



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Expression profiling reveals differences in immuno-inflammatory gene expression between the two disease forms of sheep paratuberculosis

Citation for published version:

Smeed, JA, Watkins, CA, Gossner, AG & Hopkins, J 2010, 'Expression profiling reveals differences in immuno-inflammatory gene expression between the two disease forms of sheep paratuberculosis', *Veterinary Immunology and Immunopathology*, vol. 135, no. 3-4, pp. 218-25.
<https://doi.org/10.1016/j.vetimm.2009.11.015>

Digital Object Identifier (DOI):

[10.1016/j.vetimm.2009.11.015](https://doi.org/10.1016/j.vetimm.2009.11.015)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Veterinary Immunology and Immunopathology

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Elsevier Editorial System(tm) for Veterinary Immunology and Immunopathology
Manuscript Draft

Manuscript Number: VETIMM-D-09-1576R1

Title: Expression profiling reveals differences in immuno-inflammatory gene expression between the two disease forms of sheep paratuberculosis

Article Type: Research Paper - Full length

Keywords: Mycobacterium avium subsp. paratuberculosis
MAP
Sheep
Chemokines
Genomics
Pathology.

Corresponding Author: Professor J. Hopkins,

Corresponding Author's Institution: University of Edinburgh, Royal (Dick) School of Vet. Studies

First Author: Jennifer A Smeed, BSc. PhD.

Order of Authors: Jennifer A Smeed, BSc. PhD. ; Craig A Watkins, BSc. PhD; Anton G Gossner, BSc. PhD; J. Hopkins



The Editor
Veterinary Immunology & Immunopathology

Professor John Hopkins
THE ROSLIN INSTITUTE
R(D)SVS
University of Edinburgh
Summerhall
Edinburgh, EH9 1QH
UK

Telephone +44 (0)131 650 6169
Fax +44 (0)131 650 6511
john.hopkins@ed.ac.uk
www.roslin.ed.ac.uk

26 Oct 2009

Ms. No. VETIMM-D-09-1576

Veterinary Immunology and Immunopathology

Many thanks to the referees for positive and informative comments. We have modified the manuscript in accordance with these comments.

Reviewer #1.

1. The title has now been changed to better reflect the content of the paper.
2. We have amended the Abstract to make unambiguous that we have measured transcripts and not protein.
3. Clinical signs clarified etc. Have explained and referenced IS900
4. We have included descriptions of results for chemokines/receptors/TCR molecules in the Results. I have reformatted Table 1 so that the genes are in strict alphabetic order; it is not possible to arrange them in comparison order as there would be some duplication.
5. The point of the RT-qPCR is validation of the array results – and this it does (section 3.2). The rationale for their selection is now explained in section 3.2.
6. IGFBP6 was represented in the array (see supplementary data Array Express) – but in these experiments it did not show differential expression of >1.5 and $p \leq 0.05$ in any comparison. As referenced in the manuscript we had previously identified it in a preliminary series of experiments (Roupaka MSc thesis 2004).
7. There was strong correlation between array and RT-qPCR (section 3.2) but difference in absolute values – this is a common finding and comments/references etc have now been added to the Discussion.
8. MMP9 and TYROBP were not discussed specifically because the purpose of the paper was not to cover all genes exhaustively, but to focus on areas of particular interest. If required I can add a paragraph to the discussion – but it will not add much.

Minor comments.... Asterisks explained in Table 1. SDHA/YWHAZ defined.

Reviewer #2.

1. See comment 1 and 2 (Reviewer #1).
2. Abbreviations now defined.
3. Further detailed choice of housekeeping genes. The two used were selected from six; including ribosomal protein L18, ATPase, GAPDH, β actin as well as SDHA and YWHAZ. I can list these in the text if you wish – but it is not very relevant to the paper.
4. MIQE guidelines followed for abbreviations.
5. Statistics further explained, end section 2.5.
6. References added and clarified. Rather than adding several references for well known facts I have included a reference for an informative review (Campbell *et al.* 2003).
7. Fig 1 legend better described.

Very many thanks,

A handwritten signature in black ink, appearing to read 'Jonathan P. Lewis'. The signature is stylized and cursive, with the first name 'Jonathan' and the last name 'Lewis' clearly visible.

1 **Expression profiling reveals differences in immuno-inflammatory**
2 **gene expression between the two disease forms of sheep**
3 **paratuberculosis**

4
5 J. A. Smeed^{a,c}, C. A. Watkins^{b,c}, A. G. Gossner^a, J. Hopkins^{a,*}

6
7 ^a*The Roslin Institute, Royal (Dick) School of Veterinary Studies, University of Edinburgh, Summerhall,*
8 *Edinburgh, EH9 1QH, UK.*

9 ^b*Moredun Research Institute, International Research Centre, Pentlands Science Park, Penicuik,*
10 *Midlothian EH26 0PZ, UK.*

11
12 ^c Equal contribution

13
14
15 *Corresponding author at: The Roslin Institute & Royal (Dick) School of Veterinary Studies,
16 University of Edinburgh, Summerhall, Edinburgh, EH9 1QH, UK. Tel. +44 131 650 6169;
17 Fax. +44 131 650 6511.
18 *E-mail address:* john.hopkins@ed.ac.uk

19
20
21
22
23
24
25

1 **Abstract**

2 Paratuberculosis is a chronic enteropathy of ruminants caused by *Mycobacterium avium*
3 subspecies *paratuberculosis* (MAP); infection of sheep results in two disease forms –
4 paucibacillary (tuberculoid) and multibacillary (lepromatous) associated with the differential
5 polarization of the immune response. In addition the majority of MAP-infected animals show
6 no pathology and remain asymptomatic. Microarray and real-time RT-qPCR analyses were
7 used to compare gene expression in ileum from sheep with the two disease forms and
8 asymptomatic sheep, to further understand the molecular basis of the pathologies.
9 Microarrays identified 36 genes with fold-change of >1.5 and $P \leq 0.05$ in at least one
10 comparison; eight candidates were chosen for RT-qPCR validation. Sequence analysis of two
11 candidates, CXCR4 and IGFBP6, identified three SNPs in each; five were found in all three
12 forms of disease and showed no significant relationship to pathological type. The IGFBP6 G
13 ³⁷⁴³ A SNP was not detected in asymptomatic sheep. The data show that the two forms of
14 disease are associated with distinct molecular profiles highlighted by the differential
15 expression of chemokine and chemokine receptor transcripts, the protein products of which
16 might be implicated in the different cell infiltrates of the pathologies. The cells within the
17 lesions also show evidence of abnormal activation; they express high levels of cytokine
18 transcripts but have reduced expression levels of transcripts for T cell receptor associated
19 molecules.

20

21 *Keywords:*

22 *Mycobacterium avium* subsp. *paratuberculosis*

23 MAP

24 Sheep

25 Chemokines

26 Genomics

27 Pathology

28

1 **1. Introduction**

2

3 *Mycobacterium avium* subspecies *paratuberculosis* (MAP) is a facultative
4 intracellular bacterium that primarily infects macrophages and is the causative agent of
5 paratuberculosis (Johne's disease), a common enteropathy of ruminants. In cattle an extended
6 preclinical phase is dominated by a type 1 immune response that is followed by progression to
7 fatal granulomatous disease, which is linked to a switch to a type 2 response and the
8 production of non-protective IgG1 antibodies (Koets et al., 2002; Kurade et al., 2004). In
9 sheep both type 1 and type 2 responses give rise to disease states (Clarke, 1997) and there
10 seems to be no progression from the paucibacillary (type 1 or tuberculoid) to the
11 multibacillary (type 2 or lepromatous) disease; both are fatal. As with homologous
12 pathologies in tuberculosis and leprosy, the cytokines that dominate paucibacillary
13 paratuberculosis are IFN γ and IL-12, while IL-10 dominates multibacillary disease (Smeed *et*
14 *al.*, 2007). Only a minority of infected (IS900+) sheep progress to disease and the majority
15 remain asymptomatic with no pathology (Koets et al., 2002; Smeed et al., 2007; de Silva et
16 al., 2009).

