Uitto J. Pseudoxanthoma elasticum: mutations in the *MRP6* gene encoding a transmembrane ATP-binding cassette (ABC) transporter. *Proc Natl Acad Sci U S A* 2000;97:6001–6.
- [53] Johnson K, Polewski M, van Etten D, Terkeltaub R. Chondrogenesis mediated by PP_i depletion promotes spontaneous aortic calcification in *NPP1*^{−/−} mice. *Arterioscler Thromb Vasc Biol* 2005;25:686–91.
- [54] Nakamura I, Ikegawa S, Okawa A, Okuda S, Koshizuka Y, Kawaguchi H, et al. Association of the human *NPPS* gene with ossification of the posterior longitudinal ligament of the spine (OPLL). *Hum Genet* 1999;104:492–7.
- [55] Koshizuka Y, Kawaguchi H, Ogata N, Ikeda T, Mabuchi A, Seichi A, et al. Nucleotide pyrophosphatase gene polymorphism associated with ossification of the posterior longitudinal ligament of the spine. *J Bone Miner Res* 2002;17:138–44.
- [56] Tahara M, Aiba A, Yamazaki M, Ikeda Y, Goto S, Moriya H, et al. The extent of ossification of posterior longitudinal ligament of the spine associated with nucleotide pyrophosphatase gene and leptin receptor gene polymorphisms. *Spine* 2005;30:877–80.
- [57] Zhu D, Mackenzie NCW, Millan JL, Farquharson C, MacRae VE. The appearance and modulation of osteocyte marker expression during calcification of vascular smooth muscle cells. *PLoS One* 2011;6:e19595.
- [58] Cecil DL, Terkeltaub RA. Arterial calcification is driven by RAGE in *Enpp1*^{−/−} mice. *J Vasc Res* 2011;48:227–35.
- [59] Bowman MH, Wilk J, Heydemann A, Kim G, Rehman J, Lodato JA, et al. *S100A12* mediates aortic wall remodeling and aortic aneurysm. *Circ Res* 2010;106:145–U291.
- [60] Basta G, Corci A, Vianello A, Del Turco S, Foffa I, Navarra T, et al. Circulating soluble receptor for advanced glycation end-product levels are decreased in patients with calcific aortic valve stenosis. *Atherosclerosis* 2010;210:614–8.
- [61] Hunter GK, Kyle CL, Goldberg HA. Modulation of crystal-formation by bone phosphoproteins – structural specificity of the osteopontin-mediated inhibition of hydroxyapatite formation. *Biochem J* 1994;300:723–8.
- [62] Boskey AL, Maresca M, Ullrich W, Doty SB, Butler WT, Prince CW. Osteopontin-hydroxyapatite interactions in-vitro – inhibition of hydroxyapatite formation and growth in a gelatin-gel. *Bone Miner* 1993;22:147–59.
- [63] Speer MY, McKee MD, Guldberg RE, Liaw L, Yang HY, Tung E, et al. Inactivation of the osteopontin gene enhances vascular calcification of matrix Gla protein-deficient mice: evidence for osteopontin as an inducible inhibitor of vascular calcification in vivo. *J Exp Med* 2002;196:1047–55.
- [64] Steitz SA, Speer MY, McKee MD, Liaw L, Almeida M, Yang H, et al. Osteopontin inhibits mineral deposition and promotes regression of ectopic calcification. *Am J Pathol* 2002;161:2035–46.
- [65] Matsui Y, Rittling SR, Okamoto H, Inobe M, Jia N, Shimizu T, et al. Osteopontin deficiency attenuates atherosclerosis in female apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 2003;23:1029–34.
- [66] Bruemmer D, Collins AR, Noh G, Wang W, Territo M, Arias-Magallona S, et al. Angiotensin II-accelerated atherosclerosis and aneurysm formation is attenuated in osteopontin-deficient mice. *J Clin Invest* 2003;112:1318–31.
- [67] Cote N, El Hussein D, Pepin A, Bosse Y, Bouvet C, Audet A, et al. ATP release acts as survival signal and prevents the mineralization of aortic valve by P2Y(2) and PI3K/AKT signaling. *Circulation* 2011;124:A11911.
- [68] Quarles LD. Endocrine functions of bone in mineral metabolism regulation. *J Clin Invest* 2008;118:3820–8.