17 We hypothesize that the polarization of the immune response is critical to the clinical
18 outcome of paratuberculosis infection. Intestinal tissue damage that results from a type 1
19 response involves lymphocyte and eosinophil infiltration and is fundamentally different to
20 that caused by a type 2 response, which leads to epithelioid macrophage infiltration and
21 dissemination of infection (Smeed *et al.*, 2007). The related disease leprosy also has this
22 dichotomy of immunology and pathology (Meisner *et al.*, 2001). Furthermore there is a clear
23 relationship between host genetics and disease type (Fortin *et al.*, 2007); many genes have
24 been implicated but one that has received much attention is solute carrier family 11
25 (SLC11A1, NRAMP-1 or Bcg), which is linked to both tuberculosis susceptibility (Vidal *et*
26 *al.*, 1993) and discrimination of the two forms of human leprosy (Meisner *et al.*, 2001).

1 Resistance to MAP infection in mice is also partly linked to this gene, with animals carrying
2 the *Bcg^f* allele (e.g. C3H) being refractory to infection (Veazey *et al.*, 1995).

3 The principal cell populations associated with mycobacterial infection are cells of the
4 macrophage lineage, a major function of which is the activation of T cells and initiation of the
5 immune response. Antigen-presenting cell subsets play a crucial role in the polarization of
6 responses through the differential expression of IL-10 and IL-12 (Pulendran *et al.*, 1999).

7 They also show differential expression of pattern recognition receptors (PRRs) (Nalubamba *et*
8 *al.*, 2008), and differential PRR activation can tailor the immune response (Kapsenberg,
9 2003). Both TLR2 and CARD15 (NOD2) have been shown to be up-regulated in clinical
10 paratuberculosis (Nalubamba *et al.*, 2008) and involved in both recognition (Ferwerda *et al.*,
11 2007) and response (Weiss *et al.*, 2001) to MAP. Indeed, there is growing evidence for
12 linkage of TLR2 and/or CARD15 mutations and susceptibility to human mycobacterial
13 diseases (Bochud *et al.*, 2003; Ben-Ali *et al.*, 2004) and bovine MAP infection (Mucha *et al.*,
14 2009).

15 To begin to unravel the complex interactions between MAP and the immune system
16 that gives rise to bovine paratuberculosis, Coussens and colleagues have used bovine
17 leukocyte microarrays to examine transcript changes in peripheral blood mononuclear cells
18 (PBMC) (Coussens *et al.*, 2002) and macrophages (Murphy *et al.*, 2006). In addition they
19 have examined cattle ileal tissue (Aho *et al.*, 2003) in order to understand the mechanisms
20 that lead to intestinal tissue damage and have identified a number of genes involved in the
21 immunology and pathogenesis of bovine paratuberculosis (including insulin-like growth
22 factor binding protein-6 - IGFBP6). Our own studies (Smeed *et al.*, 2007; Nalubamba *et al.*,
23 2008) have focussed on ovine paratuberculosis because of the defined pathologies in sheep
24 and exploit an oligonucleotide microarray of ~600 immuno-inflammatory genes (Watkins *et*
25 *al.*, 2008) to gain insight into the role of these genes in the development of the different
26 pathologies. The defined nature of the disease phenotype in sheep facilitates the identification
27 of genetic markers associated with pathological type, and might also inform studies in cattle.

28

1 Competitive hybridization of cDNA isolated from the ileum (the site of
2 paratuberculosis lesions) between the three infected groups was performed to identify changes
3 to transcript expression associated with each disease group and develop molecular signatures
4 of each pathological form. Quantitative real-time RT-PCR (RT-qPCR) on selected candidate
5 genes was used to validate the arrays and the data were analysed to identify and compare
6 physiological pathways associated with the contrasting pathologies. Data sets gathered from
7 such experiments provide the potential to follow pathways of immune reactivity as well as
8 assessing disease states that can then be related to specific gene expression signatures. In
9 addition, as the epidemiology of these mycobacterial diseases suggests a genetic susceptibility
10 (Ben-Ali *et al.*, 2004) we investigated this link, by a preliminary single nucleotide
11 polymorphism (SNP) analysis, between polymorphisms in two of the candidates and
12 pathological form.

16 **2. Materials and Methods**

17 *2.1. Experimental animals and tissues*

18 MAP infected animals presented with clinical Johne's disease (diagnosed by a
19 veterinary surgeon: prolonged weight loss, inappetance and depression, and occasional
20 diarrhoea) were out bred, female sheep (Table S1) of a variety of breeds and ages. All sheep
21 were euthanized and diagnosis was confirmed by histopathology, Ziehl Neelsen (ZN) staining
22 and IS900 real-time quantitative PCR with a sensitivity of 5 genome equivalents (Eishi *et al.*,
23 2002); sheep of similar ages from the same flocks (Nalubamba *et al.*, 2008), with no signs of
24 clinical Johne's disease and positive for IS900 DNA were considered asymptomatic (Smeed
25 *et al.*, 2007). IS900 is a specific marker for the precise identification of MAP (Green *et al.*,
26 1989). Animal procedures were performed under a valid Animals (Scientific Procedures) Act
27 1986 Project Licence.

28

1 2.2. *RNA isolation*

2 Terminal ileum sections (~0.5 g) were placed in five volumes of *RNAlater* (Qiagen,
3 Crawley, UK), incubated overnight at 4°C and stored at -80°C. Tissues were homogenized in
4 guanidine isothiocyanate lysis buffer (RNeasy, Qiagen) and RNA was isolated using the
5 RNeasy Maxi kit, eluted in 0.8 ml of RNase-free water, precipitated with ethanol and
6 resuspended in 0.3 ml of RNase-free water. Samples were DNase I digested (Qiagen) and
7 genomic DNA contamination assessed by GAPDH PCR and a no reverse transcriptase
8 control. RNA quality was assessed using a RNA 6000 Nano LabChip on the Agilent 2100
9 bioanalyzer and quantified using a NanoDrop ND-1000 spectrophotometer; all samples had
10 an RNA integrity number >7; n = 6 for the microarray experiments.

11

12 2.3. *Preparation of labelled cDNA and microarray hybridization*

13 cDNA generation and indirect labelling was performed using the Fairplay III
14 Microarray Labelling Kit (Stratagene, Cambridge, UK) incorporating Cy3 and Cy5 dyes (GE
15 Healthcare, Amersham, UK); the DyeEx spin 2.0 kit (Qiagen) removed unincorporated dye.
16 Pre-hybridization and hybridization was carried out using the Pronto!TM Kit (Corning,
17 Loughborough, UK). This analysis utilized the ruminant immuno-inflammatory gene
18 universal array (Watkins *et al.*, 2008); 4,824 spots from 596 genes. A 22x22 mm lifterslip
19 (VWR, VWR, Poole, UK) was then placed over the array and the slide pre-heated to 42°C.
20 Equal dye concentrations from two samples (30 pmols each) were mixed, dried and
21 resuspended in 20 µl cDNA hybridization solution and introduced onto the array. The slides
22 were incubated for 16 - 20 h at 42°C with agitation every 7 s, using a SlideBooster (Implen,
23 Stansted Mountfitchet, UK) and post-hybridization washes of the microarray slides carried
24 out using the Pronto!TM Kit, using an AdvaWash AW400 slide washer (Implen). The slides
25 were scanned using a GenePix Autoloader 4200AL, controlled by GenePix Pro v 6.1
26 (Molecular Devices, Sunnyvale, CA). In total, we performed 18 microarrays. All groups
27 contained six animals and every sample was analysed on three arrays, with each animal being

1 compared with an animal in each of the other groups. The identity of the pairings between
2 each group was chosen at random.

3 Protocols of the experimental procedures, methods of analysis and microarray data
4 are available as supplementary information in the European Bioinformatics Institute's
5 ArrayExpress database (www.ebi.ac.uk/arrayexpress). Login to www.ebi.ac.uk/aerep/login
6 and query for the accession number of the experiment or array design E-TABM-487.

7

8 2.4. *Microarray data analysis*

9 Scanned images were aligned to the GenePix Array List (GAL file); background
10 noise subtracted; spot intensity amplitudes generated and quality data extracted using
11 BlueFuse v3.3 (BlueGnome, Cambridge, UK). The log₂ ratios from the two channels for each
12 spot were calculated by Bluefuse. Data generated by Bluefuse were pre-processed using
13 Limma within BioConductor 2.2 (<http://www.bioconductor.org>). Normalization was carried
14 out using a weighted print-tip Loess method. Spots were weighted by a confidence value
15 provided by BlueFuse. Differentially expressed genes from the different group comparisons
16 (Table 1) were classified based on biological process using GO annotation. GORetriever,
17 GOanna and GoSlimViewer (<http://www.agbase.msstate.edu>) were used to annotate gene
18 products based on sequence similarity.

19

20 2.5. *Real time RT-qPCR validation*

21 RT-qPCR was performed using a Corbett Rotor-Gene™ 3000 (Qiagen). First
22 strand cDNA synthesis used 0.5 µg of total RNA and Oligo(dT) and Superscript™ III RT
23 (Invitrogen, Paisley, UK), from all RNA samples used for microarrays with a further two
24 samples from each of the groups (n = 8). Primers for candidate genes were designed (Table
25 S2) and the amplicons cloned using pGEM-T Easy kit (Promega, Southampton, UK). PCR
26 conditions were optimized using a standard curve generated from linearized plasmid. All
27 serial dilutions were generated using a CAS-1200™ Precision Liquid Handling System

1 (Qiagen). Succinate dehydrogenase complex, subunit A (SDHA) and tyrosine 3/tryptophan 5
2 -monooxygenase activation protein, zeta (YWHAZ); were chosen from a group of six using
3 GeNorm v3.4 (Vandesompele *et al.*, 2002) and NormFinder v 0.953 (Andersen *et al.*, 2004),
4 as the two most stably expressed genes to normalize the RT-qPCR assays. All reactions were
5 10 µl volumes containing 4 µl of cDNA at the optimum dilution (1:20), 5 µl 2x FastStart
6 SYBR Green Master mix (Roche, Lewes, UK) and 0.5 µl of each primer at optimum
7 concentration (Table S2). PCR conditions were: 94°C for 5 min and 45 cycles of 94°C for 20
8 s, 60°C for 20 s, 72°C for 20 s, followed by dissociation curve analysis to confirm a single
9 gene product. Each run assayed one gene and contained all samples in duplicate, a cDNA
10 standard curve and a non-template control. This was repeated twice using cDNA from two
11 additional RT reactions, resulting in 6 data points for each gene. The average Cq value for
12 each sample was calculated and converted into relative quantities, taking account of the gene
13 specific efficiency of the reaction and multiple reference gene normalization, using qBase
14 (Hellemans *et al.*, 2007). Statistical analyses were performed on data from individual animals
15 using Kolmogorov-Smirnov to test for normality of distribution and two-tailed Student's t-test
16 for each of the comparisons.

17

18 2.6. *PCR and DNA sequencing*

19 DNA was extracted from 250 mg ileum using the Qiagen DNEasy Blood and Tissue
20 kit or Qbiogene Fast DNA kit and quantified using the Nanodrop ND-1000 spectrometer. Full
21 length mRNA sequence was obtained for sheep CXCR4 and IGFBP6, and aligned to the
22 bovine genome to identify the coding regions in the genomic sequence. Primers (Table S3)
23 were designed to amplify the whole bovine coding region and the intron / exon boundaries
24 where possible (Fig. S1). Three PCR reactions were performed with each set of primers on
25 ten animals in each infected group (Table S1). The PCR products from each sample were
26 mixed, purified, ligated and cloned into the PGEM-T easy. Three independent sequences were
27 produced for each PCR product and consensus sequences produced using VectorNTI AlignX

1 (Invitrogen). Differences between the genomic sequences were considered to be SNPs when
2 the same change was found consistently in two or more of the consensus sequences. The
3 distribution of alleles and genotypes was compared between the pathological groups using the
4 χ^2 -test.

5
6 **3. Results**

7 *3.1. Microarray analysis*

8 Of the 596 genes analysed, 36 fulfilled the criteria of fold-change of ≥ 1.5 and $P \leq 0.05$
9 in at least one comparison (Table 1 and supplementary information). The fold-changes shown
10 are means of all probes for each gene. Intrinsic variation within replicate spots on single
11 arrays (technical) and between arrays (biological) was determined. Technical variation was
12 determined by comparing normalized \log_2 ratios of the spots printed in triplicate within each
13 array, for six replicate array experiments of each comparison. The median standard deviation
14 for the 3 comparisons varied from 0.24 (for multi- verses paucibacillary (M v P) to 0.26 (for
15 multi- verses asymptomatic (M v A) with a variance of between 0.06 and 0.07 across the three
16 comparisons. The biological variation was analysed by comparing the \log_2 ratios (CH1:CH2)
17 of identical spots between the 6 replicate arrays in each comparison. The median standard
18 deviation for each comparison ranged from 0.37 to 0.42, with a variance between 0.14 and
19 0.18 across all comparisons, demonstrating that within chip variability (technical variation)
20 was between third and a half of the between chip variation (biological variation).

21 ITGAL was the only gene significantly affected in all three comparisons (Table 1,
22 Fig. 1) and two other genes (SELL and TFRC) were up-regulated but $P \geq 0.05$. Five genes
23 were differentially expressed between the two disease groups and the asymptomatic group
24 (Table 1). Fifteen genes were significantly different in the paucibacillary vs asymptomatic
25 comparison including a 1.52 fold repression of CCL5 and a 2.25 fold increase in CXCL10.
26 Twenty three were significantly different in the multibacillary vs asymptomatic comparison
27 including a 1.64 fold increase in CCL2; 1.54 and 1.82 fold increases in CXCR3 and CXCR4
28 and 1.59 and 1.67 decreases in CD3E and CD3F. The pauci- vs multibacillary comparison

1 identified fifteen affected genes, but only four (C9, CLIC5, ITGAM and MMP7) were unique
2 to that comparison, as the other comparative data was not significant (<1.5 and $p>0.05$).
3 Within the limitations of this focussed microarray, it is clear that the differentially expressed
4 genes associated with each of the paratuberculosis pathologies were connected with a distinct
5 range of biological processes (Fig. 2). Both clinical diseases showed changes to genes linked
6 to responses to biotic stimuli, cell differentiation and signal transduction but only in the
7 multibacillary form were there changes to genes associated with cell communication.

8

9 3.2. *RT-qPCR validation of microarray results*

10 Of the 36 genes found to be differentially expressed in the microarray experiment,
11 seven candidates were chosen for RT-qPCR validation of the array; CD63, CXCR4, IGF2R,
12 ITGB2, MMP9, TLR2, and TYROBP; on the basis that they represent different biological
13 processes (Fig. 2) and were differentially expressed between pauci- and multibacillary
14 samples. In addition we identified IGF2R as it had been highlighted as being differentially
15 expressed in bovine paratuberculosis (Coussens *et al.*, 2004) and in a preliminary array
16 analysis (Roupaka, 2004). Table 2 shows the results expressed as relative fold-changes in the
17 same comparisons as performed in the microarray. The RT-qPCR analyses for seven original
18 candidate genes validated the results from the microarray experiment with a correlation
19 coefficient (r) 0.63 – 0.97, mean of 0.87. Comparison (two-tailed Students t test) of
20 expression levels of all eight candidates between Blackface x Blue du Maine vs other breeds
21 in the paucibacillary group and Blackface x Blue du Maine vs other breeds in the
22 asymptomatic group showed that breed had no significant impact ($P > 0.8$) on the distribution
23 of the data. Similar comparisons of the 4-5 year old vs the 2-3 year old Blackface sheep in the
24 multibacillary group also showed that age also had no effect on the data ($P > 0.9$).