- [69] Liu S, Zhou J, Tang W, Menard R, Feng JQ, Quarles LD. Pathogenic role of *Fgf23* in *Dmp1*-null mice. *Am J Physiol Endocrinol Metab* 2008;295:E254–61.
- [70] Liu SG, Zhou JP, Tang W, Jiang X, Rowe DW, Quarles LD. Pathogenic role of *Fgf23* in *Hyp* mice. *Am J Physiol Endocrinol Metab* 2006;291:E38–49.
- [71] Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, et al. The parathyroid is a target organ for *FGF23* in rats. *J Clin Invest* 2007;117:4003–8.
- [72] Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, et al. Regulation of fibroblast growth factor-23 signaling by Klotho. *J Biol Chem* 2006;281:6120–3.
- [73] Martin A, Liu S, David V, Li H, Karydis A, Feng JQ, et al. Bone proteins PHEX and *DMP1* regulate fibroblastic growth factor *Fgf23* expression in osteocytes through a common pathway involving FGF receptor (FGFR) signaling. *FASEB J* 2011;25:2551–62.
- [74] Sitara D, Kim S, Razzaque MS, Bergwitz C, Taguchi T, Schuler C, et al. Genetic evidence of serum phosphate-independent functions of *FGF-23* on bone. *PLoS Genet* 2008;4:10.
- [75] Srivaths PR, Goldstein SL, Silverstein DM, Krishnamurthy R, Brewer ED. Elevated *FGF-23* and phosphorus are associated with coronary calcification in hemodialysis patients. *Pediatr Nephrol* 2011;26:945–51.
- [76] Nasrallah MM, El-Shehaby AR, Salem MM, Osman NA, El Sheikh E, El Din UAAS. Fibroblast growth factor-23 (*FGF-23*) is independently correlated to aortic calcification in haemodialysis patients. *Nephrol Dial Transplant* 2010;25:2679–85.
- [77] Mirza MAI, Larsson A, Lind L, Larsson TE. Circulating fibroblast growth factor-23 is associated with vascular dysfunction in the community. *Atherosclerosis* 2009;205:385–90.
- [78] Mirza MAI, Hansen T, Johansson L, Ahlstrom H, Larsson A, Lind L, et al. Relationship between circulating *FGF23* and total body atherosclerosis in the community. *Nephrol Dial Transplant* 2009;24:3125–31.
- [79] Mirza MAI, Larsson A, Melhus H, Lind L, Larsson TE. Serum intact *FGF23* associate with left ventricular mass, hypertrophy and geometry in an elderly population. *Atherosclerosis* 2009;207:546–51.
- [80] Gutierrez OM, Januzzi JL, Isakova T, Laliberte K, Smith K, Collierone G, et al. Fibroblast growth factor 23 and left ventricular hypertrophy in chronic kidney disease. *Circulation* 2009;119:2545–52.
- [81] Sitara D, Razzaque MS, Hesse M, Yoganathan S, Taguchi T, Erben RG, et al. Homozygous ablation of fibroblast growth factor-23 results in hyperphosphatemia and impaired skeletogenesis, and reverses hypophosphatemia in *PheX*-deficient mice. *Matrix Biol* 2004;23:421–32.
- [82] Stubbs JR, Liu S, Tang W, Zhou J, Wang Y, Yao X, et al. Role of hyperphosphatemia and 1,25-dihydroxyvitamin D in vascular calcification and mortality in fibroblastic growth factor 23 null mice. *J Am Soc Nephrol* 2007;18:2116–24.
- [83] Bai XY, Miao DS, Li JR, Goltzman D, Karaplis AC. Transgenic mice overexpressing human fibroblast growth factor 23 (*R176Q*) delineate a putative role for parathyroid hormone in renal phosphate wasting disorders. *Endocrinology* 2004;145:5269–79.
- [84] Shimada T, Urakawa I, Yamazaki Y, Hasegawa H, Hino R, Yoneya T, et al. *FGF-23* transgenic mice demonstrate hypophosphatemic rickets with reduced expression of sodium phosphate cotransporter type IIa. *Biochem Biophys Res Commun* 2004;314:409–14.