25

26 3.2. *Single nucleotide polymorphism (SNP) analysis*

27 Analysis of the consensus CXCR4 sequences revealed the presence of three
28 synonymous SNPs; in exon 2 a C to T change at position 2080 and a G to A change at 2275

1 and in exon 3 a G to A change at 3361; the last was 3' of the stop codon. Analysis of IGFBP6
2 sequence also revealed three SNPs – a non-synonymous C to A change at position 12 in exon
3 1, which changes a histidine to a glutamine; a synonymous G to A change at 2730 in intron 3
4 and a G to A change at 3743 in exon 4, 3' of the stop codon at position 3612 (Fig. S1). When
5 these six SNPs were analysed in relation to infection there was no significant relationship
6 between any SNP and disease form (Table 3). Each of the three CXCR4 SNPs and two
7 IGFBP6 SNPs were found in all infected groups. However, IGFBP6 SNP3 (G 3822 A) was
8 only found in clinically diseased animals, and was absent from the asymptomatic group. The
9 animals tested were of a variety of breeds and crosses (Table S1) but there was no
10 discernable, significant relationship between breed and any genotype (χ^2 , $P \geq 0.79$).

11

12

13 **4. Discussion**

14 This is the first microarray analysis of ileal gene expression in sheep with clinical
15 paratuberculosis and directly compares gene expression at the site of infection in each of the
16 pathologies, and has identified novel pathways involved in MAP pathogenesis. Previous work
17 on sheep paratuberculosis has shown that the pauci- and multibacillary pathologies are
18 associated type 1 and type 2 T cell responses respectively, with the differential activation of T
19 cells (Burrells *et al.*, 1998) and the expression of quite distinct panels of both innate receptors
20 (Nalubamba *et al.*, 2008) and cytokines (Smeed *et al.*, 2007). Analysis of the data for the
21 eight candidate genes indicates that it is highly unlikely that these changes are associated with
22 the breed or age (Table SI). Paratuberculosis is characterized by extensive inflammation and
23 cellular infiltration of the terminal ileum; paucibacillary disease is associated with
24 lymphocytes and eosinophils, and multibacillary lesions with epithelioid macrophages
25 (Smeed *et al.*, 2007). These variations in the cellular composition are reflected in the
26 differences in gene expression profiles detected by microarray, with 36 genes found to be
27 differentially expressed (>1.5 fold and $P \leq 0.05$) in at least one comparison (Table 1). The RT-

1 qPCR data justify our decision to use 1.5 fold (and $p \leq 0.05$) as the cut-off point for our
2 candidate gene selection as they confirmed the array data in relation to the direction of fold
3 change. In addition, fold changes of just under 1.5 (but $p \leq 0.05$) were shown to be
4 significantly different by RT-qPCR, e.g. MMP9. RT-qPCR also confirmed the array data,
5 both quantitatively and qualitatively, when microarray showed no change (>-1.5 to <1.5), e.g.
6 CD63 in the pauci- vs asymptomatic comparison. The correlation coefficient (r) between
7 array and RT-qPCR varied between candidate genes (0.63 – 0.97, mean $r = 0.87$) and was
8 similar to that described in other systems (Dallas et al., 2005), but was much lower for genes
9 with a fold change of <1.5 . The magnitude of fold change with RT-qPCR was generally much
10 higher than with microarray, which is probably due to the different sensitivities of the two
11 assays (Morey et al., 2006); nevertheless RT-qPCR is the preferred validation method for
12 microarrays (Dallas et al., 2005).

13

14 Within the limitations of the focussed microarray, it is clear that the differentially
15 expressed genes associated with each of the pathologies were connected with a distinct range
16 of biological processes (Fig. 2) probably associated with the different cell infiltrates in the
17 two disease forms. The multibacillary samples displayed a consistent repression of the several
18 genes encoding elements of the T cell receptor complex - CD3E, CD3G (and CD3Z with –
19 1.47 fold change and $P \leq 0.05$). This might be related to the fact that the principal cells in
20 multibacillary ileum were infected epithelioid macrophages and not T cells. It is possible that
21 this reduction in antigen-receptor expression is a result of infection-associated aberrant
22 activation and may represent a specific bacterial mechanism for the modification of the host
23 immune response towards the development of disease as is seen in tuberculosis (Smith, 2003).
24 Related to this were changes to transcripts of the different immunoglobulin molecules.
25 IGHA1 and IGHM were raised in both disease forms and IGJ was increased in paucibacillary
26 lesions. In view of the fact that B cell receptor associated genes were not raised in infected
27 tissues, these increases in immunoglobulin heavy chain transcripts may be a result of greater
28 numbers of plasma cells rather than mature B cells. This pattern of immunoglobulin heavy

1 chain expression does not fit exactly into the Th1/Th2 paradigm, which would predict high
2 levels of IGHA only in the multibacillary (lepromatous) lesions (Mosmann and Coffman,
3 1989).

4 Representatives of the β 1 and β 2 integrins (ITGA4 and ITGAL) were up-regulated in
5 the paucibacillary forms. However, their behaviour was less consistent in the multibacillary
6 form where the β 1 integrin ITGA4 is repressed and the β 2 integrins ITGAL, ITGAM and
7 ITGB2 were increased. The four transmembrane domain proteins (e.g. CD63) interact with
8 integrins and modulate their function; and the high levels of CD63 in multi- relative to
9 paucibacillary lesions may be indicative of different cell populations associated with those
10 lesions. The major cellular ligand for the β 2 integrins is ICAM1. This, and the lymphocyte
11 adhesion molecule, SELL (CD62L or L selectin), were high in multibacillary diseased tissues,
12 which reflects major cellular infiltration and also inflammation, as both are up-regulated by
13 inflammatory cytokines (Dustin and Springer, 1991).

14 The stimulus for leukocyte infiltration into lesions is probably different for each
15 disease form. CCL2 was raised in multibacillary lesions (and 1.65 fold but $P \geq 0.05$ in the
16 pauci- vs asymptomatic comparison) and was probably produced by a variety of cells on
17 stimulation with pro-inflammatory cytokines and IFN γ . It is chemotactic for lymphocytes and
18 monocytes but not eosinophils and its role in inflammation is as an activator of neutrophils
19 (Campbell et al., 2003). CCL5 is associated with T cell and dendritic cell recruitment, but it
20 was repressed in the paucibacillary group. CCL5 is a product of activated T cells, but only at
21 the late stage of activation, and is down-regulated by TGF β and IFN γ both of which were
22 increased in paucibacillary tissues (Smeed *et al.*, 2007). This is additional evidence for the
23 aberrant activation of T cells in the paratuberculosis lesions. The chemokine CXCL10 is
24 associated with chemoattraction of type 1 T cells (Campbell *et al.*, 2003) and was
25 significantly raised in paucibacillary lesions and might explain the T cell infiltration.
26 However its receptor, CXCR3 was up-regulated in multibacillary tissues, probably due to the
27 effect of high levels of TGF β . The closely related chemokine receptor, CXCR4 is also

1 responsive to TGF β and was significantly increased in multibacillary tissues. This is the
2 receptor for CXCL12 (SDF-1) and one of its major functions is as a chemoattractant for Th2
3 and regulatory T cells (Campbell *et al.*, 2003).

4 IGF2R, the receptor for IGF2 and mannose-6-phosphate, was increased in the
5 multibacillary disease form and plays a role in the activation of TGF β (Melnick *et al.*, 1998),
6 a gene previously shown to be up-regulated in these samples (Smeed *et al.*, 2007). IGFBP6
7 was differentially expressed in sheep paratuberculosis (increased in paucibacillary and
8 repressed in multibacillary diseased ileum) and it was also up-regulated in cattle PBMC
9 stimulated by MAP (Coussens *et al.*, 2004). It binds with high affinity to IGF2 and inhibits
10 many of its functions, including proliferation and differentiation (Kelley *et al.*, 1996). Its
11 expression has previously been reported to be stimulated by IL-1 β and TNF α but inhibited by
12 the presence of TGF β (Martin *et al.*, 1994; Liu *et al.*, 1999), and this might explain the
13 significantly increased levels of IGFBP6 in paucibacillary lesions. In paucibacillary tissues,
14 high IGFBP6 levels could play a role in controlling cellular proliferation by inhibiting IGF2
15 and inducing apoptosis. TLR2 was greatly up-regulated in multibacillary lesions, suggesting
16 that the TLR2 signalling pathway is important in lepromatous pathogenesis. TLR2 is central
17 to the immune response to mycobacteria, as knockout mice are more susceptible to infections.
18 In human leprosy TLR2 is linked to differential pathology, as signalling through the mutated
19 receptor leads to increased IL-10 production and a type 2 immune response (Kang *et al.*,
20 2004). Consistent with this are increased levels of IL-10 in multibacillary tissues and raised
21 IFN γ in paucibacillary lesions (Smeed *et al.*, 2007). Multibacillary tissues show high levels of
22 both CXCR4 and TLR2 and, interestingly, cross-talk between these has been reported in
23 *Porphyromonas gingivalis* infection that leads to the undermining of host defence
24 (Hajishengallis *et al.*, 2008). The data on the linkage between TLR2 and MAP in ruminants is
25 not clear. Analysis of TLR2 SNPs in sheep paratuberculosis failed to identify any linkage
26 with pathological form (Nalubamba *et al.*, 2008) although in cattle there was significant
27 association of a distinct SNP with MAP infection (Mucha *et al.*, 2009).

1 Due to their differential expression in multibacillary and paucibacillary sheep,
2 CXCR4 and IGFBP6 were selected for preliminary SNP analysis and to assess whether these
3 SNPs were associated with gene expression and pathology. All SNPs except IGFBP6 at 3743
4 were found in the three infected groups and there was no significant association with any
5 disease form. The IGFBP6 SNP at 3743 is 3' to the stop codon and was not found in
6 asymptomatic sheep; there was no statistically significant association with disease form
7 possibly because of the small numbers of animals (n=10) in each group.

8 To summarize, this study identifies novel genes and pathways linked to mycobacterial
9 pathologies. The two disease forms of sheep paratuberculosis are associated with distinct
10 molecular profiles highlighted by the differential expression of chemokines, which are
11 implicated in the different cellular infiltrates of the pathologies and the polarization of the
12 immune response. Cells within the lesions show evidence of abnormal activation, with high
13 levels of cytokine transcripts and a reduction in expression levels of transcripts for
14 lymphocyte receptor molecules. Preliminary analysis could identify no significant link
15 between CXCR4 and IGFBP6 SNPs and paratuberculosis pathologies.

16
17

18 **Acknowledgements**

19 We acknowledge Jill Sales (BioSS) for help with experimental and statistical design
20 and the clinical staff at the Moredun Research Institute. This project was funded by the
21 BBSRC and SEERAD (Project Grants 15/S19602 and 15/S13964). JAS was funded by
22 BBSRC/Genesis-Faraday CASE studentship and sponsored by Moredun Scientific Ltd.
23 Midlothian, UK.

24
25
26
27
28

1 **References**

- 2 Aho, A.D., McNulty, A.M., Coussens, P.M., 2003. Enhanced expression of interleukin-1 α
3 and tumor necrosis factor receptor-associated protein 1 in ileal tissues of cattle
4 infected with *Mycobacterium avium* subsp. paratuberculosis. *Infect. Immun.* 71,
5 6486.
- 6 Andersen, C.L., Jensen, J.L., Orntoft, T.F., 2004. Normalization of real-time quantitative
7 reverse transcription-PCR data: a model-based variance estimation approach to
8 identify genes suited for normalization, applied to bladder and colon cancer data sets.
9 *Cancer Res* 64, 5245-5250.
- 10 Ben-Ali, M., Barbouche, M.R., Bousnina, S., Chabbou, A., Dellagi, K., 2004. Toll-like
11 receptor 2 Arg677Trp polymorphism is associated with susceptibility to tuberculosis
12 in Tunisian patients. *Clin Diagn Lab Immunol* 11, 625-626.
- 13 Bochud, P.Y., Hawn, T.R., Aderem, A., 2003. Cutting edge: a Toll-like receptor 2
14 polymorphism that is associated with lepromatous leprosy is unable to mediate
15 mycobacterial signaling. *J Immunol* 170, 3451-3454.
- 16 Burrells, C., Clarke, C.J., Colston, A., Kay, J.A., Porter, J., Little, D., Sharp, J.M., 1998. A
17 study of immunological responses of sheep clinically-affected with paratuberculosis
18 (Johne's disease): The relationship of blood, mesenteric lymph node and intestinal
19 lymphocyte responses to gross and microscopic pathology. *Veterinary Immunology*
20 and *Immunopathology* 66, 343-358.
- 21 Campbell, D.J., Kim, C.H., Butcher, E.C., 2003. Chemokines in the systemic organization of
22 immunity. *Immunological Reviews* 195, 58-71.
- 23 Clarke, C.J., 1997. The pathology and pathogenesis of paratuberculosis in ruminants and
24 other species. *J Comp Pathol* 116, 217-261.

1 Coussens, P.M., Colvin, C.J., Wiersma, K., Abouzied, A., Sipkovsky, S., 2002. Gene
2 expression profiling of peripheral blood mononuclear cells from cattle infected with
3 *Mycobacterium paratuberculosis*. *Infect. Immun.* 70, 5494-5502.

4 Coussens, P.M., Jeffers, A., Colvin, C.J., 2004. Rapid and transient activation of gene
5 expression in peripheral blood mononuclear cells from Johne's disease positive cows
6 exposed to *Mycobacterium paratuberculosis* in vitro. *Microbial Pathogenesis* 36, 93-
7 108.

8 Dallas, P.B., Gottardo, N.G., Firth, M.J., Beesley, A.H., Hoffmann, K., Terry, P.A., Freitas,
9 J.R., Boag, J.M., Cummings, A.J., Kees, U.R., 2005. Gene expression levels assessed
10 by oligonucleotide microarray analysis and quantitative real-time RT-PCR -- how
11 well do they correlate? *BMC Genomics* 6, 59.

12 de Silva, K., Begg, D., Carter, N., Taylor, D., Di Fiore, L., Whittington, R., 2009. The early
13 lymphocyte proliferation response in sheep exposed to *Mycobacterium avium* subsp.
14 *paratuberculosis* compared to infection status. *Immunobiology*.

15 Dustin, M.L., Springer, T.A., 1991. Role of lymphocyte adhesion receptors in transient
16 interactions and cell locomotion. *Annu Rev Immunol* 9, 27-66.

17 Eishi, Y., Suga, M., Ishige, I., Kobayashi, D., Yamada, T., Takemura, T., Takizawa, T.,
18 Koike, M., Kudoh, S., Costabel, U., Guzman, J., Rizzato, G., Gambacorta, M., du
19 Bois, R., Nicholson, A.G., Sharma, O.P., Ando, M., 2002. Quantitative analysis of
20 mycobacterial and propionibacterial DNA in lymph nodes of Japanese and European
21 patients with sarcoidosis. *J Clin Microbiol* 40, 198-204.

22 Ferwerda, G., Kullberg, B.J., de Jong, D.J., Girardin, S.E., Langenberg, D.M.L., van Crevel,
23 R., Ottenhoff, T.H.M., Van der Meer, J.W.M., Netea, M.G., 2007. *Mycobacterium*
24 *paratuberculosis* is recognized by Toll-like receptors and NOD2. *J Leukoc Biol* 82,
25 1011-1018.