- [85] Larsson T, Marsell R, Schipani E, Ohlsson C, Ljunggren O, Tenenhouse HS, et al. Transgenic mice expressing fibroblast growth factor 23 under the control of the $\alpha 1(I)$ collagen promoter exhibit growth retardation, osteomalacia, and disturbed phosphate homeostasis. *Endocrinology* 2004;145:3087–94.
- [86] Lim KLT, Molostvov G, Lee C, Lam F, Zehnder D, Hsiao LL. Vascular Klotho deficiency potentiates the development of human artery calcification and mediates resistance to *FGF-23*. *Circulation* 2012;125(18):2243–55.
- [87] Frittitta L, Spampinato D, Solini A, Nosadini R, Goldfine ID, Vigneri R, et al. Elevated *PC-1* content in cultured skin fibroblasts correlates with decreased in vivo and in vitro insulin action in nondiabetic subjects – evidence that *PC-1* may be an intrinsic factor in impaired insulin receptor signaling. *Diabetes* 1998;47:1095–100.
- [88] Maddux BA, Goldfine ID. Membrane glycoprotein *PC-1* inhibition of insulin receptor function occurs via direct interaction with the receptor α -subunit. *Diabetes* 2000;49:13–9.
- [89] Youngren JF, Maddux BA, Sasson S, Sbraccia P, Tapscott EB, Swanson MS, et al. Skeletal muscle content of membrane glycoprotein *PC-1* in obesity – relationship to muscle glucose transport. *Diabetes* 1996;45:1324–8.
- [90] Maddux BA, Chang YN, Accili D, McGuinness OP, Youngren JF, Goldfine ID. Overexpression of the insulin receptor inhibitor *PC-1/ENPP1* induces insulin resistance and hyperglycemia. *Am J Physiol Endocrinol Metab* 2006;290:E746–9.
- [91] Pan W, Ciociola E, Saraf M, Tumurbaatar B, Tuvdendorj D, Prasad S, et al. Metabolic consequences of *ENPP1* overexpression in adipose tissue. *Am J Physiol Endocrinol Metab* 2011;301:E901–11.
- [92] Zhou HH, Chin C-N, Wu M, Ni W, Quan S, Liu F, et al. Suppression of *PC-1/ENPP-1* expression improves insulin sensitivity in vitro and in vivo. *Eur J Pharmacol* 2009;616:346–52.

- [93] Di Paola R, Caporarello N, Marucci A, Dimatteo C, Iadicicco C, Del Guerra S, et al. ENPP1 affects insulin action and secretion: evidences from in vitro studies. *PLoS One* 2011;6:e19462.
- [94] Goldfine ID, Maddux BA, Youngren JF, Frittitta L, Trischitta V, Dohm GL. Membrane glycoprotein PC-1 and insulin resistance. *Mol Cell Biochem* 1998;182:177–84.
- [95] Tanyolaç S, Bremer A.A., Hodoglugil U., Movsesyan I., Pullinger C.R., Heiner S.W., Malloy M.J., Kane J.P., Goldfine I.D. Genetic variants of the *ENPP1/PC-1* gene are associated with hypertriglyceridemia in male subjects. 2009;7: 543–548.
- [96] Pizzuti A, Frittitta L, Argiolas A, Baratta R, Goldfine ID, Bozzali M, et al. A polymorphism (K121Q) of the human glycoprotein PC-1 gene coding region is strongly associated with insulin resistance. *Diabetes* 1999;48:1881–4.
- [97] Goldfine ID, Maddux BA, Youngren JF, Reaven G, Accili D, Trischitta V, et al. The role of membrane glycoprotein plasma cell antigen 1 ectonucleotide pyrophosphatase phosphodiesterase 1 in the pathogenesis of insulin resistance and related abnormalities. *Endocr Rev* 2008;29:62–75.
- [98] Willer CJ, Bonnycastle LL, Conneely KN, Duren WL, Jackson AU, Scott LJ, et al. Screening of 134 single nucleotide polymorphisms (SNPs) previously associated with type 2 diabetes replicates association with 12 SNPs in nine genes. *Diabetes* 2007;56:256–64.
- [99] McAteer JB, Prudente S, Bacci S, Lyon HN, Hirschhorn JN, Trischitta V, et al. The EBPP1 K121Q polymorphism is associated with type 2 diabetes in European populations: evidence from an updated meta-analysis in 42,042 subjects. *Diabetes* 2008;57:1125–30.