- 1 Fortin, A., Abel, L., Casanova, J.L., Gros, P., 2007. Host Genetics of Mycobacterial Diseases
2 in Mice and Men: Forward Genetic Studies of BCG-osis and Tuberculosis. Annual
3 Review of Genomics and Human Genetics 8, 163-192.
- 4 Green, E.P., Tizard, M.L., Moss, M.T., Thompson, J., Winterbourne, D.J., McFadden, J.J.,
5 Hermon-Taylor, J., 1989. Sequence and characteristics of IS900, an insertion element
6 identified in a human Crohn's disease isolate of Mycobacterium paratuberculosis.
7 Nucleic Acids Res 17, 9063-9073.
- 8 Hajishengallis, G., Wang, M., Liang, S., Triantafilou, M., Triantafilou, K., 2008. Pathogen
9 induction of CXCR4/TLR2 cross-talk impairs host defense function. Proc Natl Acad
10 Sci U S A 105, 13532-13537.
- 11 Hellemans, J., Mortier, G., De Paepe, A., Speleman, F., Vandesompele, J., 2007. qBase
12 relative quantification framework and software for management and automated
13 analysis of real-time quantitative PCR data. Genome Biol 8, R19.
- 14 Kang, T.J., Yeum, C.E., Kim, B.C., You, E.Y., Chae, G.T., 2004. Differential production of
15 interleukin-10 and interleukin-12 in mononuclear cells from leprosy patients with a
16 Toll-like receptor 2 mutation. Immunology 112, 674-680.
- 17 Kapsenberg, M.L., 2003. Dendritic cell control of pathogen-driven T-cell polarization. Nat
18 Rev Immunol 3, 984-993.
- 19 Kelley, K.M., Oh, Y., Gargosky, S.E., Gucev, Z., Matsumoto, T., Hwa, V., Ng, L., Simpson,
20 D.M., Rosenfeld, R.G., 1996. Insulin-like growth factor-binding proteins (IGFBPs)
21 and their regulatory dynamics. The International Journal of Biochemistry & Cell
22 Biology 28, 619-637.
- 23 Koets, A., Rutten, V., Hoek, A., van Mil, F., Müller, K., Bakker, D., Gruys, E., van Eden, W.,
24 2002. Progressive bovine paratuberculosis is associated with local loss of CD4+ T

- 1 cells, increased frequency of $\gamma\delta$ T cells, and related changes in T-cell function. *Infect.*
2 *Immun.* 70, 3856-3864.
- 3 Kurade, N.P., Tripathis, B.N., Rajukumar, K., Parihar, N.S., 2004. Sequential development of
4 histologic lesions and their relationship with bacterial isolation, faecal shedding and
5 immune responses during progressive stages of experimental infection of lambs with
6 *Mycobacterium avium* subsp. *paratuberculosis*. *Vet Pathol* 41, 378-387.
- 7 Liu, Y., Tsushima, T., Miyakawa, M., Isozaki, O., Yamada, H., Xu, Z.R., Iwamoto, Y., 1999.
8 Effect of cytokines on production of insulin-like growth factor binding proteins
9 (IGFBPs) from human fibroblasts in culture. *Endocr J* 46 Suppl, S63-S66.
- 10 Martin, J.L., Coverley, J.A., Baxter, R.C., 1994. Regulation of immunoreactive insulin-like
11 growth factor binding protein-6 in normal and transformed human fibroblasts. *J Biol*
12 *Chem* 269, 11470-11477.
- 13 Meisner, S.J., Mucklow, S., Warner, G., Sow, S.O., Lienhardt, C., Hill, A.V., 2001.
14 Association of NRAMP1 polymorphism with leprosy type but not susceptibility to
15 leprosy per se in west Africans. *Am J Trop Med Hyg* 65, 733-735.
- 16 Melnick, M., Chen, H., Buckley, S., Warburton, D., Jaskoll, T., 1998. Insulin-like growth
17 factor II receptor, transforming growth factor-beta, and Cdk4 expression and the
18 developmental epigenetics of mouse palate morphogenesis and dysmorphogenesis.
19 *Dev Dyn* 211, 11-25.
- 20 Morey, J.S., Ryan, J.C., Van Dolah, F.M., 2006. Microarray validation: factors influencing
21 correlation between oligonucleotide microarrays and real-time PCR. *Biol Proced*
22 *Online* 8, 175-193.
- 23 Mosmann, T.R., Coffman, R.L., 1989. TH1 and TH2 cells: different patterns of lymphokine
24 secretion lead to different functional properties. *Annu Rev Immunol* 7, 145-173.

- 1 Mucha, R., Bhide, M.R., Chakurkar, E.B., Novak, M., Mikula, S., 2009. Toll-like receptors
2 TLR1, TLR2 and TLR4 gene mutations and natural resistance to *Mycobacterium*
3 *avium* subsp. *paratuberculosis* infection in cattle. *Veterinary Immunology and*
4 *Immunopathology* 128, 381-388.
- 5 Murphy, J.T., Sommer, S., Kabara, E.A., Verman, N., Kuelbs, M.A., Saama, P., Halgren, R.,
6 Coussens, P.M., 2006. Gene Expression Profiling of Monocyte-Derived Macrophages
7 Following Infection with *Mycobacterium avium* subspecies *avium* and
8 *Mycobacterium avium* subspecies *paratuberculosis*. *Physiol Genomics* 28, 67-75.
- 9 Nalubamba, K.S., Smeed, J.A., Gossner, A.G., Watkins, C.A., Dalziel, R.G., Hopkins, J.,
10 2008. Differential expression of pattern recognition receptors in the three pathological
11 forms of sheep paratuberculosis. *Microbes and Infection* 10, 598-604.
- 12 Pulendran, B., Smith, J.L., Caspary, G., Brasel, K., Pettit, D., Maraskovsky, E., Maliszewski,
13 C.R., 1999. Distinct dendritic cell subsets differentially regulate the class of immune
14 response in vivo. *Proc Natl Acad Sci U S A* 96, 1036-1041.
- 15 Roupaka, S., 2004. Functional genomics of Johne's disease. MSc Thesis University of
16 Edinburgh.
- 17 Smeed, J.A., Watkins, C.A., Rhind, S.M., Hopkins, J., 2007. Differential cytokine gene
18 expression profiles in the three pathological forms of sheep paratuberculosis. *BMC*
19 *Vet Res* 3, 18.
- 20 Smith, I., 2003. *Mycobacterium tuberculosis* pathogenesis and molecular determinants of
21 virulence. *Clin Microbiol Rev* 16, 463-496.
- 22 Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., Speleman,
23 F., 2002. Accurate normalization of real-time quantitative RT-PCR data by geometric

1 averaging of multiple internal control genes. Genome Biology 3, research0034.1-
2 research0034.11.

3 Veazey, R.S., Horohov, D.W., Krahenbuhl, J.L., Taylor, H.W., Oliver, J.L., Snider, T.G.,
4 1995. Comparison of the resistance of C57BL/6 and C3H/He mice to infection with
5 *Mycobacterium paratuberculosis*. Vet Microbiol 47, 79-87.

6 Vidal, S.M., Malo, D., Vogan, K., Skamene, E., Gros, P., 1993. Natural resistance to infection
7 with intracellular parasites: isolation of a candidate for Bcg. Cell 73, 469-485.

8 Watkins, C.A., McKellar, A., Jensen, K., George, A., Jones, D., Sharp, J.M., Stevenson, K.,
9 Hopkins, J., 2008. Development and validation of an oligonucleotide microarray for
10 immuno-inflammatory genes of ruminants. Veterinary Research Communications 32,
11 647-657.

12 Weiss, D.J., Evanson, O.A., McClenahan, D.J., Abrahamsen, M.S., Walcheck, B.K., 2001.
13 Regulation of expression of major histocompatibility antigens by bovine
14 macrophages infected with *Mycobacterium avium* subsp. *paratuberculosis* or
15 *Mycobacterium avium* subsp. *avium*. Infect. Immun. 69, 1002-1008.

16
17
18

1 **Figure Legends**

2 **Fig. 1.** Venn diagram showing the relationships between genes differentially expressed in the
3 ileum of sheep infected with MAP; comparisons between the three infected groups; A,
4 asymptomatic; M, multibacillary; P, paucibacillary. The figures are the numbers of
5 differentially expressed genes in the different comparisons with a fold change of ≥ 1.5 and
6 $P \leq 0.05$. Figures in parentheses are the number of genes with fold change of ≥ 1.5 but $P > 0.05$.

7

8 **Fig. 2.** Distribution of GO annotations of biological processes terms of genes that were
9 differentially expressed (fold change of ≥ 1.5 and $P \leq 0.05$) in ileum tissue of MAP infected
10 sheep. (a) Comparison of paucibacillary vs asymptomatic. (b) Comparison of multibacillary
11 vs asymptomatic. (c) Comparison of multibacillary vs paucibacillary.

12

13 **Fig. S1.** Gene structures of bovine CXCR4 and IGFBP6 illustrating the positions of the
14 cloning fragments and SNPs.

15

Table 1
Mean fold–changes of significantly differentially expressed genes.

NCBI Official Symbol (Gene)	Accession Number	P v A	M v A	M v P
ANPEP (CD13)	X98240	–2.32	–2.50	
C3 (complement C3)	AF038130	1.53	1.75	
C7 (complement C7)	AF162274		1.5	
C9 (complement C9)	F14494			–1.61
CFB (complement Factor B)	NM_001040526	1.52		
CCL2 (MCP-1)	M84602	1.65*	1.64	
CCL5 (RANTES)	AJ007043	–1.52		
CD3E	Z12969		–1.59	
CD3G	X52994		–1.67	
CD63 (LAMP3)	AJ012589		1.87	1.62
CLIC5	AF109199			–1.54
CXCL10 (IP-10)	AB070717	2.25		–1.72*
CXCR3 (CD183)	X95876		1.54	
CXCR4 (CD185)	AF399642		1.82	1.9
ICAM1 (CD54)	AF110984	1.45	1.9	1.31
IGF2R (CD222)	AF327649		1.74	1.51
IGFBP3	AF327651	–1.61	–1.56	
IGHA1	AF024645	1.96	1.85	
IGHM	X59994	2.31	1.83	
IGJ (J chain)	L20311	1.89		–1.61
IL-18RAP (CD218b)	NM_010553	1.68		
ITGA4 (CD49D)	AB055841	1.52		–1.59
ITGAL (CD11A/LFA-1)	BF654844	1.9	3.58	1.95
ITGAM (CD11B/MAC-1)	AW353414			1.51
ITGB2 (CD18)	M81233		2.03	1.95
MHC-DYA	AJ251357	1.67		
MMP7 (matrilysin)	AB031323			1.84
MMP9 (gelatinase B)	X78324		1.98	1.76
pIGR (secretory component)	L04797	1.56		
SELL (CD62L)	X62882	1.58*	1.85	1.83*
SIRPA (CD172A)	Y11045	1.41	1.65	
SLC11A1 (NRAMP1)	AF005380		1.57	1.63*
TFRC (CD71)	M11507	1.78	2.79	1.81*
TLR2 (CD282)	AF310951		1.52	1.52
TNFRSF1B (CD120b)	NM_001040490		1.62	1.5
TYROBP (DAP12)	AJ419226		1.95	1.68

A – asymptomatic; M – multibacillary; P – paucibacillary.

Bold denotes genes selected for validation by qRT-PCR. Changes less than 1.5 fold (and $P \leq 0.05$) are in *italics* to show trends in the data. *Changes not statistically significant, but >1.5 fold. The blank cells are fold change <1.5 and $P > 0.05$.

Table 2
Relative fold-change of genes analysed by qRT-PCR.

	P v A	M v A	M v P
CD63	1.1 (0.7) ^a	3.2 (0.00)	3.0 (0.00)
CXCR4	2.89 (0.05)	6.13 (0.01)	2.12 (0.05)
IGFBP6	1.86 (0.05)	-1.37 (0.5)	-2.54 (0.02)
IGF2R	2.0 (0.02)	4.2 (0.00)	2.1 (0.00)
ITGB2	2.1 (0.02)	4.3 (0.00)	2.1 (0.00)
MMP9	1.6 (0.02)	14.9 (0.00)	9.3 (0.00)
TLR2	1.6 (0.4)	6.8 (0.00)	4.3 (0.00)
TYROBP	3.1 (0.01)	9.4 (0.00)	3.0 (0.00)

A – asymptomatic; M – multibacillary; P – paucibacillary.

^a mean fold change (P value). Bold, $P \leq 0.05$.

Table 3
Distribution of SNPs

Genotype	Frequency (10 in each group)			χ^2 P value
	Multi	Pauci	Asympto	
CXCR4 C ²⁰⁸⁰ T	2	4	4	0.33
CXCR4 G ²²⁷⁵ A	2	2	4	0.51
CXCR4 G ³³⁶¹ A	4	2	1	0.27
IGFBP-6 C ¹² A	1	1	1	1.00
IGFBP-6 G ²⁷³⁰ A	4	5	5	0.88
IGFBP-6 G ³⁷⁴³ A	2	1	0	0.33

Fig 1

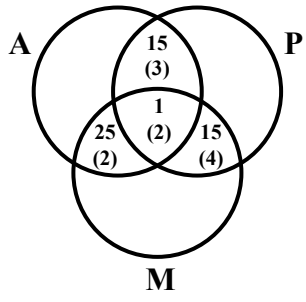
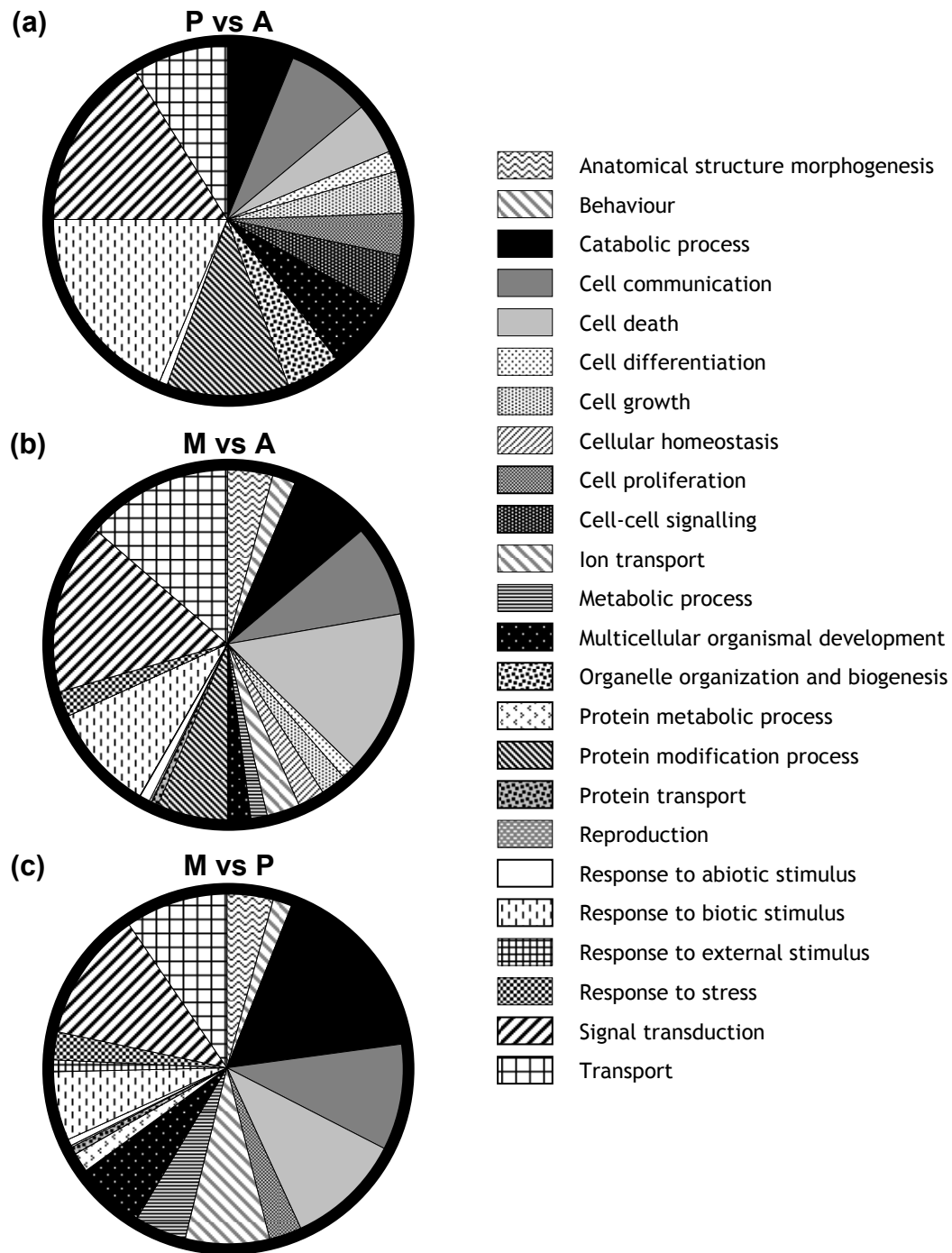
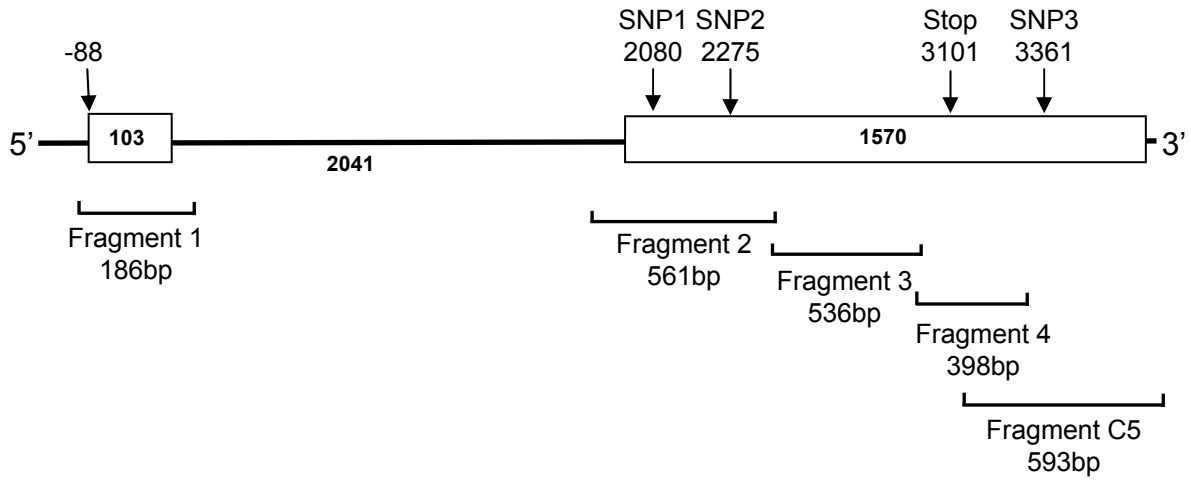


Fig 2



CXCR4



IGFBP-6

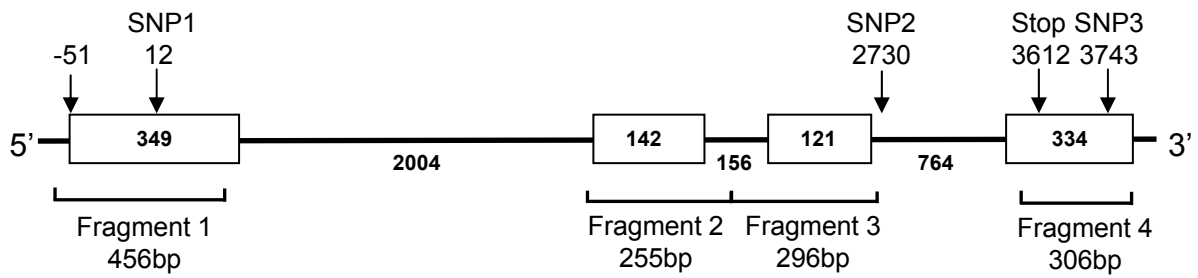


Table S1
Pathology, breed and age of sheep

Pathology	Breed	Age (y)
Pauci ^a	Blackface × Bleu du Maine	4.5
Pauci ^a	Texel	5.5
Pauci ^a	Blackface × Bleu du Maine × Lleyn × Roussin	2.5
Pauci ^a	Lleyn × Roussin	3.5
Pauci ^a	Blackface × Bleu du Maine	6
Pauci ^a	Texel	2.5
Pauci ^b	Blackface × Bleu du Maine	4.5
Pauci ^b	Blackface	2
Pauci	Blackface × Bleu du Maine	4
Pauci	Blackface	3.5
Multi ^a	Blackface	3
Multi ^a	Blackface	4
Multi ^a	Blackface	3
Multi ^a	Blackface	3
Multi ^a	Blackface	5
Multi ^a	Blackface	5
Multi ^b	Blackface	2
Multi ^b	Blackface × Bleu du Maine	4.5
Multi	Blackface × Bleu du Maine	4
Multi	Blackface	4
Asympto ^a	Blackface × Bleu du Maine	7
Asympto ^a	Greyface	4
Asympto ^a	Texel	1
Asympto ^a	Blackface	2.5
Asympto ^a	Greyface	4
Asympto ^a	Greyface	4
Asympto ^b	Blackface × Bleu du Maine	7
Asympto ^b	Blackface × Bleu du Maine	4.5
Asympto	Blackface × Bleu du Maine	3.5
Asympto	Blackface	3

^a animals used in the microarray experiments

^b additional animals used in qRT-PCR experiments

Table S2

Primer sequences for qRT-PCR

Gene Accession no	Primers	Annealing Temp °C	Product size (bp)	Primer conc (nM)
CD63 BC151412.1	F - GGGCTGTGTGGAGAAGATTG R - GATGAGGGGGCTGAAGAGAC	61	178	F: 700 R: 1000
CXCR4 AF399642	F - ACCTCCTGTTTGTCTACG R - AATGTCCACCTCGCTTGC	62	163	F: 500 R: 500
IGFBP-6 NM_001134308	F - AAGGAGAGTAAGCCCCAAGC R - CGGGAAGGAGTGGTAGAGGT	60	95	F: 500 R: 500
ITGB2 NM_001009485	F - CTCACCGACAACCTCCAAACA R - AAAGTGGAACCCATCGTCTG	57	180	F: 400 R: 400
IGF2R AF353513	F - GACGACCTGAAGACCCTGAA R - GCAAATGAAGCGGATGATG	60	150	F: 500 R: 500
MMP9 X78324	F - GAGGGTAAGGTGCTGCTGTTC R - AAGATGTCGTGCGTGCTAATG	62	133	F: 400 R: 500
TLR2 AM183218	F - GCACTTCAACCCTCCCTTTTA R - TCTCCGAAAGCACAAAGATG	57	125	F: 500 R: 500
TYROBP AJ419228.1	F - GACCTGATGCTGACCCTCC R - CTGTCTCCGTGATGTGCTGT	56	112	F: 400 R: 500
SDHA NM_174178	F - ACCTGATGCTTTGTGCTCTGC R - CCTGGATGGGCTTGGAGTAA	62	126	F: 500 R: 500
YWHAZ BC102382	F - TGTAGGAGCCCGTAGGTCATC R - TCTCTCTGTATTCTCGAGCCAT	58	101	F: 500 R: 500

Table S3. Primer sets used to sequence CXCR4 and IGFBP-6

Gene Accession Number	Primer Sequence	Size (bp)	Start position^a
CXCR4 NM_174301	F1 - GAAACTTCAGTGTGTTGGCT	186	-96
	R1 - CCACTCAGAGAGGCGGTTG		
	F2 - ATTCCCTTGCCTGTTTTTCA	561	2035
	R2 - GTACCTCTCATCCACCTCCTTG		
	F3 - GTCTGGCTACCTGCTGTCCT	536	2521
	R3 - GAATGTCCACCTCGCTTGC		
	F4 - CCCATCCTCTATGCCTTCCT	398	2938
	R4 - CGGTCCTACAATGACACACAGC		
	F5 - GAATCTTCAAGTTTTCACTCCAG	593	3076
	R5 - GCCAAAGGAATGCCAATAG		
IGFBP-6 NM_001040495	F1 - CTGGGAAGGGAGGAGGTAGAG	456	-227
	R1 - CGCAGTTGGGAGTGTAGACC		
	F2 - CTTGCGGCTGTAGGAGAGAATC	255	2291
	R2 - CAGGACTGGGATGGGAGACTT		
	F3 - AAGTCTCCCATCCCAGTCCT	296	2525
	R3 - GAAGGGCAGCGTCTCATTTA		
	F4 - GAGGTCCTTGCTGGTGTG	306	3514
R4 - GAACACAGAGACATAAACCAGAG			

^a numbered relative to start codon.